

6-9-2014

Secondary Ion Mass Spectrometry Imaging of Dictyostelium discoideum Aggregation Streams

John Daniel DeBord

Department of Chemistry and Biochemistry, Florida International University *Department of*

Donald F. Smith

FOM Institute AMOLF

Christopher R. Anderton

Pacific Northwest National Laboratory

Ron M.A. Heeren

FOM Institute AMOLF

Ljiljana Paša-Tolić

Pacific Northwest Laboratory

See next page for additional authors

Follow this and additional works at: http://digitalcommons.fiu.edu/chemistry_fac

 Part of the [Life Sciences Commons](#)

Recommended Citation

DeBord JD, Smith DF, Anderton CR, Heeren RMA, Paša-Tolić L, Gomer RH, et al. (2014) Secondary Ion Mass Spectrometry Imaging of Dictyostelium discoideum Aggregation Streams. PLoS ONE 9(6): e99319. doi:10.1371/journal.pone.0099319

This work is brought to you for free and open access by the College of Arts, Sciences & Education at FIU Digital Commons. It has been accepted for inclusion in Department of Chemistry and Biochemistry by an authorized administrator of FIU Digital Commons. For more information, please contact dcc@fiu.edu.

Authors

John Daniel DeBord, Donald F. Smith, Christopher R. Anderton, Ron M.A. Heeren, Ljiljana Paša-Tolić, Richard H. Gomer, and Francisco Fernandez-Lima



Secondary Ion Mass Spectrometry Imaging of *Dictyostelium discoideum* Aggregation Streams

John Daniel DeBord¹, Donald F. Smith², Christopher R. Anderton³, Ron M. A. Heeren², Ljiljana Paša-Tolić³, Richard H. Gomer⁴, Francisco A. Fernandez-Lima^{1*}

1 Department of Chemistry and Biochemistry, Florida International University, Miami, Florida, United States of America, **2** FOM Institute AMOLF, Science Park 104, Amsterdam, The Netherlands, **3** Environmental Molecular Sciences Laboratory, Pacific Northwest National Laboratory, Richland, Washington, United States of America, **4** Department of Biology, Texas A&M University, College Station, Texas, United States of America

Abstract

High resolution imaging mass spectrometry could become a valuable tool for cell and developmental biology, but both, high spatial and mass spectral resolution are needed to enable this. In this report, we employed Bi₃ bombardment time-of-flight (Bi₃ ToF-SIMS) and C₆₀ bombardment Fourier transform ion cyclotron resonance secondary ion mass spectrometry (C₆₀ FTICR-SIMS) to image *Dictyostelium discoideum* aggregation streams. Nearly 300 lipid species were identified from the aggregation streams. High resolution mass spectrometry imaging (FTICR-SIMS) enabled the generation of multiple molecular ion maps at the nominal mass level and provided good coverage for fatty acyls, prenol lipids, and sterol lipids. The comparison of Bi₃ ToF-SIMS and C₆₀ FTICR-SIMS suggested that while the first provides fast, high spatial resolution molecular ion images, the chemical complexity of biological samples warrants the use of high resolution analyzers for accurate ion identification.

Citation: DeBord JD, Smith DF, Anderton CR, Heeren RMA, Paša-Tolić L, et al. (2014) Secondary Ion Mass Spectrometry Imaging of *Dictyostelium discoideum* Aggregation Streams. PLoS ONE 9(6): e99319. doi:10.1371/journal.pone.0099319

Editor: Jeffrey Graham Williams, University of Dundee, United Kingdom

Received: February 14, 2014; **Accepted:** May 13, 2014; **Published:** June 9, 2014

Copyright: © 2014 DeBord et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript. This work was supported by NIGMS grants R00GM106414 to FF-L and GM102280 to RHG. A portion of this research was performed at the W. R. Wiley Environmental Molecular Sciences Laboratory, a national scientific user facility sponsored by the Department of Energy's Office of Biological and Environmental Research (DOE-BER) and located at Pacific Northwest National Laboratory (PNNL). This work is part of the research program of the Foundation for Fundamental Research on Matter (FOM), which is part of The Netherlands Organization for Scientific Research (NWO).

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: fernandf@fiu.edu

Introduction

The interrogation of biological systems with secondary ion mass spectrometry (SIMS) has seen significant growth over the last decade. [1,2,3,4,5,6,7] This relatively newfound application of a surface technique traditionally limited to the study of inorganic and small molecule analytes is largely derived from the advent of larger, cluster primary ion probes (e.g., C₆₀, [8,9,10,11] Ar clusters, [12,13] and Au nanoparticles [14,15,16,17,18,19]) which provide enhanced secondary ion yields of molecular and fragment ions from biological samples. While the use of traditional time of flight (ToF-SIMS) and magnetic sector based methodologies have intrinsic advantages for the *in situ* analysis of surfaces (e.g., speed, sensitivity, dynamic range, depth profiling), the complexity and number of components usually encountered in the analysis of biological systems warrant the coupling of these new sources to high mass accuracy and resolution analytical devices for direct identification of the molecules of interest. [20,21,22,23,24] In particular, this requirement grows out of the need for improved identification certainty for molecular ions generated from biological samples, which are substantially more complex relative to semiconductor and polymer-based applications, where the number of sample components is limited and the analyte of interest is typically predetermined.

