FUNCTIONAL RESPONSE OF HERBIVORES IN 
FOOD-CONCENTRATED PATCHES: 
TESTS OF A MECHANISTIC MODEL

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Abstract. Type II functional responses are frequently observed in herbivores feeding in patches where plants are concentrated in space. We tested a mechanistic model of regulation of intake rate of herbivores foraging in food-concentrated patches (Laca and Demment 1992, Spalinger and Hobbs 1992) that accounts for asymptotic, Type II responses. The model is based on the hypothesis that competition between cropping and chewing regulates instantaneous intake rate in response to changes in the size of bites obtained by the forager. We tested this hypothesis and examined the ability of our model to account for observations of intake rate of 12 species of mammalian herbivores ranging in body mass over 4 orders of magnitude.

We measured short-term intake rates of mammalian herbivores feeding in hand-assembled patches of plants. We varied bite size by changing plant height and density in patches offered to herbivores, and observed dry matter intake rates in response to this variation. Averaged across species, our model accounted for 77% of the variance in food intake rate (P < .001 for all species). Predictions of maximum intake rate closely resembled observations of processing capacity, demonstrating that processing rather than cropping sets an upper limit on short-term intake. Tests of model mechanisms provided strong support for the hypothesis that competition between cropping and chewing is responsible for the Type II functional response seen in herbivores feeding in food-concentrated patches. The model was able to consistently predict intake rates observed in 16 previous studies. These results indicate that plant characteristics regulating bite size (e.g., leaf size and geometry, spinescence) frequently control instantaneous rates of food intake by mammalian herbivores.

Key words: bite size; browsing; disc equation; foraging; functional response; grazing; handling time; herbivory; intake rate; mammal; patch; plant–animal interaction.

INTRODUCTION

Solomon (1949) and Holling (1959, 1965) distin- 
guished between the numerical response of animals and their functional response. In one form or another, equations representing these responses play a central role in most contemporary models of predator–prey and plant–herbivore interactions (Hassell 1978, Caughley 1982, Edelstein-Keshet 1986, Elsen et al. 1988, Gren-
functional response that are faithful to the unique features of foraging by herbivores.

Unlike most predators, herbivores often forage in patches where foods are highly concentrated in space and are readily apparent to the forager. Grass swards and stands of leafy shrubs offer examples of such food patches. Under these circumstances, the density of "prey" approaches infinity, i.e., one prey item immediately adjoins another. When this is the case, traditional models of functional response predict that intake is simply the reciprocal of handling time, which is parsimoniously assumed to be a constant function of prey size. For example, the disc equation (Holling 1959) predicts the number of prey consumed per unit time as a function of prey density. Prey consumption varies with prey handling time and predator searching efficiency,

\[ N = \frac{ADT}{1 + AHD}, \]

where \( N \) = the number of prey captured, \( A \) = the predator's searching efficiency (in square metres per minute), \( D \) = prey density (number per square metre), \( T \) = duration of foraging (in minutes), and \( H \) = handling time of prey item (minutes per prey). If we define \( N \) as the number of bites of plant tissue (i.e., bites = prey) taken by an herbivore during a given time interval \( T \), we can obtain a form that specifies the herbivore's instantaneous rate of intake of plant mass \( (I, \) in grams per minute) by dividing the disc equation by bite size \( (S, \) in grams per bite) and dividing by foraging time \( (T) \):

\[ I = \frac{AD}{1 + AHD} \cdot S. \]

When prey are infinity dense, we have

\[ \lim_{D \to \infty} \frac{AD}{1 + AHD} \cdot S = S/H = I, \]

which predicts that the intake rate of herbivores feeding in food-saturated patches should increase as a linear function of bite size (i.e., a Type I functional response). This prediction is contradicted by empirical evidence showing that intake rate of herbivores increases asymptotically as bite size increases (i.e., a Type II functional response; Allden and Whittaker 1970, Wickstrom et al. 1984, Hudson and Watkins 1986, Hudson and Frank 1987, Spalinger et al. 1988). A similar model has been developed independently by Laca and Demment (1992), and Spalinger and Hobbs (1992). Explaining this Type II behavior requires a model that allows handling time \( (H) \) to change dynamically as bite size changes.

The need for a dynamic representation of handling time motivated us to develop a new model of functional response of herbivores foraging in food-concentrated patches (Spalinger and Hobbs 1992). We began by defining a bite as a mass of plant tissue that an herbivore can remove from the plant with a single jaw movement. The size of the bite cropped \( (S) \) is determined by the interaction between the geometry of the animal's mouth (Illius and Gordon 1987, Janis and Ehrhardt 1988) and the geometric arrangement of plant tissue in space (Stobbs 1973, Chacon and Stobbs 1976, Burlison et al. 1991, Penning et al. 1991, Laca et al. 1992). If cropping rate \( (B, \) in bites per minute) were a constant function of bite size, then intake rate would be directly proportional to bite size, as suggested by the disc equation. Alternatively, we hypothesized (Spalinger and Hobbs 1992) that cropping rate must decline with increasing bite size because larger bites require an herbivore to invest more effort in masticating a bite to prepare it for swallowing, and because these chewing movements cannot occur simultaneously with cropping movements (Ardran et al. 1958, Greaves 1978, Fortelius 1985). Based on these ideas, we have shown (Spalinger and Hobbs 1992) that cropping rate, (and thus handling time), can be predicted as

\[ \frac{1}{H} = B = \frac{R_{\text{max}}}{R_{\text{max}}h + S}, \]

where \( R_{\text{max}} \) is the maximum rate of processing of plant tissue in the mouth (in grams per minute) that would occur in the absence of cropping, and \( h \) is the average time (in minutes per bite) required to crop a single bite in the absence of chewing. It follows (Spalinger and Hobbs 1992) that intake rate of herbivores feeding on spatially concentrated foods can be represented as an asymptotic function of bite size:

\[ I = \frac{R_{\text{max}}S}{R_{\text{max}}h + S}. \]

