

Mannheimia haemolytica Immunity

Are We There Yet?

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“Vaccine history: The past as prelude to the future.”

Vaccine 2012

History informs us about progress in vaccine development

- 1. Progress is made incrementally**
- 2. Progress often requires “game-changing” event or events**
- 3. Progress is closely tied to development of improved technologies from other fields**
- 4. Progress will occur through application of novel science-based technologies and strategies**

Objectives of the presentation

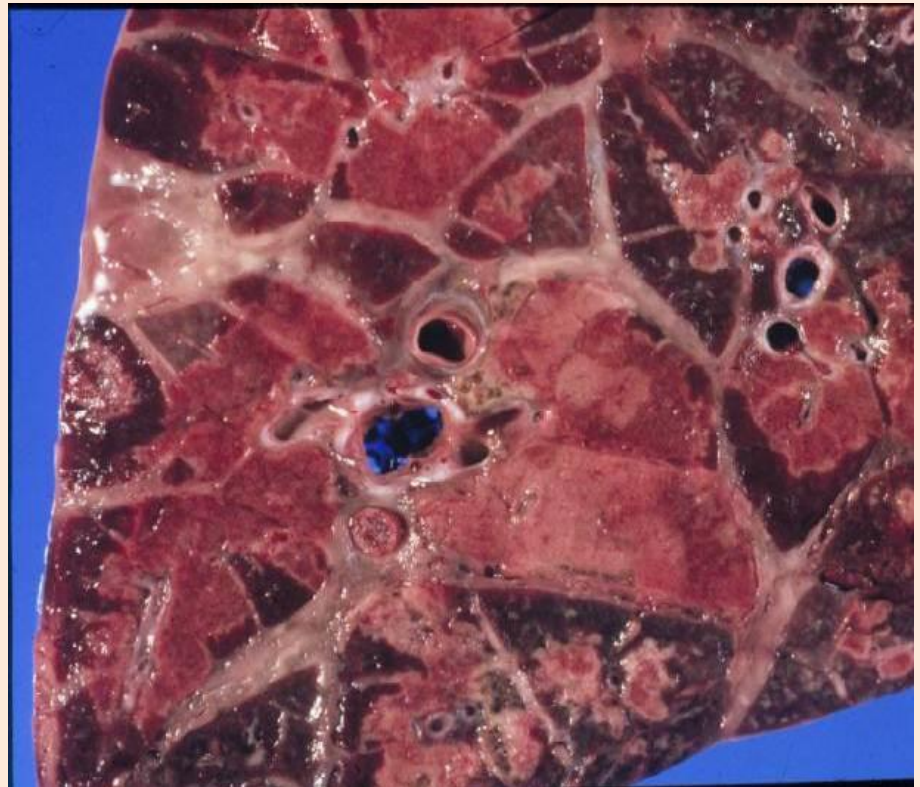
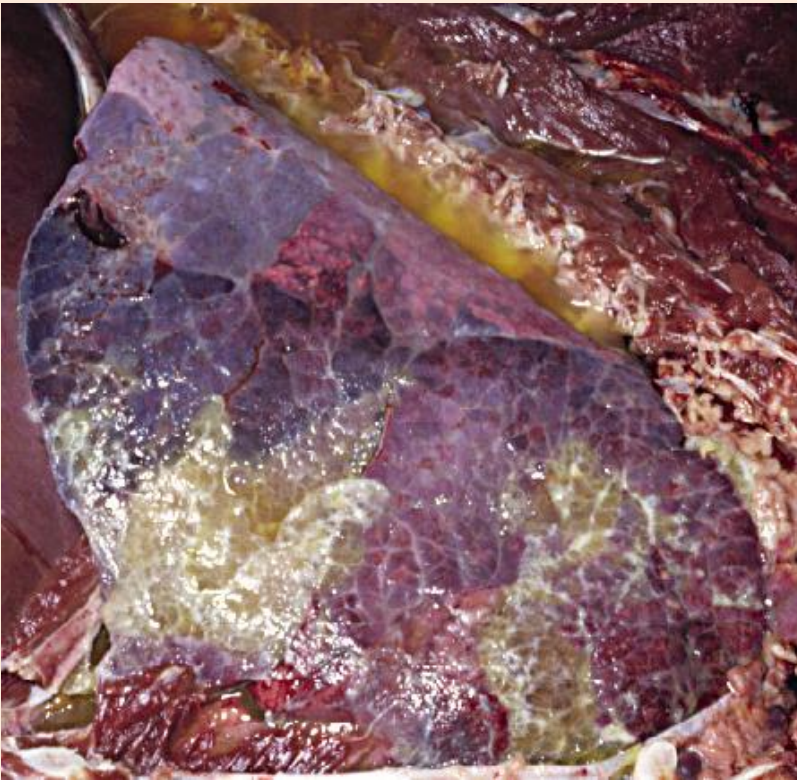
- Review several historically important findings in *M. haemolytica* pathogenesis & immunity
- Provide overview of experimental approaches in improving *M. haemolytica* vaccines