Previous mass spectrometry imaging studies have shown the advantages of correlating spatial information with molecular

composition for the study of a variety of biological systems. [25] A common drive has been the search for biological models and better interrogation probes with higher spatial resolution and improved molecular identification. To this end, we used *Dictyostelium discoideum* as a biological model for evaluating the performance of two different mass spectrometry imaging approaches. *D. discoideum* cells are eukaryotic cells that normally live on soil surfaces and eat bacteria. [26,27] An interesting feature of their biological cycle is that when the cells overgrow their food supply and starve, they aggregate together in dendritic streams to form groups of ~20,000 cells. The aggregated cells eventually form a fruiting body consisting of a 1–2 mm tall stalk supporting a mass of spore cells which can then be dispersed by the wind to start new colonies. Because soil surfaces are exposed to rain water, the cells can survive and undergo development in water. This feature makes *D. discoideum* a good model for *in situ* mass spectrometry imaging since it does not require the use of cleaning protocols that can potentially compromise the spatial information (e.g., removal of buffer salts and/or media components). In addition, this cell averages 10 μm in size, which is at the frontier of various surface interrogation techniques (e.g., SIMS, DESI and MALDI). [5,25,28] Although the lipid composition of *D. discoideum* has been studied at different developmental stages using traditional chromatographic techniques and mass spectrometry, [26,29,30,31,32] nothing is known about their distribution during chemotaxis and the aggregation process. In this article, we explore the potential for

SIMS imaging of unknown biological samples by employing traditional TOF-SIMS and accurate mass determination via FTICR-SIMS for direct molecular ion identification of biological components in *D. discoideum* during aggregation.

Experimental Method

Sample Preparation

D. discoideum Ax2 cells were grown in shaking culture at 21°C in Formedium HL-5 as previously described. [33] Mid-log cells ($1\text{--}2 \times 10^6$ cells/ml) were collected by centrifugation at $1,500 \times g$ for 4 minutes, resuspended in PBM (20 mM KH_2PO_4 , 10 μM CaCl_2 , 1 mM MgCl_2 , pH 6.1), and collected by centrifugation. The resuspension and centrifugation were repeated two more times. The cells were resuspended in PBM to 5×10^6 cells/ml, and 10 ml of cells was placed in a 125 ml Erlenmeyer flask and shaken at room temperature for 4 hours. The cells were then diluted 1:6 with PBM, collected by centrifugation, and resuspended in deionized water. The collection and resuspension in deionized water were repeated twice, and the cells were diluted to 9×10^5 cells/ml. 80 μl droplets of the cells were then spotted onto gold-coated silicon chips (Sigma Aldrich). After allowing cells to settle for 30 minutes, 40 μl of the overlaying water was removed and the chips were placed in a humid box at 21°C. 17 hours later, the chips with aggregating cells were gently drained by touching to a kimwipe, and placed cell-side down on a piece of dry ice. This was covered by a piece of aluminum foil, inverted, and placed in a vacuum chamber. After 12 hours, the dry ice had evaporated and the sample was desiccated. The chips with cells were then stored over a CaCl_2 desiccant at room temperature.

Instrumentation

Duplicate *D. discoideum* samples were analyzed in positive ion mode using a ToF SIMS⁵ instrument (ION-TOF, Münster, Germany) and a custom C_{60} FTICR-SIMS. The custom C_{60} FTICR-SIMS instrument (more details in refs [21,23]) utilizes a 40 keV C_{60} primary ion gun (Ionoptika Ltd., Hampshire, England) that is coupled to a SolariX 9.4T FTICR mass spectrometer (Bruker Daltonics Inc, Billerica, MA). The vacuum pumping scheme of the SolariX cart was modified so that the pressure in the source chamber was reduced to 3×10^{-5} mbar instead of the ~ 3 mbar at which it typically operates. The C_{60} FTICR-SIMS images were acquired using 40 keV C_{60}^+ projectiles over a field of view of approximately 4 mm \times 6 mm with a pixel size of 125 μm and a total primary ion dose of 2.78×10^{13} ions/cm². Spectra were acquired using a broadband excitation over the $100 < m/z < 1,500$ range, with 1.0 s transients collected for each pixel. Transients were zero-filled and Sine-Bell apodized prior to fast Fourier transformation. An ion accumulation time of 0.40 s was used to obtain sufficient S/N in the resulting spectra. In the case of the ToF-SIMS analysis, no modifications were made to the instrument. The analysis was performed by rastering the 25 keV Bi_3^+ beam over a 500 μm^2 field of view with a pixel size of 3.9 μm and a total primary ion dose of 8.16×10^{12} ions/cm².

Data Analysis

Spectra and images from the Bi_3 ToF-SIMS analysis were processed using SurfaceLab 6 software (ION-TOF, Münster, Germany). C_{60} FTICR-SIMS images were visualized using FlexImaging software (Bruker Daltonics Inc., Billerica, MA). Peak signals were identified using mMass software [34,35] from the summed spectrum of all pixels within the region of interest. A signal to noise threshold of 10 was used to generate a peak list containing 2,595 peaks. This peak list was then searched against

the LIPID MAPS database (www.lipimaps.org), which contains $\sim 37,000$ entries, using the mMass compound search tool.[34,35] In addition to the typical protonated ions, sodium and potassium adducts as well as dehydration rearrangement products ($-\text{H}_2\text{O}$) were considered. These assignment criteria returned 293 peaks which could be matched to a lipid ion with better than 5 ppm mass accuracy. All reported ion masses were measured from the total spectrum summed over all image pixels.