Thus, Eq. 5 represents the hypothesis that competition between cropping and chewing is responsible for the Type II functional response frequently seen in herbivores feeding in food-concentrated patches (Allden and Whittaker 1970, Wickstrom et al. 1984, Hudson and Watkins 1986, Hudson and Frank 1987, Spalinger et al. 1988). A similar model has been developed independently by Laca and Demment (1992), and Spalinger and Hobbs (1992) derived explicit conditions defining when these models should apply.

Here, we report experimental tests of our model of functional response (Eq. 5) for mammalian herbivores differing in body size and feeding style. We offer tests of the model's mechanism and discuss the significance of that mechanism for understanding foraging behavior of herbivores.

**Materials and Methods**

**Experimental manipulation of bite size**

We observed changes in food intake rate \( (I, \) Eq. 5) that occurred in response to changes in bite size \( (S, \) Eq. 5) of 12 species of herbivores spanning 4 orders of magnitude in body mass (Table 1). The animals in our experiments represented a variety of feeding styles, dentition patterns, and digestive adaptations. Differences in bite size were imposed by varying the size of
Table 1. Species used in tests of the functional response model.

<table>
<thead>
<tr>
<th>Species</th>
<th>Mass (kg)</th>
<th>(n)*</th>
<th>Origin/rearing†</th>
<th>Molar dentition style</th>
<th>Feeding style</th>
<th>Site‡</th>
<th>Distance§</th>
<th>No. trials</th>
</tr>
</thead>
<tbody>
<tr>
<td>Collared lemming</td>
<td>0.05</td>
<td>(13)</td>
<td>C/M</td>
<td>hypsodont</td>
<td>grazer</td>
<td>CO</td>
<td>1</td>
<td>57</td>
</tr>
<tr>
<td>(Dicrostonyx groenlandicus)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Black-tailed prairie dog</td>
<td>0.75</td>
<td>(3/2)</td>
<td>W/M</td>
<td>hypsodont</td>
<td>mixed</td>
<td>CO</td>
<td>4.5</td>
<td>108</td>
</tr>
<tr>
<td>(Cynomys ludovicianus)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Domestic rabbit (Oryctolagus cuniculus)</td>
<td>2.9</td>
<td>(2/3)</td>
<td>C/M</td>
<td>hypsodont</td>
<td>grazer</td>
<td>TX</td>
<td>9</td>
<td>125</td>
</tr>
<tr>
<td>Collared peccary (Tayassu tajacu)</td>
<td>27.2</td>
<td>(3/0)</td>
<td>C/M</td>
<td>bunodont</td>
<td>concentrate†</td>
<td>TX</td>
<td>9</td>
<td>124</td>
</tr>
<tr>
<td>White-tailed deer (fawn)</td>
<td>25.5</td>
<td>(2/2)</td>
<td>C/B</td>
<td>selenodont</td>
<td>mixed</td>
<td>TX</td>
<td>18</td>
<td>90</td>
</tr>
<tr>
<td>(Odocoileus virginianus)</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>White-tailed deer (adult)</td>
<td>45.0</td>
<td>(1/3)</td>
<td>C/B</td>
<td>selenodont</td>
<td>mixed</td>
<td>TX</td>
<td>18</td>
<td>67</td>
</tr>
<tr>
<td>Axis deer (Axis axis)</td>
<td>53.0</td>
<td>(0/3)</td>
<td>W/B</td>
<td>selenodont</td>
<td>grazer</td>
<td>TX</td>
<td>18</td>
<td>62</td>
</tr>
<tr>
<td>Caribou (Rangifer tarandus)</td>
<td>104</td>
<td>(2/0)</td>
<td>W/B</td>
<td>selenodont</td>
<td>mixed</td>
<td>WA</td>
<td>18</td>
<td>46</td>
</tr>
<tr>
<td>Grizzly bear (Ursus arctos)</td>
<td>161</td>
<td>(2/3)</td>
<td>W/M</td>
<td>bunodont</td>
<td>omnivore</td>
<td>WA</td>
<td>18</td>
<td>36</td>
</tr>
<tr>
<td>Elk (Cervus eleaphus)</td>
<td>266</td>
<td>(0/3)</td>
<td>C/B</td>
<td>selenodont</td>
<td>grazer</td>
<td>CO</td>
<td>10</td>
<td>46</td>
</tr>
<tr>
<td>Moose (Alces alces)</td>
<td>309</td>
<td>(1/1)</td>
<td>W/B</td>
<td>lophodont</td>
<td>grazer</td>
<td>WA</td>
<td>18</td>
<td>43</td>
</tr>
<tr>
<td>Horse (Equus caballus)</td>
<td>432</td>
<td>(3/0)</td>
<td>C/M</td>
<td>lophodont</td>
<td>grazer</td>
<td>TX</td>
<td>18</td>
<td>41</td>
</tr>
<tr>
<td>Cow (Bos taurus)</td>
<td>548</td>
<td>(2/2)</td>
<td>C/M</td>
<td>lophodont</td>
<td>grazer</td>
<td>TX</td>
<td>18</td>
<td>73</td>
</tr>
</tbody>
</table>