***M. haemolytica* is associated with severe bovine
bacterial pneumonia**

Acute fibrinous pleuropneumonia.

- Shipping fever – beef cattle
- Enzootic Pneumonia – dairy calves



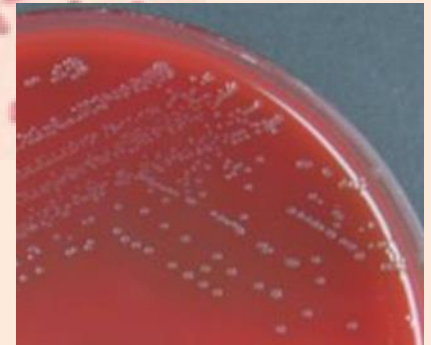
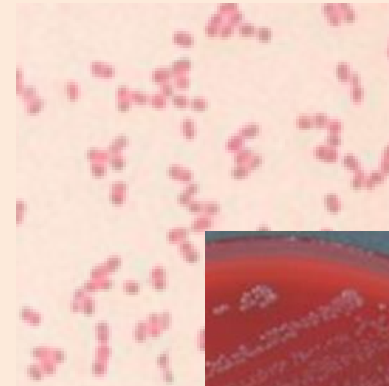
“Failure of innate immunity!” Robert Fulton

Mannheimia haemolytica

Gram negative coccobacillus

Previous names:

- *Bacillus bovisepiticus*
- *Pasteurella haemolytica*
 - Biotype A – Arabinose fermenters
 - Biotype T – Trehalose fermenters
 - Divided among serotypes based on capsular antigens



As of 1999: *Mannheimia haemolytica*

- 11 Biotype **A** serotypes - 1,2,5,6,7,8,9,12, 13, 14, and 16 (Angen et al. *IJSEM* 49:67086, 1999)
- Serotype 1 (S1) is responsible for 60% or more of pneumonia cases

Critical findings 1970s & 1980s

Pathogenesis: Changes in the nasopharyngeal flora of stressed or viral infected calves.

- In stressed calves, *M. haemolytica* proliferate and are in increased concentrations in the tracheal air (Grey & Thomson, CJCM, 1971)
- Serotype 1 is in low nasal concentrations until stressed or viral infected; then S1 is readily isolated (Frank and Smith, AJVR, 1983; 1986)

Pathogenesis: Discovery of leukotoxin

- *M. haemolytica* secretes a leukotoxin (then called “cytotoxin”) that kills leukocytes from ruminants (Shewen & Wilkie, I & I, 1982)



R.G. Thomson



G.H. Frank &
R.E. Briggs



B.N. Wilkie &
P.E. Shewen

Critical findings 1970s & 1980s

Immunity

- **At feedlot entry, cattle with higher anti-*M. haemolytica* antibodies have less respiratory disease than those with low antibodies** (Thomson et al., CJCM, 1975)
- **Bacterins do not protect and may enhance disease** (Friend et al., CJCM 1977; Wilkie et al., AJVR 1980)
- **Cattle dying of shipping fever had lower LKT neutralizing titers than did those that died of other causes** (Shewen & Wilkie, CJCM 1983)
- **Direct correlation of LKT neutralizing antibody titers and resistance to *M. haemolytica* challenge** (Gentry et al., VI & I, 1985)
- **Immunity requires antibodies to leukotoxin and to surface antigens** (Shewen & Wilkie, CJVR, 1988)
- **Major surface antigens are not LPS or capsule but OMPs** (Mosier et al., I & I, 1989; Confer et al., AJVR 1986; Confer et al., AJVR 1989)

Central Dogma of Vaccine-induced Immunity to *M. haemolytica*

- Immunity is serum antibody-mediated
- Antibodies **MUST** neutralize leukotoxin
- Antibodies against surface antigens (OMPs) must stimulate complement-mediated killing and/or phagocytosis & killing
- When given properly, vaccines that stimulate antibodies to surface antigens and to leukotoxin **SHOULD** reduce colonization of the lower respiratory tract & protect cattle

Commercial *M. haemolytica* Vaccines currently available or available in the past

- Bacterin (“antigens from chemically inactivated cultures”)
- Bacterin – leukotoxoid combination
- Leukotoxin-rich culture supernatant
- Recombinant leukotoxin-outer membrane combination
- Live streptomycin-dependent mutant vaccine (parenteral or intranasal delivery)
- Other avirulent(?) live cultures*
- Autogenous vaccines

*no longer marketed

Commercial *M. haemolytica* vaccines: Do they work?