Results and Discussion

We observed that when *D. discoideum* cells are starved in water, aggregation stream formation begins at about 16 hours, compared to the ~ 8 hours when cells are starved in buffer. However, stream formation was not further delayed when cells were starved on a gold surface instead of the usual glass or plastic surfaces used for most work with this organism. Typical mass spectra of aggregating *D. discoideum* cells from Bi_3 ToF-SIMS and C_{60} FTICR-SIMS are shown in Figure 1. The mass range and ion relative abundances are similar for each instrument and both are characteristic of SIMS analyses of biological targets. That is, the SIMS spectra are dominated by singly charged ions in the $0 < m/z < 500$ range with some larger ion species ($500 < m/z < 1200$) present at lower abundance. A common feature between the spectra is the fact that the most intense peaks correspond to gold cluster and gold cluster hydrocarbon adduct ions derived from the gold-coated silicon wafer substrate (as expected, since this constitutes the majority of the surface area within the analyzed region). The gold cluster species were used to internally calibrate the FTICR-SIMS spectrum summed over all pixels to a mass accuracy below 5 ppm. As a figure of merit, a mass resolving power of $\sim 150,000$ ($m/\Delta m_{50\%}$) was measured at $m/z = 393.9326$ (Au_2^+ peak), where $\Delta m_{50\%}$ is the magnitude mode spectral peak width at half-maximum peak height. The C_{60} FTICR-SIMS spectrum also shows numerous lipid-specific fragments, with the most abundant being the phosphatidylcholine head group ($\text{C}_5\text{H}_{15}\text{NPO}_4^+$) at m/z 184. A total of 293 peaks in the C_{60} FTICR-SIMS spectrum can be attributed to lipid species. When comparing the Bi_3 ToF-SIMS and C_{60} FTICR-SIMS spectra, there are some key differences that become apparent. (1) The radio-frequency ion guides and quadrupole (set to transmit m/z 160 and above) used to transfer ions from the source to the ICR cell induce a low mass cutoff as seen by the significant reduction in ion signal below $m/z = 200$ (relative to the ToF-SIMS spectrum). [36] (2) The greater number of lipid signals detected in the $650 < m/z < 900$ range for C_{60} show that this large cluster projectile is more efficient for generating intact lipid molecular ions than smaller primary ions such as Bi_3 (as previously noted in ref [37]). It is important to note that the ion fluences used were 2.78×10^{13} ions/cm² and 8.16×10^{12} ions/cm² for the C_{60} and Bi_3 analysis, respectively. These values are at or slightly above the static limit, meaning that erosion of the sample is expected. According to the reported sputter yields for these projectiles in organic matrices at the similar fluences and kinetic energies, [38,39,40] the sampled depths are estimated to be approximately 50 nm and 15 nm for the C_{60}^+ and Bi_3^+ analyses, respectively.

Figure 2 shows optical and selected ion images from the Bi_3 ToF-SIMS (Figure 2: B–F) and C_{60} FTICR-SIMS (Figure 2: H–L) spectra. The optical microscopy images (Figure 2: A, G) clearly show the cellular aggregation streams which form branched structures $< 200 \mu\text{m}$ in width and a few millimeters in length. The ToF-SIMS total secondary ion image (Figure 2B) shows higher intensity for ions originating from the aggregation streams and lower overall intensity from the gold substrate. $\text{C}_5\text{H}_{13}\text{NPO}_3^+$ (m/z

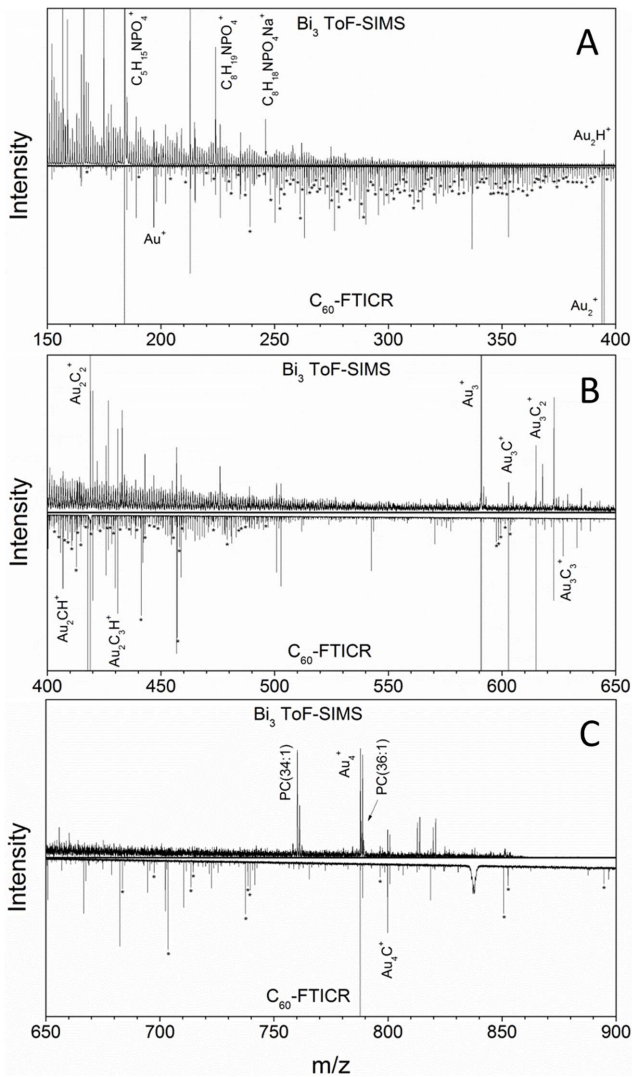


Figure 1. Comparison of Bi_3 ToF-SIMS and C_{60} FTICR-SIMS spectra. Bi_3 ToF-SIMS (top of each panel) and C_{60} FTICR-SIMS (inverted in each panel) spectra of aggregating *D. discoideum* cells. Lipid fragments, gold clusters, and gold cluster adduct peaks are labeled with their corresponding molecular formulas while lipid species identified from the LIPID MAPS database are denoted with asterisks (*). doi:10.1371/journal.pone.0099319.g001

= 166.1), $\text{C}_5\text{H}_{15}\text{NPO}_4^+$ ($m/z = 184.1$), and $m/z = 760.6$ give spatial distributions corresponding to the aggregation streams. The $\text{C}_5\text{H}_{13}\text{NPO}_3^+$ and $\text{C}_5\text{H}_{15}\text{NPO}_4^+$ species are head group fragment ions from glycerophosphatidylcholines, which make up ~25% of all lipids present in *D. discoideum*. [31] The signal at $m/z = 760.6$ appears to be a lipid molecular ion due to its co-localization with the aggregations streams, the observed isotopic pattern which contains significant ^{13}C contributions, and the presence of another peak at $m/z = 788.6$ corresponding to the same molecule with a fatty acyl chain two carbons longer. [41,42] However, due to the limited mass accuracy afforded by ToF analysis and their absence from the C_{60} FTICR-SIMS spectrum, the precise identities of these supposed lipids was not determined. Viewing the sample as a binary system containing signals from the cellular aggregations and from the substrate, we are also able to show that the Au_3^+ ion image represents only the substrate as this signal is not observed from the aggregations streams.

