

**EPIGENETIC ALTERATIONS INDUCED BY GENOTOXIC
OCCUPATIONAL AND ENVIRONMENTAL HUMAN CHEMICAL
CARCINOGENS: AN UPDATE OF A SYSTEMATIC LITERATURE
REVIEW**

A Thesis

by

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ABSTRACT

Epigenetic alterations, such as changes in DNA methylation, histones/chromatin structure, nucleosome positioning, and expression of non-coding RNAs, are recognized among key characteristics of carcinogens; they may occur independently or concomitantly with genotoxic effects. While data on genotoxicity are collected through standardized guideline tests, data collected on epigenetic effects is far less systematic. In 2016, we conducted a systematic review of published studies of genotoxic carcinogens that reported epigenetic endpoints to better understand the evidence for epigenetic alterations of human carcinogens, and the potential association with genotoxic endpoints. Since then, the number of studies of epigenetic effects of chemicals has nearly doubled. This project stands as an update on epigenetic alterations induced by genotoxic occupational and environmental human chemical carcinogens that were previously and recently classified as Group 1 by the International Agency for Research on Cancer. We found that the evidence of epigenetic effects remains uneven across agents. Studies of DNA methylation are most abundant, while reports concerning effects on non-coding RNA have increased over the past 5 years. By contrast, mechanistic toxicology studies of histone modifications and chromatin state alterations remain few. We found that most publications of epigenetic effects of genotoxic carcinogens were studies in exposed humans or human cells. Studies in rodents represent the second most common species used for epigenetic studies in toxicology; *in vivo* exposures being the most predominant. Future studies should incorporate dose- and time-dependent study designs and also investigate the persistence

of effects following cessation of exposure, considering the dynamic nature of most epigenetic alterations.

DEDICATION

I would like to dedicate this work to my parents, Ivey and Pam Goodman. For me to pursue my passions, you have both sacrificed an immeasurable amount. I am grateful every day to have been raised by loving parents who never stopped pushing me towards my dreams.

I love you to the moon and back, Mom and Dad.

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Contributors

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Previous literature review referenced within this document was completed by Dr. Grace Chappell with the assistance of Drs. Igor Pogribny and Kate Guyton and published in 2016. This work, including study categorization, summarization, and statistical analysis, was completed by the student independently.

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NOMENCLATURE

1-OHP	1-hydroxypyrene
5caC	5-carboxycytosine
5fC	5-formylcytosine
5hmC	5-hydroxymethylcytosine
5mC	5-methylcytosine
ACLAM	Activated leukocyte cell adhesion molecule
AFB1	Aflatoxin B1
AFB2	Aflatoxin B2
AhRR	Aryl-hydrocarbon receptor repressor
BER	Base excision repair
C/EBP α	CCAAT/enhancer-binding protein alpha promoter
CBMN	Cytokinesis-block micronucleus
CC	Collaborative Cross
CH3	Methyl group
ChIP	Chromatin immunoprecipitation
COX-2	Cyclooxygenase-2
CpG	Cytosine-guanine dinucleotides
CREB	cAMP-response element-binding protein
CTPE	Coal tar pitch extract
CYPs	Cytochrome P450s

DNA	Deoxyribose nucleic acid
DNMT	DNA methyltransferases
EGCG	Epigallocatechin-3-gallate
HAWC	Health Assessment Workspace Collaborative
HBEC	Human bronchial endothelial cells
HCC	Hepatocellular carcinoma
HDAC	Histone deacetylase
HUVECS	Human umbilical vein endothelial cells
IARC	International Agency for the Research on Cancer
IL-6	Interleukin 6
ISO	International Organization for Standardization
JMJD2b	Jumonji domain-contain protein 2b
LINE-1	Long interspersed nuclear element-1
lncRNA	Long non-coding RNA
MGMT	O6-methylguanine-DNA methyl-transferase
miRNA	MicroRNA
MOCA	4,4'-Methylenebis(2-chloroaniline)
MRL	Murphy Roths Large
mRNA	Messenger RNA
NF-kB	Nuclear factor kappa-light-chain-enhancer of activated B cells
nt	Nucleotides
PAHs	Polycyclic aromatic hydrocarbons