- **18 *M. haemolytica* or *M. haemolytica* + *P. multocida* vaccine field trials**
 - **3/18 significant reduction in BRD morbidity**
 - **4/18 increased BRD morbidity**
 - **11/18 decreased morbidity but not statistically significant**
- **“the published body of evidence does not provide a consistent estimate of the direction and magnitude of effectiveness in feedlot cattle vaccination against *Mannheimia haemolytica*, *Pasteurella multocida*, or *Histophilus somni*.” (Larson & Step, *Vet Clin N Amer Food An Prac* 2012)**

What are approaches to potentially improve *M. haemolytica* vaccines?

Better Than Ever

UPDATED



New And Improved

RESULTS GUARANTEED

Potential modern approaches to bacterial vaccines

- **Recombinant protein subunit vaccines**
- **Chimeric protein vaccines**
- **Genetically modified bacterial vaccines**
- **Live recombinant organisms**
- **DNA vaccines**
- **Bacterial ghosts**
- **Bacterial vesicles**
- **Alternative delivery methods**
- **Immunostimulants**

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Recombinant protein vaccines: Addition of Immunogenic Recombinant *M. haemolytica* Proteins to Commercial Vaccines: Recombinant LKT (rLKT)

Conlon et al. (*Infect. & Immun.* 1991)

- Vaccination with rLKT alone did NOT stimulate protection
- Addition of rLKT increased efficacy of a culture supernatant vaccine (Presponse) against experimental challenge with *M. haemolytica* with reduced clinical signs and lesions.

Recombinant protein vaccines: Addition of Immunogenic Recombinant *M. haemolytica* Proteins to Commercial Vaccines: Recombinant Sialoglycoprotease (rSGP)

Shewen et al. (*Vaccine* 2003) — SGP, a protease in culture supernatant.

- Addition of rSGP fusion protein (Gcp-F) & rLKT increased efficacy of a culture supernatant vaccine (Presponse) against experimental challenge with *M. haemolytica* with lower mean clinical scores, but the differences were not significant.

Recombinant protein vaccines: Addition of Recombinant *M. haemolytica* Proteins to Commercial Vaccines: Recombinant OMP PlpE (rPlpE)

PlpE: a major surface-exposed 45 kDa outer membrane lipoprotein of *M. haemolytica* with sequence homology between serotypes 1 & 6. (Ayalew et al., *Vet Microbiol* 2006; Confer et al., *Vaccine* 2003 & 2006; Pandher et al., *I & I* 1998)

- **Addition of 100 µg of rPlpE increased efficacy of a culture supernatant vaccine (Presponse) or bacterin toxoid (One Shot) against experimental challenge with *M. haemolytica* S1 or S6.**

Addition of rPlpE to Presponse[®]: Mean Lung Lesion Scores \pm SD after challenge with *M. haemolytica* Serotype 1

Group	Lesion score (% reduction)
Control	7.75 \pm 3.58
Presponse[®]	3.00 \pm 1.26 (67.9%)
Presponse/PlpE	1.08 \pm 0.92 (95.3%)

Addition of PlpE improved resistance by 27.4%

Addition of rPlpE to Presponse® followed by Serotype 6 challenge

Vaccine	No. of cattle	Mean lesion \pm SD (% reduction)
Control – adjuvant only	6	8.1 \pm 2.2
100 μ g PlpE + adjuvant	8	4.4 \pm 4.7 (45.1%)
Presponse®	8	4.8 \pm 2.2 (41.2%)
Presponse® + 100 μ g PlpE	8	2.0 \pm 1.2 (75.3%)

Addition of PlpE improved resistance by 34.1%

Other potential recombinant *M. haemolytica* OMPs for vaccine consideration

Serotype-specific antigen-1

- **Highly conserved between S1 & S2 (Gonzalez et al., *Infect & Immun* 1995)**
- **Highly immunogenic mice and cattle (Ayalew et al., *CVI* 2011, Lo et al. *Infect & Immun* 1991)**
- **Addition of rSSA-1 to other recombinant proteins enhanced responses to those proteins (Ayalew et al., *CVI* 2011)**

GS60 – Surface-exposed outer membrane lipoprotein (Weldon et al., *Vet Microbiol* 1994; Lo & Mellors, *Vet Microbiol* 1996).