* Number of individuals (male/female). All male horses, caribou, and cattle were castrated. Sex of lemmings unknown.
† C = captive born, W = wild born, M = maternal raised, B = bottle (human) reared.
‡ CO = Fort Collins, Colorado; TX = Uvalde, Texas; WA = Pullman, Washington.
§ Distance between bites.
† Concentrate selector (e.g., cactus fruits, pods from legumes).

Plants offered to animals in patches (Black and Kenney 1984, Spalinger et al. 1988) that were assembled as follows. We offered animals preweighed amounts of fresh alfalfa (Medicago sativa) anchored to plywood boards (25.5 cm x 28.5 cm x 12.7 mm thick for prairie dogs, and 61 cm x 61 cm x 19 mm thick for all other species except lemmings). We used alfalfa because it was palatable to a broad range of herbivores and was consistent in quality among study locations. Holes were drilled in plywood boards to accept alfalfa stems or petioles, which were firmly held with rubber stoppers inserted from the bottom of the board. Plants were uniformly spaced 4.5, 9, 10, or 18 cm apart to scale plant distribution to the size of the animal being observed (Table 1). To complete the patch, boards were bolted to frames 1.2 x 2.4 m (for prairie dogs 0.31 x 1.22 m) to form a surface where the animal foraged. Because of their small size, lemmings required a different experimental apparatus. Lemmings were offered leaves placed at 1-cm intervals in plastic clips (commonly sold as binders for report covers). These clips were roughly triangular in cross section, 13 mm high, 5 mm wide at the base, and 280 mm long. Petioles were securely held by pressure exerted at the apex of the clip. Clips were attached to an acrylic-fronted runway 9 cm wide x 61 cm long.

Hereafter, we will refer to plant size (in grams per plant) as the mass of plant tissue anchored at a single location in a patch, and bite size (S, in grams per bite) as the mass of tissue removed from the plant by a single cropping motion of an herbivore. When animals consumed the entire plant from a single location, then plant size equalled bite size. We varied plant size by changing height and bulk density of plants on the boards. In so doing, we were able to force a 10-fold or greater difference in mass between the largest and smallest bites taken by each species.

**Behavioral measures**

We measured dry matter intake rates of 2–13 individuals of each species (Table 1) feeding in artificial patches during foraging trials conducted during September 1990 through September 1991. All animals were habituated to the experimental protocols before trials were initiated. For each trial, a single animal was released into a patch consisting of plants of roughly equal size. As the animal foraged, we counted the number of bites cropped and recorded the elapsed time of active foraging. We used video tapes of trials replayed in slow motion to enhance the accuracy of our counts of cropped bites.

Because we were interested in the maximum instantaneous intake rate achievable at a given bite size, trials were kept brief, typically lasting ≈ 1.5 min. Moreover, species were fasted before observations were taken to assure a high level of interest in feeding. Food was withheld from rabbits and smaller species for 6–7 h before intake measurements. Larger species were fasted overnight (12 h). Otherwise, animals were allowed ad libitum access to a maintenance diet.

We estimated total dry matter intake during a trial as the difference between the dry mass of forage offered at the beginning of the trial and the dry mass remaining on the boards at the end of the trial. The amount of forage offered was determined by weighing the plant material before it was placed in patches. Upon completion of a trial, all plant remains were removed, dried to a constant mass in a convection or microwave oven, and weighed. Because orts were held firmly in place in the patch, we were able to collect all of the plant mass.
removing at the end of a feeding trial. The average bite size consumed during each trial was determined by dividing total amount of forage removed (i.e., total intake) by the number of cropping bites observed during that trial.

All calculations were corrected for dry matter. Dry matter corrections were determined daily from several subsamples of offered material, and varied little among days or sites.

**Model tests**

We designed three model tests. First, we examined whether the model could provide a reasonable statistical fit to the data. Model parameters were estimated by nonlinear regression (SAS 1988), and the residuals were analyzed for a systematic lack of fit using a runs test (Sokal and Rohlf 1981). Values of $r^2$ for nonlinear regressions were calculated using corrected sums of squares (Motulsky and Ransnas 1987). We separately analyzed plant size and bite size as independent variables. Both quantities offer useful features in our analyses. Clearly, plant size ultimately controls bite size, while bite size is proximally responsible for the mechanism we propose (Eq. 5). Bite size is virtually impossible to measure directly, particularly for small animals. So, we estimated bite size indirectly as a ratio (total mass removed per total number of cropping bites). The numerator of this ratio was also present in the dependent variable, intake rate (total mass removed per time). Regression analyses based on ratios are controversial (Prairie and Bird 1989, Jackson and Somers 1991, Kenney 1991). So, we also analyzed our data by regressing intake rate against direct measurements of plant size, which shares no term in common with intake rate.