PARG	Poly(ADP-ribose) glycohydrolase
PARP-1	Poly(ADP-ribose) polymerase-1
PCBDs	Polychlorinated dibenzodioxins
PCBs	Polychlorinated biphenyls
PCR	Polymerase chain reaction
REST	Repressor element 1-silencing transcription factor
RNA	Ribose nucleic acid
ROS	Reactive oxidative species
SAEG	Small human airway epithelial cells
SAHH	S-adenosylhomocysteine hydrolase
SINE B1	Short Interspersed Nuclear Element B1
SMAD	Fusion of Sma genes and the Drosophila Mad, Mothers against decapentaplegic
SNP	Single nucleotide polymorphism
SVOCs	Semi-volatile organic compounds
TCE	Trichloroethylene
TDG	Thymine DNA glycosylase
TET	Ten eleven translocation enzymes
Topo II α	Topoisomerase II α
VOCs	Volatile organic solvents

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1. INTRODUCTION

Evaluation of the causative linkages between exposures to environmental agents and cancer includes considerations of the mechanistic evidence; the process of cancer hazard evaluation is now increasingly relying on the evidence from studies other than epidemiology or rodent cancer bioassays [1]. The practice of cancer hazard assessments and incorporation of mechanistic evidence has been streamlined and systematized through the introduction of the key characteristics of carcinogens organizational approach [2, 3]. The key characteristics approach provides guidance and structure for the consistent utilization of the mechanistic evidence and has been developed for many environmental agent-associated toxicities beyond cancer [4-8]. Included among the ten key characteristics of carcinogens is “induces epigenetic effects” and “is genotoxic.” These two key characteristics represent one of the least and one of the most data-rich key characteristics across the peer-reviewed literature related to human carcinogens, respectively, and are the focus of the project presented herein.

While many known carcinogens are genotoxic, others cause epigenetic alterations [9]; however, most studies consider one or the other even though the linkages between these key characteristics are many [10]. Epigenetic modifications are heritable non-mutational events. They can operate independently or interdependently with genetic alterations to cause dysregulation of gene expression and subsequent molecular changes [11, 12]. Epigenetic events include stable changes in gene expression that are not attributed to the modification of the genomic sequence. These include changes in DNA

methylation patterns, histone modifications, alteration of chromatin structure, and dysregulation of non-coding RNA expression. Environmental factors can impact these epigenetic marks, causing phenotypic variation and may potentiate human disease including cancer [13-15].

Studies of environmental agents suggest that epigenetic alterations play an important role in chemically-induced carcinogenesis [15, 16]. As compared to the decades of studies of genotoxicity of chemical agents and exposure-related gene expression responses, studies of epigenetics have been more recently enabled by the advances in sequencing technology. The evidence for epigenetic alterations caused by carcinogens, and the potential association with genotoxic endpoints, is rapidly growing [10]. As recently as five years ago, a systematic literature review of epigenetic alterations induced by genotoxic occupational and environmental human chemical carcinogens showed that the database was relatively sparse for environmental chemicals [10]. Only three known human chemical carcinogens (aflatoxins, benzene and benzo[a]pyrene), had ten or more studies with reported epigenetic effects. Other chemical carcinogens had very few studies of this key characteristic. As epigenetics continues to be an area of focus for mechanistic studies in toxicology, the evidentiary basis for the role of these non-genetic changes to DNA and transcription accumulates. This project summarizes the current status of the evidence on the role epigenetic alterations play in the induction of carcinogenesis following exposure to DNA damaging chemical agents (and related occupations) to enable a better understanding of the interplay of epigenetics and DNA damage in chemical carcinogenesis.

2. METHODOLOGY

A multi-step, systematic literature review was conducted to analyze published scientific evidence concerning epigenetic mechanisms of action of human carcinogens that are genotoxic. An update of the recent (since 2016) publications on environmental or occupational hazards classified as Group 1 (“carcinogenic to humans”) by the International Agency for Research on Cancer (IARC) in Monograph Volume 100F was conducted. In addition, carcinogens classified as Group 1 in IARC Monograph Volumes 105-118 were evaluated. The list of substances included in this analysis and their associated IARC monograph reference is provided in **Table 1**. In summary, substances included in this analysis were (i) classified as IARC Group 1 “human carcinogen”, (ii) identified by IARC Monographs working groups having strong evidence for a genotoxic mechanism of carcinogenesis, and (iii) had published reports of epigenetic alterations.