- **Conserved among all *M. haemolytica* serotypes**
- **Correlation between antibodies to Gs60 and resistance to challenge (Orouji et al., *CJVR* 2012)**

Other potential recombinant *M. haemolytica* OMPs for vaccine consideration

OmpA – Conserved OMP with adhesin properties (Kisiela & Czuprynski, I & I 2009; Lo & Sorensen, FEMS Microbiol Lett. 2007)

- High antibodies correlate with resistance against experimental challenge (Mahasreshti et al., Infect & Immun, 1997)
- Highly immunogenic (Ayalew et al., CVI 2011)
- Anti-OmpA antibodies stimulate complement-mediated killing
- Addition of rOmpA to other recombinant OMP may reduce responses (Ayalew et al., CVI 2011, Zeng et al., PhD dissertation, 1999)

PlpF – outer membrane lipoprotein (Ayalew et al., Vaccine 2011)

- Conserved among S1, S2, & S6 with variations in repeats regions
- Highly immunogenic in mice and cattle
- Stimulates high titers of C'-mediated bactericidal antibodies
- Protection studies not done

Chimeric (fusion) protein vaccines

Recombinant proteins derived from the spliced genes for multiple proteins.

Experimental *Bordetella bronchiseptica* fimbrial protein-*M. haemolytica* LKT Chimeric Protein Vaccine: Recombinant genes expressing a fusion protein composed of combinations of

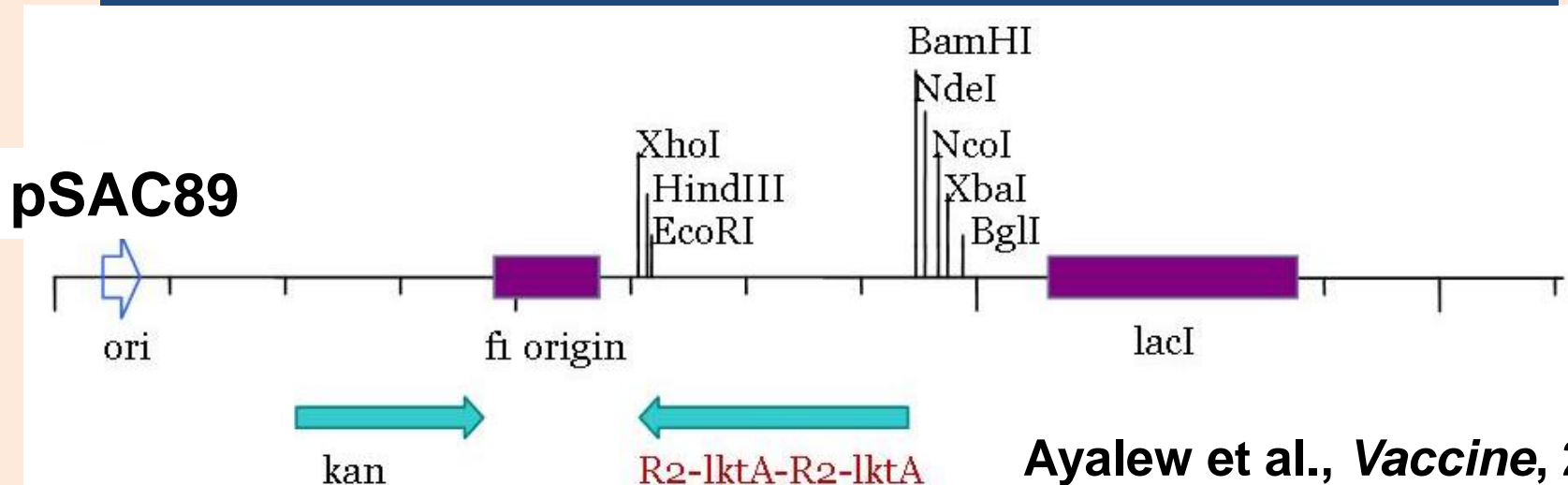
- C-terminus-neutralizing region of *lktA*
 - Fimbrial protein (*fim N* gene)
 - Glutathione-S-transferase (GST)
- Vaccination of mice resulted in anti-LKT antibodies
- Rajeev et al., *Vaccine*, 2001