Examining statistical fit of a model to data provides an opportunity for the model to fail, but it does not validate mechanisms the model represents. To do so, we derived predictions from the model and compared those predictions with independent observations. Two predictions are central to our model. The first is that cropping motions of the jaw compete with chewing motions, thereby producing a decelerating rate of intake with increasing bite size. Second, the model predicts that the asymptotic maximum intake rate (i.e., $R_{\text{max}}$) of herbivores feeding in food-concentrated patches is set by constraints on food processing in the mouth. We tested these predictions as follows.

The functional form of the competitive relationship between cropping and chewing is given by Eq. 4. If the deceleration in intake rate in Eq. 5 is produced by competition between cropping and chewing, then cropping rate must decline asymptotically as bite size increases according to Eq. 4 (Spalinger and Hobbs 1992).

Our model (Eq. 5) also predicts that maximum instantaneous intake rate is set by a processing constraint, $R_{\text{max}}$. In the absence of cropping, intake rate is theoretically limited only by the ability of the animal to masticate food sufficiently to render it suitable for swallowing. We estimated $R_{\text{max}}$ (in grams per minute) directly as the product of $R_{\text{max}}$ (in grams per minute) directly as the product of $C_e$, $C_f$ and chewing frequency (in chews per minute):

$$R_{\text{max}} = C_e \times C_f.$$  

We defined a chew to consist of a single up and down cycle of the jaw terminating when the molars occluded. We observed chewing behavior using video tapes of feeding animals replayed in slow motion. Values for chewing effort ($C_e$, Eq. 6) were determined by counting at least five, consecutive, uninterrupted chews and dividing the number of chews by the time elapsed between the first and last chew. To test the proposed mechanism producing the asymptote in our model (Eq. 5), we compared observations of $R_{\text{max}}$ (estimated as the product of $C_e \times C_f$) with predictions of $R_{\text{max}}$ obtained from the statistical fit of Eq. 5 to intake rate data.

As a final model test, we compared published observations of intake rates for a variety of species to those predicted by Eq. 5. To generate predicted intake rates, we first used body mass to match species represented in previous studies with species used in our experiments. We then substituted parameter values from the least squares fit of our functional response model (Eq. 5) to calculate a predicted intake rate for each bite size observed in previous studies. We excluded values only when the author noted that mechanical structures interfered with foraging (e.g., spines). We compared our model predictions against the observations of other workers.

**Results**

Following a brief training period, all species readily consumed alfalfa offered in the foraging trials. Species used in our experiments consumed bites ranging in mass from 0.0004 g dry matter (lemming) to 8.1 g dry matter (cow). Analyses of the model for food-concentrated patches (Eq. 5) were based on 36–125 trials (Table 1; mean = 72 trials/species). Each species consumed bites spanning at least an order of magnitude in mass, and plant size offered varied by >2 orders of magnitude for some species. We observed intake rates of <0.03 g/min for lemmings to >80 g/min for cattle. Overall, estimates of maximum intake rate ($R_{\text{max}}$) increased with body mass (Tables 1 and 2), but $h$ was independent of body mass, averaging ≈0.014 min/bite for all species. Details of our work on the allometry of model parameters will be reported elsewhere.

**Fidelity to observations**

As predicted by our model (Eq. 5), we observed an asymptotic relationship between plant size and intake rate of mammalian herbivores (Fig. 1). Plant size proved
Fig. 1. Intake rate of mammalian herbivores increased asymptotically as a function of plant size (left column on each page) and bite size. The solid line is nonlinear least squares fit of functional response model (see Eq. 5 in Introduction). Plant size is the mass of plant material available to the herbivore at a point in a hand-constructed patch. Bite size is the average mass of plant tissue cropped by an herbivore with a single cropping motion. When herbivores consumed an entire plant in a single bite then plant size equalled bite size.
FIG. 1. Continued.
to be an accurate predictor of intake rate, with an average corrected r² of 72% (Fig. 1; Table 2, all regressions were significant at P < .001). We found a systematic lack of fit (i.e., a nonrandom distribution of residuals) in only one species (rabbits, runs test, T₇ = 2.11, P < .05).

In general, using bite size (S) as an independent variable improved the fit of the model (Table 2, Fig. 1). On average, S accounted for 77% of the variance in intake rate (Table 2). A runs test performed on the residuals from Eq. 5 confirmed that there was no systematic lack of fit for 11 of the 13 species (Table 2). Asymptotes (R_max) estimated from nonlinear regression of intake rate on bite or plant size (from Eq. 2). All regressions are significant (P < .001).

### Table 2. Estimates of maximum processing rate (R_max) and cropping time (h) from nonlinear regression of intake rate on bite or plant size (from Eq. 2). All regressions are significant (P < .001).