The systematic literature review was conducted using the Health Assessment Workspace Collaborative (HAWC) web-based tool [17] and was based exclusively on the publications indexed in PubMed. The search terms published in the previous literature review of epigenetic alterations induced by genotoxic occupational and environmental human chemical carcinogens [10] were used; a list of the search terms used for this assessment is listed in **Supplementary Table 1** (Appendix A). Inclusion and exclusion criteria are summarized in **Supplementary Table 2** (Appendix A). Included studies were classified by, in order: chemical, species (human, mouse, rat, or other), and study type (*in*

vitro or *in vivo*). The final organization of included studies was based on the type of epigenetic alterations in DNA methylation, histone modifications, or non-coding RNA.

Studies that were excluded from the final analyses were grouped by topics or areas of knowledge that were not related to the focus of this project, or not clearly defined. *In silico* studies were excluded even if they reported epigenetic effects of a chemical of interest. Studies were classified as “not chemical of interest” if the studied agents or substances were not listed in **Table 1**. Publications that were not available in full text, or were a review/commentary, were excluded. Publications that did report on a chemical of interest but did not report epigenomic alterations were placed within the "No epigenetics" category. A sub-category within this category included “ γ -H2AX”, which is an initial cellular response to DNA double-strand break induction that is technically an epigenetic alteration (histone phosphorylation), if this endpoint was used solely as an indicator of DNA damage and the study did not otherwise report on epigenetic alterations.

Dates of the literature search for the IARC Monograph 100F carcinogens ranged from 12/2015 to 12/2020, to capture literature published since the previous literature review of epigenetic alterations related to genotoxic Group 1 agents [10]. Of the 526 articles identified in the search, 391 (74%) were excluded because they did not report epigenetic effects. Other reasons for exclusion included, but were not limited to, the following: research did not pertain to the chemicals of interest, unavailability of full text in English, or no reporting of the primary data. The assessment concerning the Group 1 genotoxic carcinogens included in IARC Monographs 105-118 identified studies published from 01/1968 to 12/2020. A total of 903 scientific reports were identified in the

search, with 300 meeting inclusion standards (603 reports were excluded for prior stated reasons).

Visualizations of the literature tree demonstrating the previously described inclusion and exclusion criteria for Monograph 100F and Monographs 105-118 Group 1 carcinogens with evidence of genotoxicity are shown in **Figures 1** and **2**, respectively. The systematic literature review information is public and freely accessible. The assessment for the update to Monograph 100F carcinogens is available at: <https://hawcproject.org/assessment/1046/>. The assessment for Monographs 105-118 carcinogens is available at: <https://hawcproject.org/assessment/1063/>. While this project aimed to cite most of the studies identified in the systematic literature review, not all of them are cited and the reader is welcome to explore the literature trees at the links above. The categorization of all studies is shown within **Table 2**.

Table 1. Carcinogens classified in Group 1 in IARC Monographs Volume 100F-118 that were included in the systematic literature review.

Group 1 Carcinogen	Last Monograph Number	Evidence of Genotoxicity: Animals	Evidence of Genotoxicity: Humans	Epigenetic data in last monograph	Epigenetic data in this review
1,3-Butadiene	100F	Yes	Yes	Yes	Yes
4,4'-Methylenebis (2-Chlorobensenamine)	100F	Yes	Yes	Yes	Yes
4-Aminobiphenyl	100F	Yes	Yes	Yes	Yes
Aflatoxins (naturally occurring mixtures)	100F	Yes	Yes	Yes	Yes
Benzene	100F	Yes	Yes	Yes	Yes
Benzidine	100F	Yes	Yes	Yes	Yes
Benzo[a]pyrene	100F	Yes	Yes	Yes	Yes
Coal tar pitch	100F	Yes	Yes	No	Yes
Coke production, occupational exposure	100F	Yes	Yes	Yes	Yes
Engine exhaust, diesel	105	Yes	Yes	No	Yes
Formaldehyde	100F	Yes	Yes	Yes	Yes
Lindane	113	Yes	Yes	No	Yes
Occupational exposure as a painter	100F	No	Yes	No	Yes
Outdoor air pollution	109	Yes	Yes	Yes	Yes
Pentachlorophenol	117	No	Yes	Yes	Yes
Polychlorinated biphenyls	107	Yes	Yes	Yes	Yes
Polychlorinated biphenyls, dioxin-like	107	Yes	Yes	Yes	Yes
Sulfur mustard	100F	Yes	Yes	Yes	Yes
Trichloroethylene	106	Yes	Yes	Yes	Yes
Vinyl chloride	100F	Yes	Yes	Yes	Yes
Welding fumes	118	Yes	Yes	Yes	Yes
1,2-Dichloropropane	110	Yes	Yes	No	No
2-Naphthylamine	100F	Yes	Yes	No	No
Bis(chloromethyl)ether and chloromethyl methyl ether	100F	No	Yes	No	No
Coal gasification	100F	Yes	No	No	No
Ethylene oxide	100F	Yes	Yes	No	No
Isopropyl alcohol manufacture by the strong-acid process	100F	No	Yes	No	No
Mineral oils, untreated or mildly treated	100F	No	Yes	No	No
Mists from strong inorganic acids	100F	No	Yes	No	No
Occupation exposures during coal-tar distillation	100F	Yes	Yes	No	No