Chimeric protein vaccines: PlpE-LKT

Chimeric Protein Vaccine

- Plasmids developed that expressed several chimeric genes (pSAC86-89, pSAC91) composed of various combinations of:
 - C-terminus-neutralizing region of *lktA* (NLKT)
 - N-terminus – major surface epitope (R2) of *PlpE*

SAC89 protein: R2-NLKT-R2-NLKT



Ayalew et al., *Vaccine*, 2008

Vaccination of cattle with 100 µg SAC89 + bacterin: *M. haemolytica* challenge

Group	Lesion score (% reduction)
SAC89 + adjuvant	7.1 ± 6.3 (39.6%)
SAC89 + Bacterin + adjuvant	3.1 ± 1.2 (73.7%)
Bacterin + adjuvant	7.6 ± 6.8 (34.7%)
PBS + adjuvant	11.7 ± 9.7

Addition of PlpE/LKT chimeric protein enhanced protection of a bacterin by 39%

Confer et al., *Vaccine*, 2009

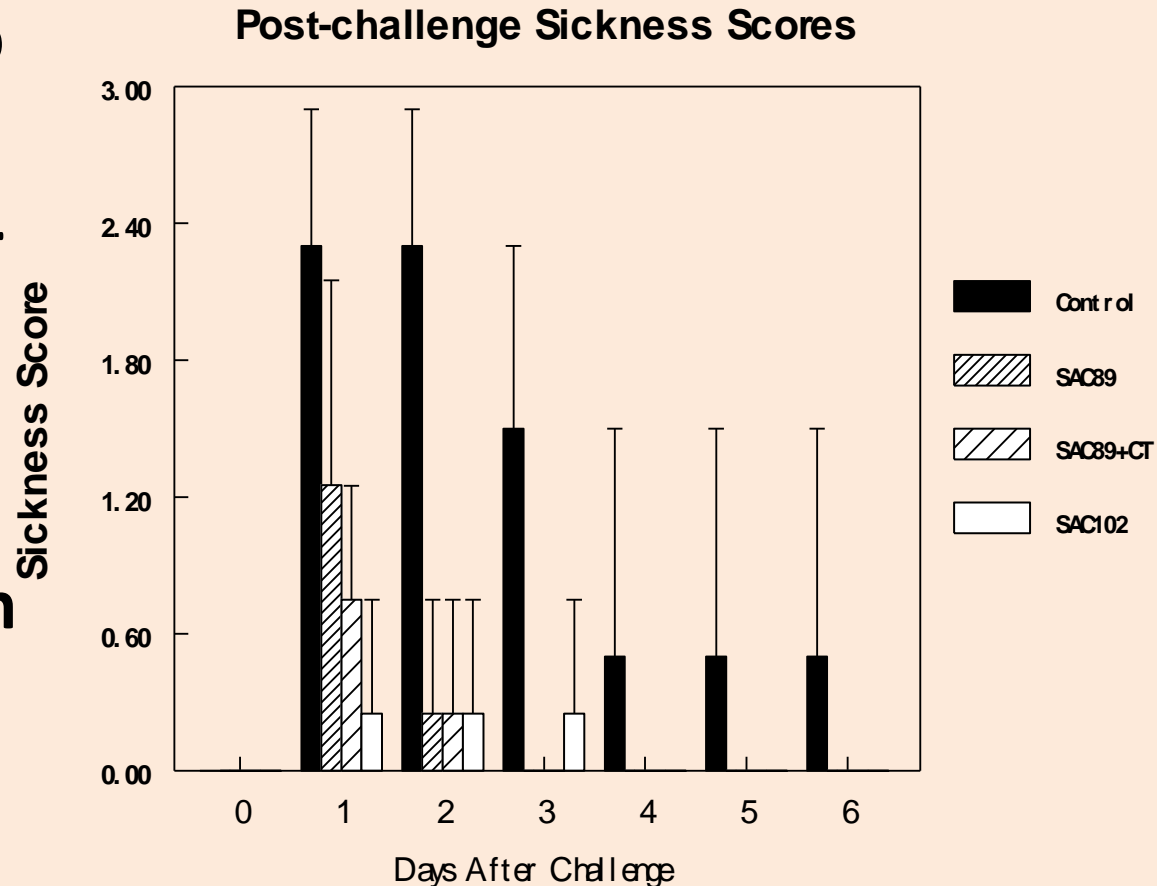
Intranasal CTB-R2-NLKT Chimeric Protein Vaccine in Cattle