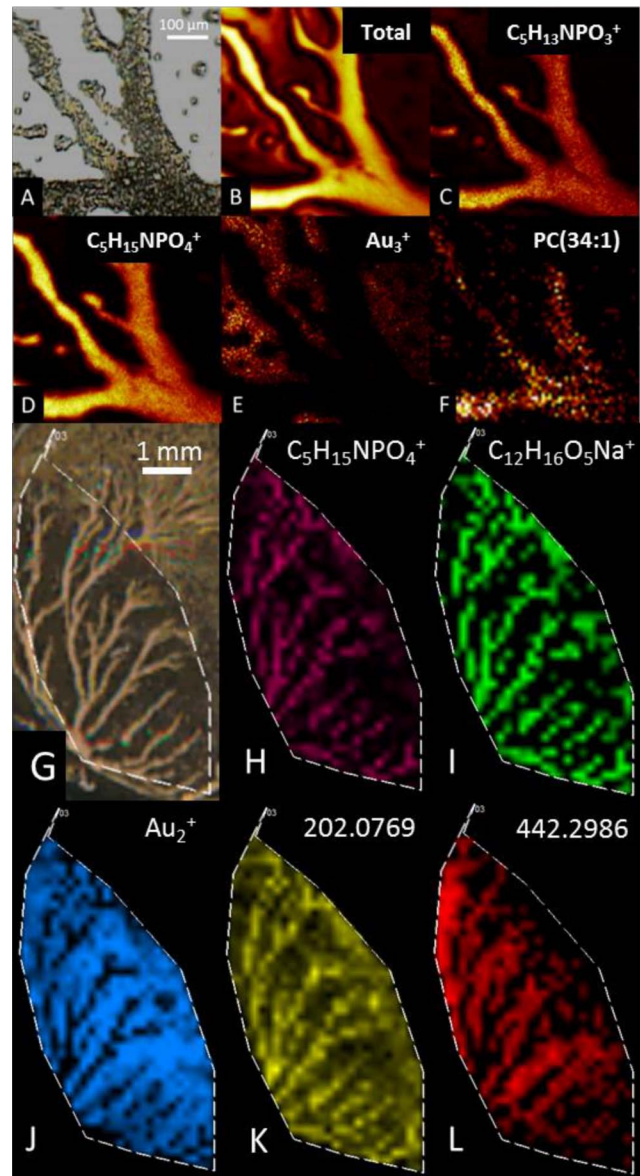


Figure 2. Optical and secondary ion images of *D. discoideum* aggregation streams. (A) Optical and (B–F) ion images of aggregation streams generated by 25 keV Bi_3 TOF-SIMS analysis. (G) Optical and (H–L) ion images of aggregation streams generated by 40 keV C_{60} FTICR-SIMS analysis. doi:10.1371/journal.pone.0099319.g002

Analogously, molecular ion images can also be obtained from the C_{60} FTICR-SIMS spectra. Two of the mass spectral features (Figure 2: H, I) which display spatial distributions corresponding to the aggregation streams are $\text{C}_5\text{H}_{15}\text{NPO}_4^+$ ($m/z = 184.0737$, $\delta = 2.1$ ppm) and $\text{C}_{12}\text{H}_{16}\text{O}_5\text{Na}^+$ ($m/z = 263.0889$, $\delta = -0.2$ ppm). As mentioned above, $\text{C}_5\text{H}_{15}\text{NPO}_4^+$ corresponds to the phosphatidylcholine head group, while according to the LIPID MAPS database, the $\text{C}_{12}\text{H}_{16}\text{O}_5\text{Na}^+$ species corresponds to the heterocyclic fatty acyl 3-carboxy-4-methyl-propyl-2-furanpropionic acid (LIPID MAPS ID: LMFA01150004), which has previously been detected from human uremic serum as a sodiated ion using SIMS. [43] Ion images for two unidentified peaks from the FTICR spectrum are shown in panels K and L. The image of $m/z 202.0769$ shows a distribution consistent with the aggregation