Table 1 Continued

Group 1 Carcinogen	Last Monograph Number	Evidence of Genotoxicity: Animals	Evidence of Genotoxicity: Humans	Epigenetic data in last monograph	Epigenetic data in this review
Occupational exposure during aluminum production	100F	No	Yes	No	No
Occupational exposures during iron and steel founding	100F	No	Yes	No	No
Occupational exposures in the rubber manufacturing industry	100F	No	Yes	No	No
Ortho-toluidine	100F	Yes	Yes	No	No
Shale oils	100F	Yes	No	No	No
Soot, as found in the occupation exposure of chimney-sweeps	100F	No	Yes	No	No

Table 2. Number of publications with data related to epigenetic alterations for each Group 1 carcinogen included in the systematic literature review.

Group 1 carcinogen	Last Monogr. Listed		DNA Methylation			Histone Modifications			Non-Coding RNAs			Total
			Humans	Rodents (mouse or rat)	Other (See footnotes)	Humans	Rodents (mouse or rat)	Other (See footnotes)	Humans	Rodents (mouse or rat)	Other (See footnotes)	
1,3-Butadiene	100F	<i>in vitro</i>										0
		<i>in vivo</i>		5			5			1		11
4-Aminobiphenyl	100F	<i>in vitro</i>				1			2			3
		<i>in vivo</i>										0
Aflatoxins (naturally occurring mixtures)	100F	<i>in vitro</i>	8		1 ⁿ , 2 ^o	1		1 ^o	8	1		22
		<i>in vivo</i>	11	4	1 ^r		1	1 ^r	4	8	1 ^c , 1 ^p	32
Benzene	100F	<i>in vitro</i>	11	2		3	1		10	1		28
		<i>in vivo</i>	23	2		5	1		12	2		45
Benzidine	100F	<i>in vitro</i>										0
		<i>in vivo</i>		1								1
Benzo[a]pyrene	100F	<i>in vitro</i>	18	6	3 ⁿ , 2 ⁱ	15	2	1 ^h	26	4		77
		<i>in vivo</i>	11	12	6 ^w , 2 ^m , 1 ^g , 1 ^r , 1 ^s		3		2	13	1 ^g , 1 ^j , 1 ^t , 1 ^k , 1 ^f	57
Coal-tar pitch	100F	<i>in vitro</i>	1						2			3
		<i>in vivo</i>										0
Coke production, occupational exposure	100F	<i>in vitro</i>	1									1
		<i>in vivo</i>	10			3			4			14
1,2-Dichloro-propane	110	<i>in vitro</i>										0
		<i>in vivo</i>										0

Table 2 Continued

Group 1 carcinogen	Last Monogr. Listed		DNA Methylation			Histone Modifications			Non-Coding RNAs			Total
			Humans	Rodents (mouse or rat)	Other (See footnotes)	Humans	Rodents (mouse or rat)	Other (See footnotes)	Humans	Rodents (mouse or rat)	Other (See footnotes)	
Engine exhaust, diesel	105	<i>in vitro</i>	2			2			5			9
		<i>in vivo</i>	6	5					3			14
Formaldehyde	100F	<i>in vitro</i>	1			3		1 ^v	1			7
		<i>in vivo</i>	1							3	1 ^d	5
Lindane	113	<i>in vitro</i>										0
		<i>in vivo</i>		1								1
4-4'-Methylenebis (2-chloro-benzenamine)	100F	<i>in vitro</i>					1					1
		<i>in vivo</i>										0
Occupational exposure as a painter	100F	<i>in vitro</i>										0
		<i>in vivo</i>	1						1			2
Outdoor air pollution	109	<i>in vitro</i>	7			3			16			26
		<i>in vivo</i>	83	12	1 ^b , 1 ^e	4	3		26	12	1 ^w	143
Pentachlorophenol	117	<i>in vitro</i>										0
		<i>in vivo</i>								1		1
Polychlorinated biphenyls (non-dioxin-like)	107	<i>in vitro</i>	3	1	1 ^u	1	1		2	1	2 ^c	12
		<i>in vivo</i>	19	11	1 ^l , 1 ^q , 1 ^a , 1 ^m		1		2	3	2 ^w	42
Polychlorinated biphenyls (dioxin-like)	107	<i>in vitro</i>	1			2	1		1			6
		<i>in vivo</i>	3	3	1 ^w				2			8