- Cholera toxin is one of best mucosal adjuvants
- Because of potential hazard in using cholera toxin, SAC102 was developed: Protein derived from a chimeric gene for Cholera Toxin Subunit B (CTB)-major epitope of PlpE (R2)-neutralizing epitope of leukotoxin (NLKT).
- In IN vaccinated calves, SAC102 stimulated serum antibodies against formalin-killed *M. haemolytica*, PlpE, and LKT

Ayalew et al., *Vet Immunol & Immunopathol*, 2009

Clinical responses of SAC102 vaccinates after intrabronchial challenge with *M. haemolytica*

Clinical responses to challenge evaluated using 0-4 scale criteria (DART™). Significantly less clinical disease with SAC102 vaccinates.



M. haemolytica chimeric vaccine – Bighorn sheep

- Vaccination of mice with mammalian cell culture-expressed LKT/PlpE chimeric protein stimulated antibodies to LKT and PlpE Batra et al., *Vet Immunol Immunopathol* 2016
- Bighorn sheep vaccinated intranasally with recombinant BHV-1 vectored vaccine encoding LKT neutralizing epitope and surface-dominant epitope of PlpE. Batra et al., *Vaccine* 2017
- Sheep developed antibodies but were not protected against *M. haemolytica* challenge.



So, where are we with commercial *M. haemolytica* vaccines “spiked” with recombinant proteins?

- Under experimental conditions, supernatant and bacterin-toxoid vaccines can be enhanced by adding recombinant antigens.
- Chimeric vaccines alone may not induce complete protection
- Understandably, animal health companies have been reluctant to add recombinant proteins to their current vaccines due to increasing cost of production and cost to producers.
- One vaccine, NUPLURA™ PH by Elanco, contains rLKT and “extracted and purified outer membrane proteins”.

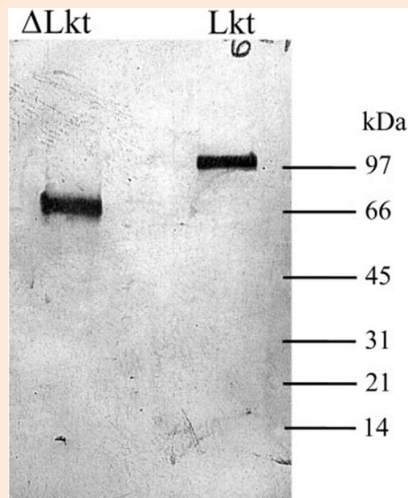
Genetically modified *M. haemolytica* vaccines

- **Streptomycin-dependent mutant *M. haemolytica***
 - Induced by N-Methyl-N'-nitro-N-nitrosoguanidine (Chengappa & Carter, *AJVR* 1979)
 - Streptomycin-dependent *Pasteurella multocida* (type A:3) and *M. haemolytica* (type 1) vaccination improved performance in a field trial (Kadel et al., *AJVR* 1985)
 - Commercial vaccine ONCE PMH[®]
- **AroA deletion mutants (AroA required for synthesis of aromatic amino acids)**
 - Homchampa et al. (*Vet Microbiol* 1994) reported generation of *M. haemolytica* aroA mutant. Mutant highly attenuated in a mouse challenge model and mice immunized with the mutant were protected against challenge.

Genetically modified *M. haemolytica* vaccines

LKT mutants

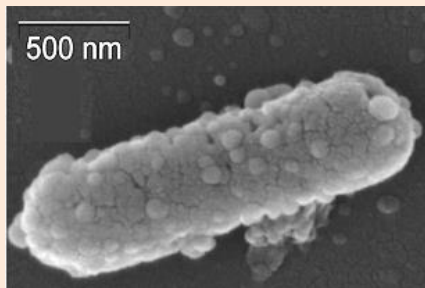
- Multiple isogenic *lkt*⁻ mutants *M. haemolytica* studied mainly related to pathogenesis
- Briggs et al. (*Microb Pathog* 2012) reported *lktA* deletion mutant that is a non-hemolytic truncated form of LKT (Δ LKT) that stimulates anti-LKT but is not leukotoxic.
 - Subcutaneous and oral vaccination of the MLV – Δ LKT *M. haemolytica* had significantly reduced lung lesions following challenge than did controls.