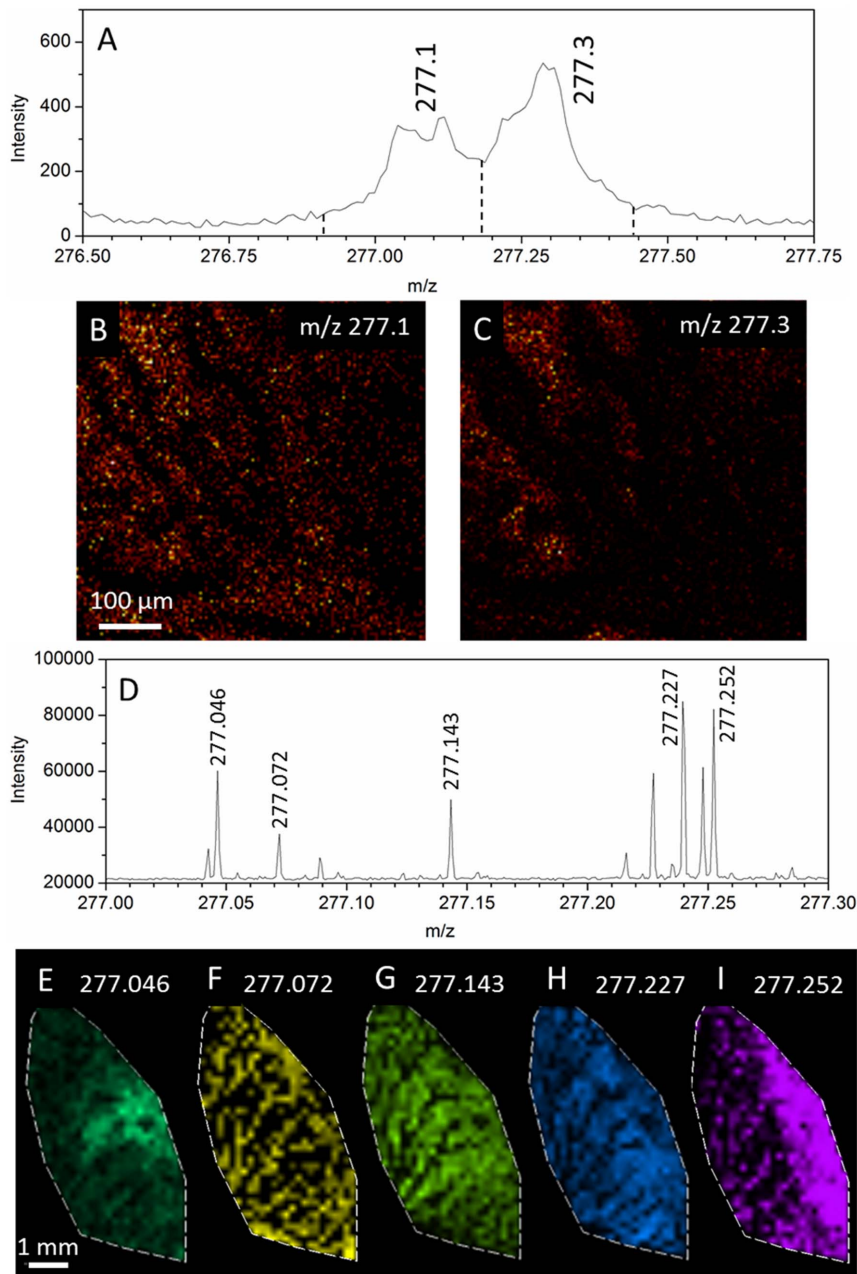


Figure 3. Secondary ion images from within the $m/z = 277$ nominal mass. (A) Bi_3 ToF-SIMS and (D) C_{60} FTICR-SIMS spectra excerpts showing multiple peaks within the 277 nominal mass. (B,C) Bi_3 ToF-SIMS ion images obtained from the first “peak” and second “peak” within the 277 nominal mass. (E–I) C_{60} FTICR-SIMS ion images generated for the corresponding peaks in D with a m/z bin size of ± 0.001 . doi:10.1371/journal.pone.0099319.g003

streams with lower level concentrations between the aggregation streams. The m/z 442.2986 ion is located on the surface in proximity to, but not within the aggregation streams. Such an arrangement may mean this ion corresponds to a metabolite which is secreted from the *D. discoideum* cells. The m/z 202.0769 and 442.2986 ions did not return lipid matches within the 5 ppm mass accuracy threshold, suggesting these lipids are not contained in the database, these compounds are not lipids, or the mass errors for these peaks fall outside the applied threshold range. As such, identities for these ions can not be determined from this analysis. As in the ToF-SIMS analysis, a Au-related ion, Au_2^+ (m/z

393.9326, $\delta = -0.1$ ppm), can be used to visualize the substrate and not the aggregation streams.

The molecular ion images shown in Figure 2 demonstrate that ions throughout the mass range can be used to display meaningful spatial distributions. Moreover, the mass resolving power of the FTICR-SIMS instrument is most apparent when the true complexity of the sample is revealed. The excerpted mass spectrum (from the sum of all spectra) shown in Figure 3D shows that within the spectrum, there can be upwards of 10 ions within a given nominal mass, and that each of these ions may arise from different regions within the sample. Assuming a composition of carbon, hydrogen, nitrogen, oxygen, and phosphorus and a 5 ppm

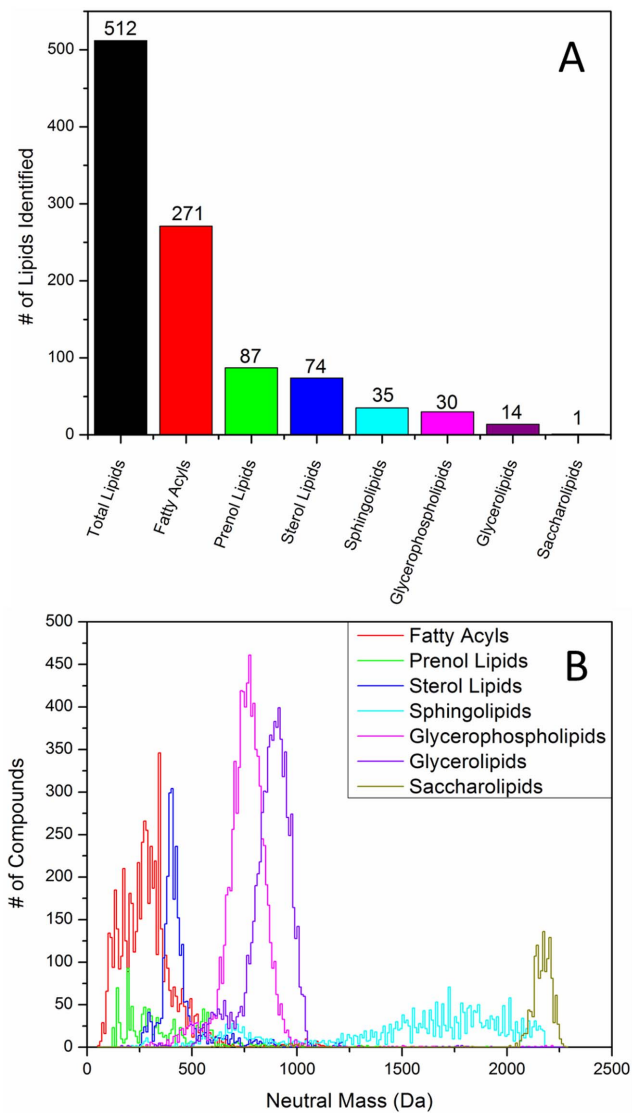


Figure 4. Overlap between the detected lipid classes and the LIPID MAPS database. (A) The number of peaks identified from the C_{60} -FTICR-SIMS spectrum by lipid class. (B) Mass distributions for compounds from the LIPID MAPS database organized by lipid class. Each data point represents the number of lipids for a given class binned every 10 mass units.