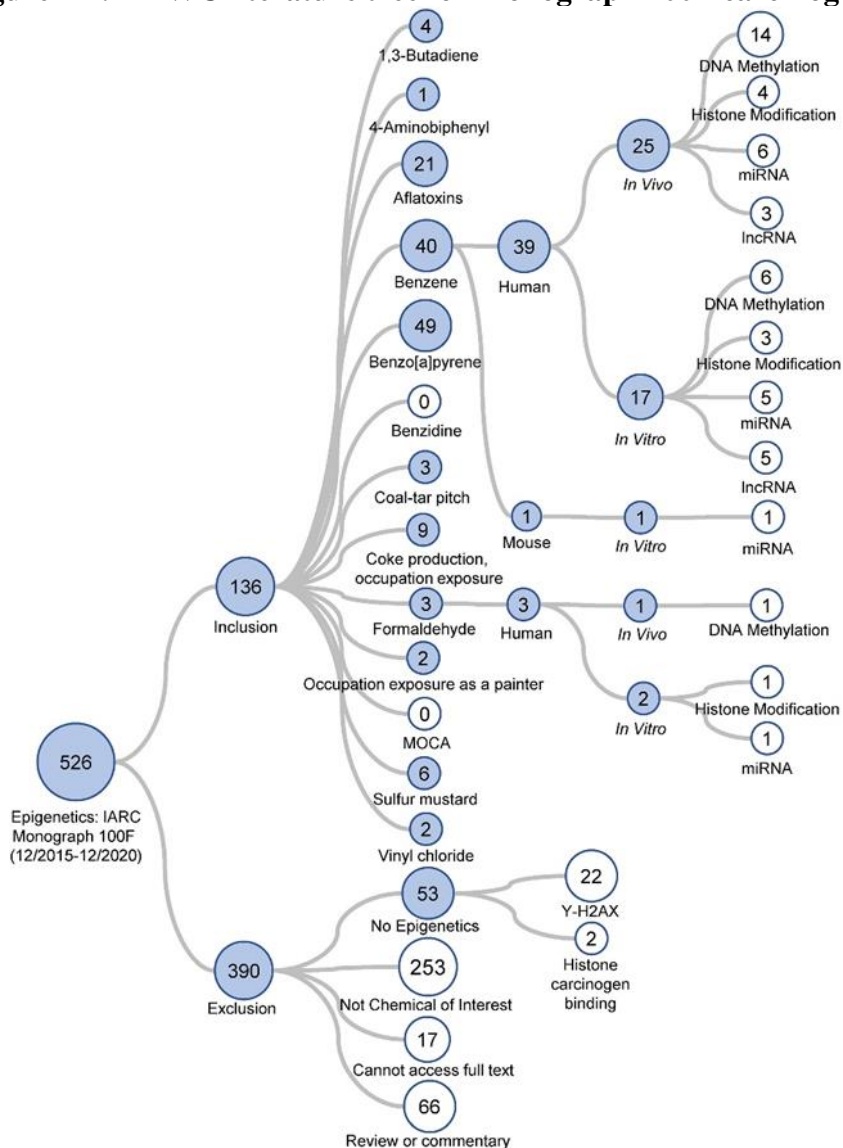
Table 2 Continued

Group 1 carcinogen	Last Monogr. Listed		DNA Methylation			Histone Modifications			Non-Coding RNAs			Total
			Humans	Rodents (mouse or rat)	Other (See footnotes)	Humans	Rodents (mouse or rat)	Other (See footnotes)	Humans	Rodents (mouse or rat)	Other (See footnotes)	
Sulfur mustard	100F	<i>in vitro</i>	2			1			2			5
		<i>in vivo</i>							6	2		8
Trichloroethylene	106	<i>in vitro</i>	8	2		4			2	2	2 ^e	20
		<i>in vivo</i>	1	18			2		1	4	1 ^w	27
Vinyl chloride	100F	<i>in vitro</i>										0
		<i>in vivo</i>	3	1					1			5
Welding fumes	118	<i>in vitro</i>	1	1								2
		<i>in vivo</i>	3	2					1			6