<table>
<thead>
<tr>
<th>Species</th>
<th>R_max (g/min)</th>
<th>h (min/bite)</th>
<th>r²</th>
<th>Runs test</th>
<th>R_max (g/min)</th>
<th>h (min/bite)</th>
<th>r²</th>
<th>Runs test</th>
</tr>
</thead>
<tbody>
<tr>
<td>Lemming</td>
<td>0.15</td>
<td>0.013</td>
<td>0.74</td>
<td>NS</td>
<td>0.15</td>
<td>0.016</td>
<td>0.75</td>
<td>NS</td>
</tr>
<tr>
<td>Prairie dog</td>
<td>0.61</td>
<td>0.006</td>
<td>0.59</td>
<td>NS</td>
<td>0.53</td>
<td>0.005</td>
<td>0.41</td>
<td>NS</td>
</tr>
<tr>
<td>Rabbit</td>
<td>1.04</td>
<td>0.016</td>
<td>0.68</td>
<td>**</td>
<td>1.01</td>
<td>0.017</td>
<td>0.63</td>
<td>*</td>
</tr>
<tr>
<td>Peccary</td>
<td>4.14</td>
<td>0.026</td>
<td>0.57</td>
<td>NS</td>
<td>3.86</td>
<td>0.033</td>
<td>0.53</td>
<td>NS</td>
</tr>
<tr>
<td>White-tailed deer (fawn)</td>
<td>6.32</td>
<td>0.010</td>
<td>0.73</td>
<td>NS</td>
<td>5.99</td>
<td>0.011</td>
<td>0.72</td>
<td>NS</td>
</tr>
<tr>
<td>White-tailed deer (adult)</td>
<td>9.85</td>
<td>0.011</td>
<td>0.84</td>
<td>**</td>
<td>9.17</td>
<td>0.011</td>
<td>0.78</td>
<td>NS</td>
</tr>
<tr>
<td>Axis deer</td>
<td>8.65</td>
<td>0.008</td>
<td>0.74</td>
<td>NS</td>
<td>8.42</td>
<td>0.009</td>
<td>0.73</td>
<td>NS</td>
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<tr>
<td>Caribou</td>
<td>16.73</td>
<td>0.013</td>
<td>0.78</td>
<td>NS</td>
<td>16.29</td>
<td>0.016</td>
<td>0.75</td>
<td>NS</td>
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<tr>
<td>Grizzly bear</td>
<td>43.06</td>
<td>0.015</td>
<td>0.87</td>
<td>NS</td>
<td>40.90</td>
<td>0.022</td>
<td>0.81</td>
<td>NS</td>
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<tr>
<td>Elk</td>
<td>52.95</td>
<td>0.012</td>
<td>0.96</td>
<td>NS</td>
<td>47.41</td>
<td>0.015</td>
<td>0.89</td>
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<tr>
<td>Moose</td>
<td>31.11</td>
<td>0.038</td>
<td>0.83</td>
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<td>29.78</td>
<td>0.041</td>
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<tr>
<td>Horse</td>
<td>40.39</td>
<td>0.010</td>
<td>0.90</td>
<td>NS</td>
<td>39.31</td>
<td>0.010</td>
<td>0.89</td>
<td>NS</td>
</tr>
<tr>
<td>Cow</td>
<td>74.16</td>
<td>0.009</td>
<td>0.75</td>
<td>NS</td>
<td>74.57</td>
<td>0.011</td>
<td>0.75</td>
<td>NS</td>
</tr>
</tbody>
</table>

* P < .05, ** P < .01, NS = not significant.
† Runs test on residuals (Sokal and Rohlf 1981:782).

We estimated the functional response of the predator herbivore to plant size by feeding the alfalfa stems directly to the molars via the diastema between the incisors and molars. This allowed them to crop bites that exceeded the dimensions of the mouth. In contrast, lemmings, elk, and prairie dogs tended to restrict bite size as plant size increased (for example, Fig. 2D). Body size and feeding style did not appear to influence these patterns.

### Tests of model mechanisms

Consistent with our hypothesis, cropping rate declined at a decelerating rate as bite size increased for all species observed (Fig. 3). Maximum intake rates calculated from direct measurements of chewing rate (chews per minute) and chewing effort (chews per gram; Table 3) compared favorably with R_max calculated from Eq. 5 (r² = 0.98 for log-transformed values, r² = 0.91 for untransformed data; Fig. 4).

Our functional response model (Eq. 5) accounted for 83% of the variance in previously published observations of intake rate (Fig. 5, n = 153, log y = -0.085 + 0.988 log x, r² = 0.83, P < .001, for untransformed data y = -2.09 + 1.07 x, r² = 0.86, P < .001). The largest residuals from the regression (circled points in Fig. 5) were obtained by recording pressure changes in a pneumatic bellows that encircled the jaw of feeding caribou (Trudell and White 1981). This device recorded both jaw and lip movements and may therefore have overestimated the number of prehending bites as we defined them.

### Discussion

#### Relationship to empirical studies of functional response

Our observations of asymptotic intake rate (R_max) consistently exceeded intake rates reported for similar species of herbivores (Tables 2 and 4). However, we suggest that intake rates observed in many previous
studies never approached $R_{max}$ because bite sizes consumed in those studies severely limited intake rate (Table 4). When we used the bite sizes observed in previous studies to drive our functional response model (Eq. 5), it accounted for 83% of the variation in intake rate for a broad array of herbivores. Our model tended to overestimate intake rate (intercept < 0), yet it achieved a level of generality not exhibited by other models of herbivore functional response.

We find this generality somewhat surprising because we believed that several factors inherent to the design of our experiments should limit their applicability. We

Table 3. Chewing frequency and chewing effort for herbivores consuming fresh alfalfa.