doi:10.1371/journal.pone.0099319.g004

threshold, we can suggest molecular formulas for the peaks at $m/z = 277.046$, 277.072 , 277.143 , 277.227 , and 277.252 m/z to be $C_{12}H_9N_2O_6^+$ or $C_{10}H_{14}O_7P^+$, $C_{14}H_{13}O_6^+$, $C_{16}H_{21}O_4^+$, $C_{17}H_{29}N_2O^+$, and $C_{19}H_{33}O^+$ respectively. From the ion images it is apparent that two of the ions (277.072 and 277.252) originate from the aggregation streams. As expected, these two ions are the only two which match entries from the LIPID MAPS database. The $m/z = 277.072$ ion can be identified as Thysanone (LIPID MAPS ID: LMPK13030001, $[C_{14}H_{12}O_6+H]^+$, $\delta = -4.9$ ppm). [44] The peak at 277.252 has the formula $[C_{19}H_{32}O+H]^+$ ($\delta = -0.2$ ppm) and has 13 possible lipid matches with the same stoichiometry. This list of matches includes a sterol lipid, 4 prenol lipids, and 8 sphingolipids. Analysis with ToF-SIMS (Figure 3A) reveals at least two unresolved peaks within the m/z 277 nominal mass. The selected ion images generated by integrating the left and right halves of the peak cluster (3B,3C) show some differences in

spatial distribution, but the unique distributions of the summed peaks are lost due to insufficient mass resolving power. Further attempts to segment the peaks resulted in insufficient counts per window to generate ion images (see Figure S1).

The search for lipid IDs from the high resolution FTICR-SIMS spectrum against the LIPID MAPS database resulted in 293 hits throughout the spectrum within 5 ppm mass measurement accuracy. Depending on the uniqueness of each detected m/z , peaks can be assigned to a single lipid, any of multiple isomers within a given class, or to any of multiple isomers from multiple lipid classes. A summary plot showing the 512 lipid class assignments for the 293 peaks is provided in Figure 4A, with the detailed list of peak assignments included as Table S1. Figure 4B shows the number of compounds from each lipid class in the LIPID MAPS database binned every 10 Da. The three most commonly detected classes of lipids, the fatty acyls, prenol lipids, and sterol lipids, feature mass distributions which reside almost entirely within the 200–500 Da range. This happens to be the range over which most of the C_{60} FTICR-SIMS signal is observed. As a general trend, as the mass of the compound increases, the probability of ion formation/survival decreases in SIMS analyses. Only one saccharolipid (LMSL05000001, $C_{18}H_{32}O_8$) was detected due to the fact that nearly all saccharolipids in the database reside at $>2,000$ Da. Most of the detected species, especially the nonpolar and electronegative compounds (like fatty acyls), were detected as sodiated or potassiated ions, with most undergoing dehydration reactions in order to generate positive ions.

Previous reports of the *D. discoideum* lipid profile are almost exclusively limited thin-layer chromatographic measurements of the types and relative abundances of the general lipid classes without regard to the specific lipids present. As an example, Paquet et al. recently reported that neutral, phosphoethanolamine, and phosphocholine lipids constitute over 80% of a total lipid extract from *D. discoideum*. However, variations in ionization probability between lipid classes and a mass dependent detection probability preclude quantitative comparisons of this type from mass spectrometric data. There have also been many reports of the fatty acid profile of *D. discoideum* obtained from hydrolyzed lipid extracts, but these fatty acyls have not been linked back to their parent lipid class. In order to obtain a more detailed lipid profile, the fatty acyls should be detected along with their corresponding head groups. This could be done either by using a solvent pre-fractionation method to isolate the various lipid class prior to hydrolysis and subsequent GC analysis [45] or by analysis of the original intact molecular ions [24] as was done here. This has been done for the most abundant sphingolipids from *D. discoideum* using liquid chromatography mass spectrometry; [32] however, this analysis was performed in negative ion mode while our MS analyses were acquired in positive ion mode. The author did propose identities for the four most abundant lipids observed in positive ion mode to be PC(36:4), PC(34:4), PC(32:2), and PS(32:1), but these ions were not observed in the C_{60} FTICR-SIMS spectrum.

Despite the fact that lipid profiling using this approach is biased by the mass range and ionization probability of the desorbed molecules, it does offer a rapid tool for molecular differentiation and cell state classification. A current limitation of this approach (e.g., compared to LC-MS lipid profiling) lies in the inability to differentiate isobaric species. The identity of structural isomers is often important in lipid analysis and efforts have been made to incorporate MS/MS capabilities into SIMS analysis. [46,47]. The current FTICR-SIMS instrument is also capable of MS/MS measurements, [21] though none were performed during the course of this study. Another limitation of the current prototype

lies in the sub-optimal focusing of the C_{60}^+ primary ion beam which has a diameter of $\sim 75 \mu\text{m}$ and the lack of ion raster optics which means mechanical stage movement must be used to generate ion images rather than the more precise method of beam rastering. Other groups have shown that C_{60} beams can be focused down to 200 nm and rastered to create images with sub-micron spatial resolution. [48] Such improvements would be necessary for the current instrument to resolve smaller surface features such as lipid distributions within *Dictyostelium* aggregation streams or individual *Dictyostelium* cells.

Conclusions

Bi_3 ToF-SIMS and C_{60} FTICR-MS offer complementary information, where the first analysis provides short analysis times and high spatial resolution while the second demonstrates the need for higher mass resolving power when interrogating biological samples. In particular, the use of high mass resolving power in SIMS (e.g., FTICR-SIMS) was shown to be effective for the analysis of a variety of chemical classes with molecular ion masses $< 1,000 \text{ Da}$ (e.g., fatty acyls, prenol lipids, and sterol lipids). Further incorporation of high resolution mass analyzers with high spatial resolution surface probes will permit a better identification of molecular components in biological matrices, a necessary step in the progression towards single cell mass spectrometry imaging.