- a Alaska blackfish (*Dallia pectoralis*)
- b Barn swallow (*Hirundo rustica*)
- b Chicken (*Gallus gallus domesticus*)
- d Cynomolgus macaques (*Macaca fascicularis*)
- e Dog (*Canis lupis familiaris*)
- f Firefly (*Coleoptera lampyridea*)
- g Fish (*Oeyzias melastigma*)
- h Frog (*Xenopus borealis*)
- i Hamster (*Mesocricetus auratus*)
- j Japanese Rice Fish (*Oryzias latipes*)
- k Lancelet (*Brachiostoma belcheri*)
- l Loggerhead sea turtle (*Caretta caretta*)

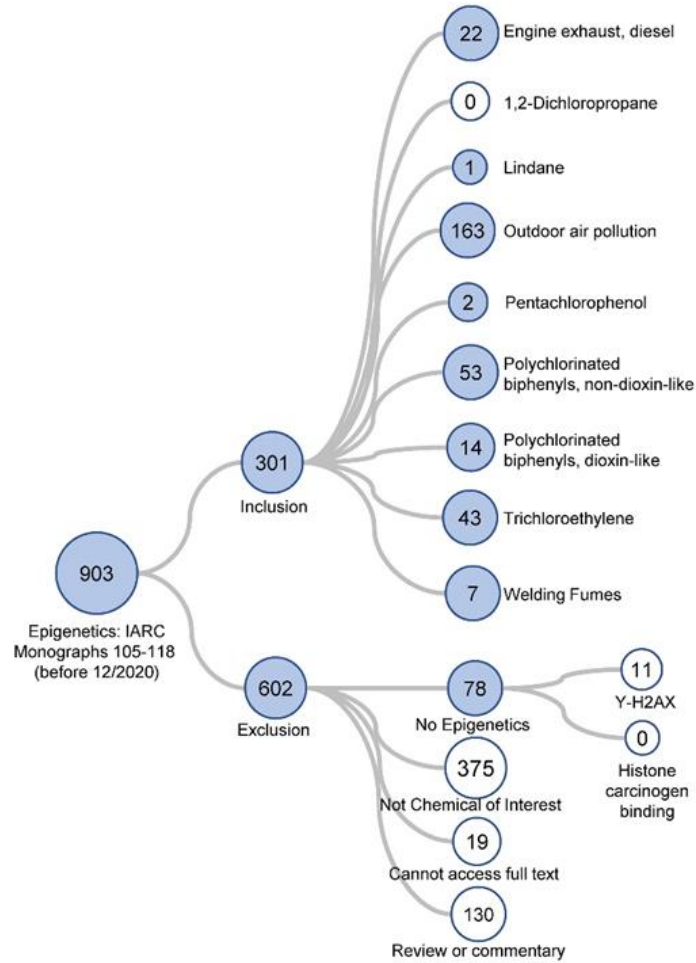
- m Mumichog fish (*Fundulus heteroclitus*)
- n Oligodeozynucleotides
- o Pig (*Sus domesticus*)
- p Pig (*Sus scrofa domesticus*)
- q Pond Slider (*Trachemys scripta*)
- r Rainbow trout (*Oncorhynchus mykiss*)
- s Rock dove (*Columba livia*)
- t Roundworm (*Caenorhabditis elegans*)
- u Sheep (*Ovis aries*)
- v Synthetic peptide
- w Zebrafish (*Danio rerio*)

Figure 2-1. HAWC literature tree for Monograph 100F carcinogens.



Literature tree of the 526 studies returned from the PubMed search concerning epigenetic endpoints for the human carcinogens included in IARC Monograph 100F (all classified as “Group 1”). The studies were categorized based on inclusion or exclusion criteria as shown. Included studies were further sorted, in order, by cancer hazard, species studied (human, rat, mouse, or other), study design (in vivo or in vitro), and epigenetic endpoint. The numbers within each circle denote how many studies were allocated to that specific category. Blue circles denote that they contain additional sub-categories. White circles indicate a terminal category. The literature tree branches for included studies for carcinogens “Benzene” and “Formaldehyde” are expanded for illustration purposes. The branch for “No Epigenetics” category is shown as an example of an exclusion category. The searches covered literature published between 12/2015 and 12/2020

Figure 2-2. HAWC literature tree for Monographs 105-118 carcinogens.