<table>
<thead>
<tr>
<th>Species</th>
<th>Chews/min</th>
<th>Chews/g intake</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>$\bar{x}$</td>
<td>$\text{sd}$</td>
</tr>
<tr>
<td>Lemming</td>
<td>474</td>
<td>51.4</td>
</tr>
<tr>
<td>Prairie dog</td>
<td>276</td>
<td>21.4</td>
</tr>
<tr>
<td>Rabbit</td>
<td>282</td>
<td>17.6</td>
</tr>
<tr>
<td>Peccary</td>
<td>121</td>
<td>3.3</td>
</tr>
<tr>
<td>White-tailed deer (fawn)</td>
<td>145</td>
<td>8.2</td>
</tr>
<tr>
<td>White-tailed deer (adult)</td>
<td>101</td>
<td>0.5</td>
</tr>
<tr>
<td>Axis deer</td>
<td>121</td>
<td>6.3</td>
</tr>
<tr>
<td>Caribou</td>
<td>150</td>
<td>2.1</td>
</tr>
<tr>
<td>Grizzly bear</td>
<td>139</td>
<td>2.4</td>
</tr>
<tr>
<td>Elk</td>
<td>112</td>
<td>8.4</td>
</tr>
<tr>
<td>Moose</td>
<td>117</td>
<td>9.6</td>
</tr>
<tr>
<td>Horse</td>
<td>91</td>
<td>7.3</td>
</tr>
<tr>
<td>Cow</td>
<td>67</td>
<td>2.1</td>
</tr>
</tbody>
</table>
offered each animal a homogeneous patch of high-quality, low-fiber forage (fresh alfalfa). In so doing, we eliminated characteristics that could potentially reduce intake rate or feeding efficiency, such as high fiber levels (Balch 1971, Spalinger et al. 1988), a densely branched plant architecture (McNaughton 1984), or the inclusion of inedible material in the feeding station (Arnold 1964, Bell 1970, McNaughton 1978). Moreover, because our objective was to test a model of mechanical (rather than volitional) controls on intake rate, we conducted trials with highly motivated, hungry animals that were most likely to maximize short-term intake rate (Demment and Greenwood 1988, Dougherty et al. 1989c). We caution that in some situations, these influences may strongly impact intake rate (Dunham 1980, Arthur 1984) and may limit the utility of our model.

Temporal scale is central to understanding an animal's functional response. We focused on instantaneous intake rate and the processes that control it. Because animals typically consumed our hand-assembled patches in a systematic manner, there was essentially no patch depletion during the course of a measurement; the distance between bites was the same for the first two bites of a trial and the last two. In addition, because the mechanical processes responsible for the functional response operate on a time scale of seconds or minutes, we limited the length of feeding trials. Studies of the relationship between food availability and intake rate of herbivores have focused on different time

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**Fig. 3.** Cropping rate declined non-linearly with increased bite size for all species, as predicted by Eq. 4. We show six representative species differing in body size and feeding style; other species show similar trends.

**Fig. 4.** Predicted maximum intake rate ($R_{\text{max}}$) was highly correlated with observations based on chewing rate and chewing effort. Best fit to line shown is $\log_{10}y = 0.039 + 0.948 \log_{10}x (r^2 = 0.98, P < .001)$. For untransformed data, $y = 0.432 + 0.87x (r^2 = 0.91, P < .001)$. 
scales, ranging from periods of a few minutes (this study, Trudell and White 1981, Black and Kenney 1984, Hudson and Watkins 1986, Spalinger et al. 1988), 10's of minutes (Batzli et al. 1981, Milne et al. 1982, Lundberg and Danell 1990), one to several hours (Allden and Whittaker 1970, Short 1986, Lundberg 1988, Dougherty et al. 1989a), or ≥24 h (Chacon and Stobbs 1976, Jamieson and Hodgson 1979, Hodgson and Jamieson 1981, Short 1985). Direct comparisons between these studies are difficult because constraints imposed by short-duration mechanical processes, such as cropping and chewing time, are confounded with processes that operate over longer time frames, such as digestion, gut fill, or satiety (e.g., Suzuki et al. 1969, Dougherty et al. 1989c). The interaction among processes regulating intake over different time scales offers an important, unresolved question in foraging ecology.

Our results suggest that bite size is a useful predictor of short-term intake rate of herbivores (Figs. 1 and 5). Traditionally, biomass density (in grams per square metre) has been used for this purpose. The use of biomass density to predict intake can be traced to the historical predominance of studies of domestic ruminants feeding in grasslands (reviewed by Hodgson 1985 and Forbes 1988). In habitats with a relatively simple structure, such as grass swards, there is a strong correlation between bite size and biomass density (Stobbs 1975, Black and Kenney 1984, Hudson and Nietfeld 1985, Hudson and Watkins 1986, Penning et al. 1991), and consequently either measure can effectively be used to predict intake rate of grazers. However, a bite cropped by a browser typically consists of a leaf, or a twig with a few leaves. Hence, the size of bites cropped by browsers is often controlled by leaf size. When leaf size con-