Supporting Information

Figure S1 Bi_3 ToF-SIMS secondary ion images from within the $m/z = 277$ nominal mass. (A–D) Bi_3 ToF-SIMS

References

- Gormanns P, Reckow S, Poczatek JC, Turck CW, Lechene C (2012) Segmentation of multi-isotope imaging mass spectrometry data for semi-automatic detection of regions of interest. *Plos One* 7: e30576.
- Touboul D, Brunelle A, Lapr v te O (2011) Mass spectrometry imaging: Towards a lipid microscope? *Biochimie* 93: 113–119.
- Fletcher JS, Lockyer NP, Vickerman JC (2011) Developments in Molecular SIMS Depth Profiling and 3D Imaging of Biological Systems using Polyatomic Primary Ions. *Mass Spectrometry Reviews* 30: 142–174.
- Benabdellah F, Seyer A, Quinton L, Touboul D, Brunelle A, et al. (2010) Mass spectrometry imaging of rat brain sections: nanomolar sensitivity with MALDI versus nanometer resolution by TOF-SIMS. *Analytical & Bioanalytical Chemistry* 396: 151–162.
- Chughtai K, Heeren RMA (2010) Mass Spectrometric Imaging for Biomedical Tissue Analysis. *Chemical Reviews* 110: 3237–3277.
- Frisz JF, Lou K, Klitzing HA, Hanafin WP, Lizunov V, et al. (2013) Direct chemical evidence for sphingolipid domains in the plasma membranes of fibroblasts. *Proceedings of the National Academy of Sciences* 110: E613–E622.
- Kraft ML, Weber PK, Longo ML, Hutcheon ID, Boxer SG (2006) Phase Separation of Lipid Membranes Analyzed with High-Resolution Secondary Ion Mass Spectrometry. *Science* 313: 1948–1951.
- van Stipdonk MJ, Harris RD, Schweikert EA (1996) A Comparison of Desorption Yields from C_{60} to Atomic and Polyatomic Projectiles at keV Energies. *Rapid Communications in Mass Spectrometry* 10: 1987–1991.
- Cheng J, Winograd N (2005) Depth Profiling of Peptide Films with TOF-SIMS and a C_{60} Probe. *Analytical Chemistry* 77: 3651–3659.
- Fletcher JS, Conlan XA, Jones EA, Biddulph G, Lockyer NP, et al. (2006) TOF-SIMS Analysis Using C_{60} . Effect of Impact Energy on Yield and Damage. *Analytical Chemistry* 78: 1827–1831.
- Fletcher JS, Lockyer NP, Vaidyanathan S, Vickerman JC (2007) TOF-SIMS 3D Biomolecular Imaging of *Xenopus laevis* Oocytes Using Buckminsterfullerene (C_{60}) Primary Ions. *Analytical Chemistry* 79: 2199–2206.
- Rabbani S, Barber AM, Fletcher JS, Lockyer NP, Vickerman JC (2011) TOF-SIMS with argon gas cluster ion beams: a comparison with C_{60}^+ . *Analytical Chemistry* 83: 3793–3800.
- Bich C, Havelund R, Moellers R, Touboul D, Kollmer F, et al. (2013) Argon cluster ion source evaluation on lipid standards and rat brain tissue samples. *Analytical Chemistry* 85: 7745–7752.
- Fernandez-Lima FA, DeBord JD, Schweikert EA, Della-Negra S, Kellersberger KA, et al. (2013) Surface characterization of biological nanodomains using NP-ToF-SIMS. *Surface and Interface Analysis* 45: 294–297.
- Fernandez-Lima FA, Post J, DeBord JD, Eller MJ, Verkhoturov SV, et al. (2011) Analysis of Native Biological Surfaces Using a 100 kV Massive Gold Cluster Source. *Analytical Chemistry* 83: 8448–8453.
- Della-Negra S, Depauw J, Guillemier C, Schweikert EA (2011) Massive clusters: Secondary emission from keV to qMeV. New emission processes? *New SIMS probe? Surface and Interface Analysis* 43: 62–65.
- Novikov A, Caroff M, Della-Negra S, Depauw J, Fallavier M, et al. (2005) The Aun cluster probe in secondary ion mass spectrometry: Influence of the projectile size and energy on the desorption/ionization rate from biomolecular solids. *Rapid Communications in Mass Spectrometry* 19: 1851–1857.
- Bouneau S, Della-Negra S, Depauw J, Jacquet D, Le Beyec Y, et al. (2004) Heavy gold cluster beams production and identification. *Nuclear Instruments and Methods in Physics Research Section B: Beam Interactions with Materials and Atoms* 225: 579–589.
- Brunelle A, Della-Negra S, Depauw J, Jacquet D, Le Beyec Y, et al. (2001) Enhanced secondary-ion emission under gold-cluster bombardment with energies from keV to MeV per atom. *Physical Review A* 63: 022902.
- Gilmore IS (2013) SIMS of organics—Advances in 2D and 3D imaging and future outlook. *Journal of Vacuum Science & Technology A* 31: 050819.
- Smith DF, Robinson EW, Tolmachev AV, Heeren RMA, Pasa-Tolic L (2011) C_{60} Secondary Ion Fourier Transform Ion Cyclotron Resonance Mass Spectrometry. *Analytical Chemistry* 83: 9552–9556.
- Green FM, Gilmore IS, Seah MP (2011) Mass Spectrometry and Informatics: Distribution of Molecules in the PubChem Database and General Requirements for Mass Accuracy in Surface Analysis. *Analytical Chemistry* 83: 3239–3243.
- Smith D, Kiss A, Leach F III, Robinson E, Pasa-Tolic L, et al. (2013) High mass accuracy and high mass resolving power FT-ICR secondary ion mass spectrometry for biological tissue imaging. *Analytical and Bioanalytical Chemistry* 405: 6069–6076.
- Fhaner CJ, Liu S, Ji H, Simpson RJ, Reid GE (2012) Comprehensive Lipidome Profiling of Isogenic Primary and Metastatic Colon Adenocarcinoma Cell Lines. *Analytical Chemistry* 84: 8917–8926.
- Ellis S, Bruinen A, Heeren RA (2013) A critical evaluation of the current state-of-the-art in quantitative imaging mass spectrometry. *Analytical and Bioanalytical Chemistry*: 1–15.
- Paquet VE, Lessire R, Domergue F, Fouillen L, Filion G, et al. (2013) Lipid Composition of Multilamellar Bodies Secreted by *Dictyostelium discoideum* Reveals Their Amoebal Origin. *Eukaryotic Cell* 12: 1326–1334.
- Jang W, Gomer RH (2011) Initial Cell Type Choice in *Dictyostelium*. *Eukaryotic Cell* 10: 150–155.
- Wu C, Dill AL, Eberlin LS, Cooks RG, Ifa DR (2013) Mass spectrometry imaging under ambient conditions. *Mass Spectrometry Reviews* 32: 218–243.