Literature tree of the 903 studies returned from the PubMed search concerning the human genotoxic carcinogens included in IARC Monographs 105-118 (all classified as “Group 1”) with evaluation of epigenetic alterations. The studies were categorized based on inclusion or exclusion criteria as shown. See Figure 1 legend for further explanation of the figure structure. The searches covered literature published before 12/2020.

3. CATEGORIES OF EPIGENETIC ALTERATIONS INDUCED BY THE CHEMICALS AND ASSOCIATED OCCUPATIONS INCLUDED IN THE SYSTEMATIC LITERATURE REVIEW

3.1. DNA Methylation

DNA methylation (**Figure 3**) is the covalent addition of a methyl group (CH₃) at the carbon-5 position of cytosine, resulting in 5-methylcytosine (5mC). DNA methylation is typically found at cytosine-guanine dinucleotides (CpG sites) and is catalyzed by *de novo* DNA methyltransferases (DNMT3A and DNMT3B) which transfer a methyl group from *S*-adenyl methionine (SAM) to the carbon-5 location of the cytosine [18]. DNMT1, a maintenance DNMT, is responsible for maintaining the DNA methylation patterns following replication. DNA methylation is a dynamic, multi-step process that is counter-balanced by DNA demethylation. DNA demethylation, or the removal of methyl groups from cytosines, can occur through passive (spontaneous de-methylation) or active mechanisms. The latter involves the oxidation of 5mC by ten-eleven-translocation (TET) proteins, creating 5-hydroxymethylcytosine (5hmC), 5-formylcytosine (5fC), and 5-carboxycytosine (5caC) through hydroxylase activity [19]. Thymine DNA glycosylase (TDG) via the base excision repair (BER) pathway can excise 5hmC, 5fC, and 5caC which will be replaced with an unaltered cytosine base [20].

DNA methylation is a major epigenetic factor and has been implicated in the modulation of cell biology [21]; gene expression regulation, genomic stability, and overall chromosomal inactivation [22, 23]; and silencing of imprinted genes [24]. Aberrant DNA

methylation results in malignant tumors through DNA hyper- or hypo-methylation [25], with hypermethylation indicating increased methylation with subsequent transcriptional suppression and decreased gene expression [26, 27] and hypomethylation indicating decreased methylation with influence over chromosomal stability [28]. Global DNA hypomethylation is normally utilized as a biomarker of cancer induction as it causes genomic instability [25], a hallmark of carcinogens [2].

Previous research concerning genomic DNA methylation and identification of DNA-methylation-based biomarkers for disease diagnosis has focused on hypermethylation of promoter CpG islands within tumor-suppressor genes [29, 30]. CpG islands represent clusters of CpG sites; when hypermethylated, expression is repressed. Recent studies focus on identifying the CpG loci as biomarkers. For example, methylation of specific locations within a promoter has been found to control transcriptional silencing, in contrast to indiscriminate methylation throughout the entire promoter region [31-35]. Quantification of DNA methylation can be achieved through various techniques, most recent techniques are based on advancements in nucleotide sequencing [36]. Non-sequencing methods include mass spectrometry for global DNA methylation levels or combined bisulfite restriction analysis for gene-specific DNA methylation, warrant consideration [37-39].

3.2. Histone Modifications

Post-translational histone modifications (**Figure 3**) comprise several epigenetic marks that have been associated with transcription, DNA repair, and the maintenance of chromatin structure [40]. Nucleosomes, the fundamental building blocks of chromatin, are

composed of octamers of the four core histone proteins (H3, H4, H2A, and H2B) around which is wrapped with a 147 base-pair segment of DNA. Each histone protein possesses an N-terminal tail (a side chain) populated with cationic lysine and arginine residues; these side chains may be post-translationally modified by acetylation, citrullination, methylation, phosphorylation, or ubiquitination. Histone modification “writers” (e.g. histone acetyltransferases) enzymatically add alterations while “erasers” (e.g. histone deacetylases) enzymatically remove alterations [41], representing a dynamic system modulating these alterations on the histone side chains. These histone modifications are then recognized by "reader" proteins (e.g. bromodomains and chromodomains) which affect transcription.