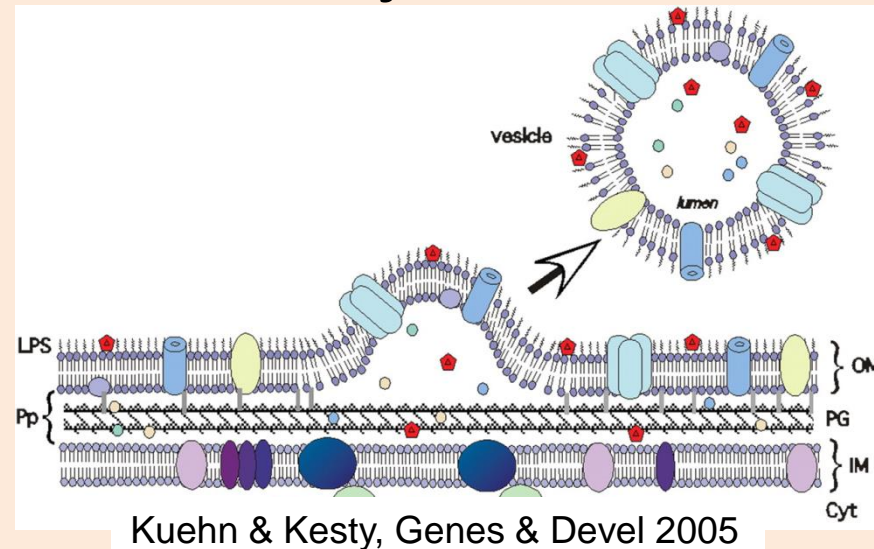
Group	% Lung lesions (% reduction)
IM vaccinates	7.0 \pm 7.3 (78%)
Oral vaccinates	4.4 \pm 4.5 (86%)
Controls	32.0 \pm 13.4

Bacterial vesicle vaccines

- Rapidly growing bacteria produce outer membrane “blebs” that detach as vesicles (outer membrane vesicles or OMV).
- Vesicles contain full complement of membrane proteins and secreted proteins, such as toxins.
- Highly immunogenic and do not require bactericidal treatments that can damage immunogenicity of proteins.
- In some cases serve as their own adjuvant



Ellis & Kuehn, Microbiol Mol Biol Rev 2010



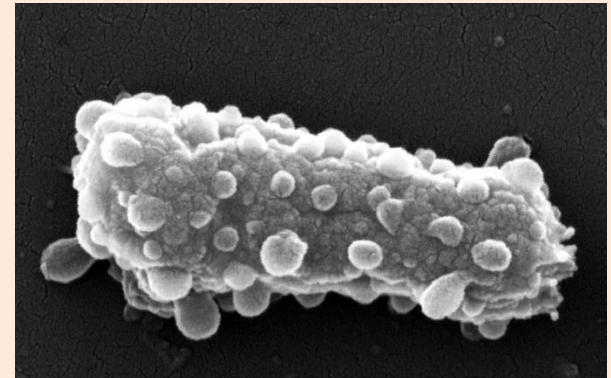
Kuehn & Kesty, Genes & Devel 2005

M. haemolytica vesicle (MHV) vaccines

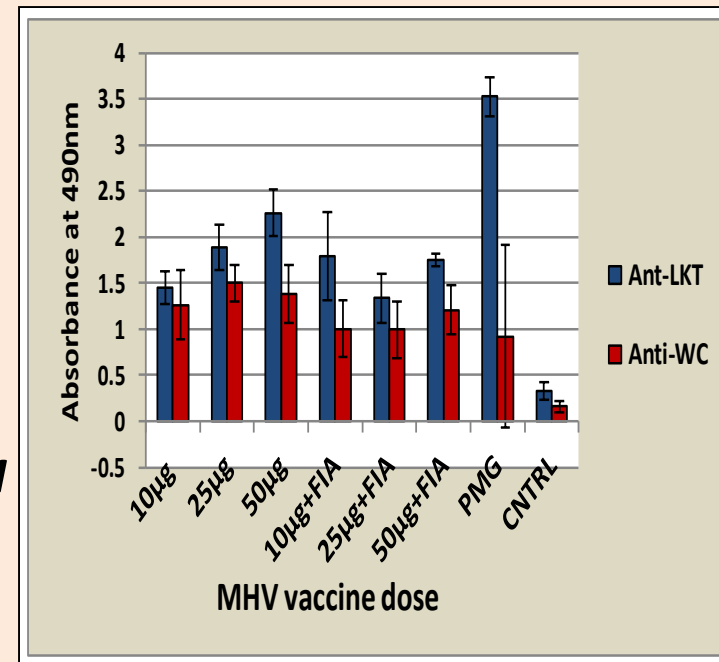
Ayalew et al., *CVI* 2013 - Proteomic analyses – MHV full complement of OMPs + many secreted proteins including LKT