### Table 4. Maximum intake rates of mammalian herbivores consuming fresh forages.

<table>
<thead>
<tr>
<th>Species</th>
<th>Body mass (kg)</th>
<th>Bite size (g)</th>
<th>Intake rate (g/min)</th>
<th>Forage or habitat</th>
<th>Source</th>
</tr>
</thead>
<tbody>
<tr>
<td>Bank vole</td>
<td>0.04*</td>
<td></td>
<td>0.012</td>
<td>willow shoots</td>
<td>Lundberg 1988</td>
</tr>
<tr>
<td>Brown lemming</td>
<td>0.05</td>
<td></td>
<td>0.11</td>
<td>arctic sod</td>
<td>Batzli et al. 1981</td>
</tr>
<tr>
<td>Domestic lamb</td>
<td>40†</td>
<td>0.20</td>
<td>5.0</td>
<td>service-berry shrub</td>
<td>Flores et al. 1989a</td>
</tr>
<tr>
<td>Impala</td>
<td>40</td>
<td>0.37</td>
<td>7.48</td>
<td>Diospyros lycioides</td>
<td>Dunham 1980</td>
</tr>
<tr>
<td>Ewe</td>
<td>40†</td>
<td>0.168</td>
<td>5.7</td>
<td>perennial ryegrass</td>
<td>Penning et al. 1991</td>
</tr>
<tr>
<td>Sheep</td>
<td>41.5</td>
<td></td>
<td>6</td>
<td>Kikuyu grass</td>
<td>Black and Kenney 1984</td>
</tr>
<tr>
<td>Mule deer</td>
<td>42</td>
<td>0.35</td>
<td>6</td>
<td>mixed diet</td>
<td>Wickstrom et al. 1984</td>
</tr>
<tr>
<td>Sheep</td>
<td>43</td>
<td>0.4</td>
<td>7.5</td>
<td>ryegrass</td>
<td>Allden and Whittaker 1970</td>
</tr>
<tr>
<td>Black-tailed deer</td>
<td>48</td>
<td>0.28</td>
<td>7.5</td>
<td>Rubus spectabilis</td>
<td>Spalinger et al. 1988</td>
</tr>
<tr>
<td>Mule deer</td>
<td>60</td>
<td></td>
<td>3.3</td>
<td>clear-cut forest</td>
<td>Collins and Unness 1983</td>
</tr>
<tr>
<td>Reindeer</td>
<td>70</td>
<td></td>
<td>12.7</td>
<td>Pedicularis landgorfii</td>
<td>Trudell and White 1981</td>
</tr>
<tr>
<td>Elk</td>
<td>225†</td>
<td>0.36</td>
<td>12.6</td>
<td>grass sward</td>
<td>Hudson and Nietfeld 1985</td>
</tr>
<tr>
<td>Elk</td>
<td>160</td>
<td>1.6</td>
<td>20</td>
<td>mixed diet</td>
<td>Wickstrom et al. 1984</td>
</tr>
<tr>
<td>Elk</td>
<td>231</td>
<td></td>
<td>17.2</td>
<td>clear-cut forest</td>
<td>Collins and Unness 1983</td>
</tr>
<tr>
<td>Elk</td>
<td>250†</td>
<td>0.5</td>
<td>22.5</td>
<td>wet meadow</td>
<td>Collins et al. 1978</td>
</tr>
<tr>
<td>Elk</td>
<td>250†</td>
<td>0.7</td>
<td>21</td>
<td>cured pasture</td>
<td>Hudson and Watkins 1986</td>
</tr>
<tr>
<td>Moose</td>
<td>340</td>
<td>1.2</td>
<td>22.5</td>
<td>summer forages</td>
<td>Renecker and Hudson 1986</td>
</tr>
<tr>
<td>Moose</td>
<td>350*</td>
<td></td>
<td>29</td>
<td>birch trees</td>
<td>Astrøm et al. 1990</td>
</tr>
<tr>
<td>Moose</td>
<td>360</td>
<td></td>
<td>16.7</td>
<td>deciduous leaves</td>
<td>Belovsky and Jordan 1978</td>
</tr>
<tr>
<td>Cow</td>
<td>471</td>
<td>1.46</td>
<td>49</td>
<td>fresh alfalfa</td>
<td>Dougherty et al. 1989c</td>
</tr>
<tr>
<td>Cow</td>
<td>400†</td>
<td>1.22</td>
<td>74.1</td>
<td>open grassland</td>
<td>Arthur 1984</td>
</tr>
</tbody>
</table>

* Nowak and Paradiso 1983.
† Personal estimate.
trols bite size, bite size and biomass density are un-
coupled (Spalinger et al. 1988, Spalinger and Hobbs
1992), and the predictive power of biomass density is
lost (Trudell and White 1981, Wickstrom et al. 1984,
Renecker and Hudson 1986, Spalinger et al. 1988). The
failure of biomass density to provide general predic-
tions of herbivore functional response emphasizes the
need for mechanistic models that respond to the variety
of foraging situations encountered by herbivores. We
think that a better understanding of the features of
plants and animals that control bite size is central to
this development.

Relationship to theory on functional response

Most contemporary models of functional response
represent foraging by predators. Because herbivores
encounter foraging situations that differ markedly from
those confronted by predators, traditional functional
response models often fail to describe processes regu-
lating intake rate by herbivores. For example, Holling's
disc equation and the random parasitoid model require
that searching for food and handling it occur as mu-
ally exclusive events (Holling 1959, Rogers 1972).
Asymptotic forms result in these models because
searching competes with handling, i.e., the predator
can allocate its time to one activity or the other, but
not both simultaneously. In addition, functional re-
sponse models designed for predators predict the num-
ber of similar-sized prey captured per unit time (Holl-
ing 1959, Rogers 1972, Stephens and Krebs 1986,
herbivores varies dynamically; it can change by more
than an order of magnitude even when animals are
feeding on the same plant species (Fig. 2). Moreover,
while predators must often reside at a prey item to
consume it, herbivores can process bites from one plant
as they move through the environment looking for
another. Because searching and handling overlap in
time, searching does not compete with handling when
plants are sufficiently concentrated in space.