29. Long BH, Coe EL (1974) Changes in Neutral Lipid Constituents during Differentiation of the Cellular Slime Mold, *Dictyostelium discoideum*. *Journal of Biological Chemistry* 249: 521–529.
30. Birch GL (2011) Lipidomic profiling of *Dictyostelium Discoideum*. Purdue University.
31. Weeks G, Herring FG (1980) The lipid composition and membrane fluidity of *Dictyostelium discoideum* plasma membranes at various stages during differentiation. *Journal of Lipid Research* 21: 681–686.
32. Birch GL (2011) Lipidomic Profiling of *Dictyostelium Discoideum*. Purdue University.
33. Brock DA, Gomer RH (2005) A secreted factor represses cell proliferation in *Dictyostelium*. *Development* 132: 4553–4562.
34. Strohal M, Kavan D, Novák P, Volný M, Havlíček VR (2010) mMass 3: A Cross-Platform Software Environment for Precise Analysis of Mass Spectrometric Data. *Analytical Chemistry* 82: 4648–4651.
35. Strohal M, Hassman M, Kořata B, Kodiček M (2008) mMass data miner: an open source alternative for mass spectrometric data analysis. *Rapid Communications in Mass Spectrometry* 22: 905–908.
36. Beu S, Hendrickson C, Marshall A (2011) Excitation of Radial Ion Motion in an rf-Only Multipole Ion Guide Immersed in a Strong Magnetic Field Gradient. *Journal of the American Society for Mass Spectrometry* 22: 591–601.
37. Ostrowski SG, Szakal C, Kozole J, Roddy TP, Xu J, et al. (2005) Secondary Ion MS Imaging of Lipids in Picoliter Vials with a Buckminsterfullerene Ion Source. *Analytical Chemistry* 77: 6190–6196.
38. Brison J, Muramoto S, Castner DG (2010) ToF-SIMS Depth Profiling of Organic Films: A Comparison between Single-Beam and Dual-Beam Analysis. *The Journal of Physical Chemistry C* 114: 5565–5573.
39. Muramoto S, Brison J, Castner DG (2011) Exploring the Surface Sensitivity of TOF-Secondary Ion Mass Spectrometry by Measuring the Implantation and Sampling Depths of Bin and C60 Ions in Organic Films. *Analytical Chemistry* 84: 365–372.
40. Delcorte A, Leblanc C, Poleunis C, Hamraoui K (2013) Computer Simulations of the Sputtering of Metallic, Organic, and Metal–Organic Surfaces with Bin and C60 Projectiles. *The Journal of Physical Chemistry C* 117: 2740–2752.
41. Petković M, Schiller J, Müller M, Benard S, Reichl S, et al. (2001) Detection of Individual Phospholipids in Lipid Mixtures by Matrix-Assisted Laser Desorption/Ionization Time-of-Flight Mass Spectrometry: Phosphatidylcholine Prevents the Detection of Further Species. *Analytical Biochemistry* 289: 202–216.
42. Hase A (1981) Fatty acid composition and sterol content of *Dictyostelium discoideum* cells at various stages of development. *Journal of the Faculty of Science, Hokkaido University* 12: 183–194.
43. Takeda N, Niwa T, Tatematsu A, Suzuki M (1987) Identification and quantification of a protein-bound ligand in uremic serum. *Clinical chemistry* 33: 682–685.
44. Singh SB, Cordingley MG, Ball RG, Smith JL, Dombrowski AW, et al. (1991) Structure of stereochemistry of thysanone: a novel human rhinovirus 3C-protease inhibitor from *Thysanophora penicilloides*. *Tetrahedron Letters* 32: 5279–5282.
45. Yoshioka S, Nakashima S, Okano Y, Hasegawa H, Ichiyama A, et al. (1985) Phospholipid (diacyl, alkylacyl, alkenylacyl) and fatty acyl chain composition in murine mastocytoma cells. *Journal of Lipid Research* 26: 1134–1141.
46. Ferreri C, Chatgililoglu C (2005) Geometrical trans Lipid Isomers: A New Target for Lipidomics. *ChemBioChem* 6: 1722–1734.
47. Piehowski PD, Carado AJ, Kurczy ME, Ostrowski SG, Heien ML, et al. (2008) MS/MS Methodology To Improve Subcellular Mapping of Cholesterol Using TOF-SIMS. *Analytical Chemistry* 80: 8662–8667.
48. Fletcher JS, Rabbani S, Henderson A, Blenkinsopp P, Thompson SP, et al. (2008) A New Dynamic in Mass Spectral Imaging of Single Biological Cells. *Analytical Chemistry* 80: 9058–9064.