- Vaccination with MHV stimulated high anti-whole cell and anti-LKT antibodies in mice and calves.
- **After challenge, MHV-vaccinated calves compared to controls**
 - 44.2% lower clinical scores ($p < 0.05$)
 - 62.8% less severe pneumonia ($p < 0.05$)

Roier et al., *Int J Med Microbiol* 2013 – Similarly demonstrated *M. haemolytica* vesicle vaccination of mice stimulated antibody responses.



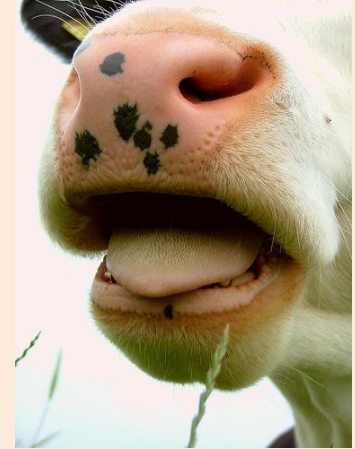
M. haemolytica – SEM photo by Dr. K. Kocan



Additional approaches

Alternative routes

- **Intranasal**
 - One commercial vaccine for IN delivery
 - Recently, we found and characterized two *M. haemolytica* IgA proteases, which may assist the bacterium to evade mucosal immunity. IgA proteases are potential intranasal vaccine targets.
(Ayalew et al., *Vet Microbiol* 2017)
- **Oral**
 - Attempts by R. Lo & Shewen to make an edible vaccine using transgenic alfalfa. Seems to be off the table



Immunostimulants

- **Probiotics**
 - Diaz et al., *Benef microbes*, 2018 –Mice vaccinated with *M. haemolytica*/*P. multocida* vaccine and given intragastric *Enterococcus faecalis* CECT7121 had enhanced antibody response, antibody avidity, and higher interferon- γ than with vaccine alone.
- **Unmethylated CpG DNA dinucleotides**
 - Stimulate innate and adaptive immunity through TLR9. Role in enhancing *M. haemolytica* vaccine?
 - Addition of CpG to *Bordetella pertussis* antigens enhanced production of IFN- γ in mice following vaccination. (Bakhshaei et al., *J IFN Cytokine Res*, 2018)

ARE WE THERE YET !?!



Relative to the four points made on vaccine history (*Vaccine* 2012)

- 1. Progress is made incrementally**
 - **Incremental progress made since 1980s by incorporating LKT into vaccines and understanding the role of surface antigens.**
- 2. Progress often requires “game-changing” event or events**
 - **Discovery of LKT was A MAJOR game changer. What are the next ones?**
- 3. Progress is closely tied to development of improved technologies from other fields**
 - **Through molecular biology and genomics, we better understand the antigens and epitopes involved in immunity and starting to better understand the respiratory microbiome.**
- 4. Progress will occur through application of novel science-based technologies and strategies**
 - **Future, more efficacious vaccines will apply molecular techniques, improved novel production techniques, immunostimulants, and/or better vaccine delivery methods.**

A. W. Confer – Official retirement date, July 1, 2019

“Regrets, I’ve had a few. But then again, too few to mention.” Frank Sinatra - *I did it my way*



My *M. haemolytica* regrets that I WILL mention

- **Our lab:** Focused too much on the organism and serum antibody and not enough on *M. haemolytica*/host interactions & innate immunity.
- ***M. haemolytica* research community:** Focused too much on LKT, not enough on other virulence factors, or the bacterial/host interaction.
- **Animal health companies:** *M. haemolytica* vaccines not improved beyond those from early 1990s.

Acknowledgements

The OSU BRD project was started by R.J. Panciera & R.E. Corstvet ~1970. Many good faculty and staff have kept it alive. Most recently:

- Sahlu Ayalew
- Mady Dabo
- Robert Fulton
- Marie Montelongo
- Jerry Ritchey
- Tim Snider
- D. L. Step
- Jared Taylor

Funding – Grants from: USDA AFRI, Noble Foundation, Oklahoma Agricultural Experiment Station, & several animal health companies



Thank you.

**Are there any
Questions?**