Our model is a Michaelis-Menten function that can
be shown to be algebraically identical to the disc equa-
tion of Holling (1965) under the assumption that the
cropping rate is a linear function of bite (i.e., prey)
density (Real 1977). However, this assumption clearly
does not hold for herbivores feeding in food-concen-
trated patches. Cropping rates changed over an order of
magnitude even when the density of plants (or bites)
offered to animals was held constant (Fig. 3). Thus,
although our model is mathematically consistent with
the disc equation, it is functionally different.

The primary difference between our model and pre-
vious functional response models follows from as-
sumptions of the foraging process. Our experiments
substantiate the idea that competition between crop-
ning and chewing regulate the instantaneous intake rate
of herbivores feeding in food-concentrated patches.
These results are analogous to the competition between
searching and handling in predator-based models; the
herbivore can allocate its time to cropping or to chew-
ing but not both simultaneously (Fig. 3, also see Ardran
between cropping and chewing does not require
that an herbivore finish chewing one bite before crop-
ping another, i.e., at any instant an animal may be
chewing several bites at once. But, this does mean that
the overall cropping rate is limited by the chewing rate,
i.e., the rate of input to the mouth cannot exceed the
rate of output. Large bites require longer chewing times
than small ones and, hence, depress cropping rate
through competition for jaw movements. We surmise
from our experiments in food-concentrated patches that
the mechanism represented in the functional response
models of Spalinger and Hobbs (1992) and Laca and
Demmert (1992) faithfully represents these features of
short-term regulation of intake. We suggest that the
fidelity of the model to this mechanism explains its
broad applicability to herbivores of vastly different body
size and feeding style (Figs. 1 and 5).

Our observations allow us to reject the hypothesis
that input to the mouth regulates maximum intake rate
in mammalian herbivores. Clutton-Brock and Harvey
(1983) proposed that because buccal capacity deter-
mines bite size, maximum intake rate is proximally
limited by the volume of the mouth. They suggested
that intake rate responds directly to the amount of food
consumed by each bite, because cropping rate is rela-
tively constant. Their hypothesis leads to predictions
inconsistent with our observations. First, it suggests a
linear increase of intake rate with bite size (i.e., Eq. 5),
that is, a linear Type I as opposed to an asymptotic
Type II functional response. In contrast, we observed
a nonlinear, asymptotic response of intake rate to bite
size (Fig. 1). Second, Clutton-Brock and Harvey (1983)
suggested that cropping rate is insensitive to bite size,
but we observed an inverse relationship between crop-
ning rate and bite size (Fig. 2). The close correspon-
dence of our observed maximum intake rate and that
predicted from chewing rate and chewing effort (Fig.
4) lends strong support to the idea that the most im-
portant determinant of intake rate is processing ca-
pacity. Hence, we conclude that although cropping rate
is proximally responsible for intake regulation, pro-
cessing bites for swallowing ultimately controls intake
by regulating cropping rate.

Identifying how cropping and chewing compete to
regulate intake rate provides a foundation for predict-
ing how plant morphology and structure can influence
foraging strategies of herbivores. Plant fibrosity (Balch
1971) or cell wall thickness (Spalinger et al. 1988) will
reduce the processing rate \( R_{\text{max}} \) by increasing chewing
effort (Spalinger et al. 1988), but may have little or no
direct effect on cropping time \( \left( \overline{h} \right) \). On the other hand,
we postulate that reductions in intake rate related to
spinescence (Dunham 1980, Cooper and Owen-Smith
1986) or changes in plant morphology (McNaughton
result from an increase in the time required to crop bites (\(\bar{H}\)), but are unrelated to processing rate (\(R_{\text{max}}\)). Because cropping time is relatively more important to functional response as bite size diminishes, we expect that plants with small bites (e.g., small leaves) will be much more effectively protected by mechanical structures than plants with large bites. When leaves are large, more time is spent chewing and features that increase chewing effort (e.g., lignin) will have more impact on intake rate. These factors also emphasize the limitations on inferences offered by our data. We conducted trials with alfalfa plants in peak condition, with very high palatability and relatively low fiber contents. Therefore, our measures of cropping rate, chewing effort, and the resulting \(R_{\text{max}}\) are likely to differ from those obtained in other settings.

Although previous models of herbivore functional response have provided useful statistical descriptions of observations of intake rate (e.g., Allden and Whitaker 1970, Wickstrom et al. 1984, Hudson and Watkins 1986, Hudson and Frank 1987, Lundberg and Danell 1990), they failed to provide testable hypotheses on the processes regulating intake. In contrast, our model describes a specific mechanism. Our tests of the model offer evidence that this mechanism, competition between cropping and processing, is responsible for Type II functional responses frequently observed for herbivores feeding in food-concentrated patches.

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**LITERATURE CITED**


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