Most studies of histone modifications covered in this project focus on acetylation and methylation of lysine residues. Histone acetylation modifies the ϵ -amino group of lysine residues, neutralizing the positive charge. This alteration has been suggested to disrupt the interaction between the histone side chain and DNA nucleosome, resulting in increased chromatin availability and the promotion of active transcription. Acetylated lysine further promotes chromatin accessibility by attaching to bromodomain-containing transcription factors within chromatin remodeling complexes [42, 43]. Histone methylation has been identified as a key mechanism in development and differentiation [44, 45]. Histone methylation occurs on both lysine and arginine residues with preferential occurrence on histone H3 and H4 [46]. Various methylation states exist (mono-, di-, and trimethylation) and all can elicit different transcriptional outcomes. Recent studies investigate histone modifications through ADP-ribosylation, butyrylation, citrullination,

crotonylation, deamination, formylation, O-GlcNAcylation, propionylation, and proline isomerization [40]. Dysregulation of histone modifications can alter gene expression and therefore has been associated with carcinogenesis [47, 48]. Histone modifications may be involved in carcinogenesis through transcriptional [49] and DNA repair mechanisms [50, 51]. Alterations of histone post-translational modification patterns are most frequently studied either globally or at specific loci using ChIP (chromatin immunoprecipitation) followed by sequencing [40].

3.3. Non-Coding RNAs

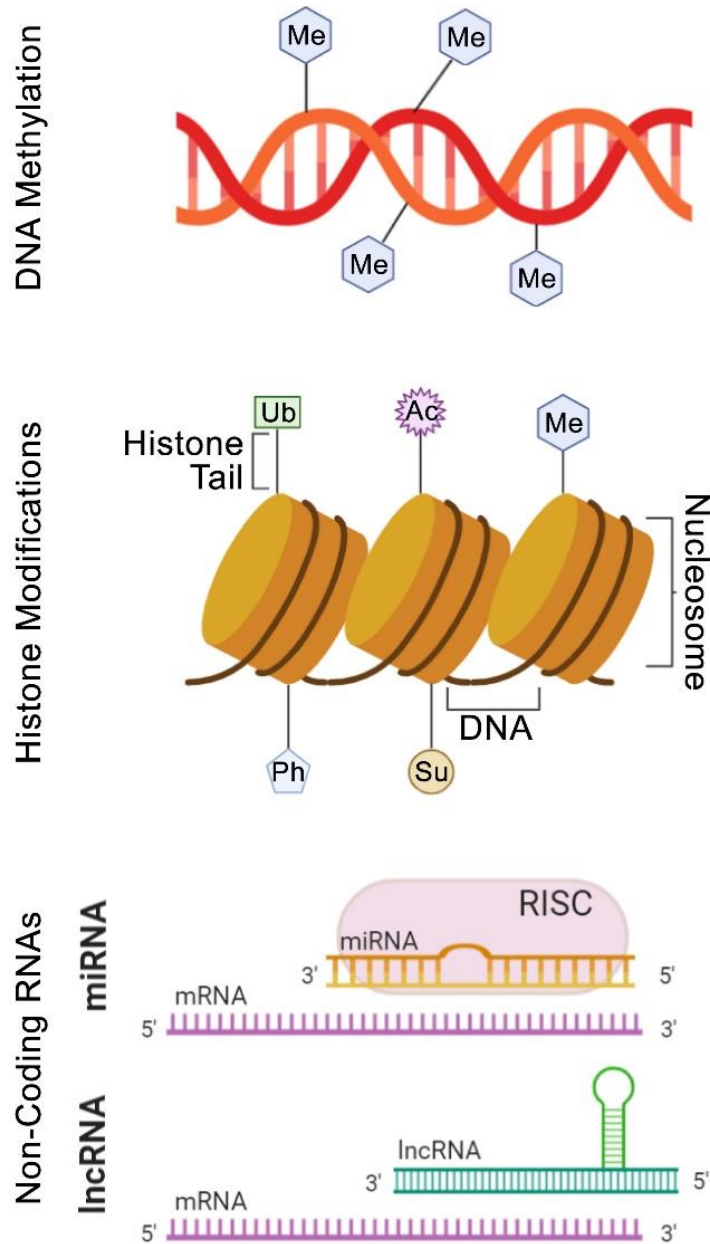
Non-coding RNAs (**Figure 3**) are a class of RNA molecules that are not translated into a protein. Non-coding RNAs that possess a regulatory role in transcription are typically sub-divided into two categories: short-chain non-coding RNAs (miRNA) and long non-coding RNAs (lncRNA). These non-encoding RNAs have been shown to have significant roles in epigenetic modifications and regulation on both the gene and chromosomal level [52-54]. Quantitative RT-PCR, RNA sequencing, and microarrays are the most commonly utilized methods for quantitation of non-coding RNAs.

miRNA are short, single-stranded RNA (~19-25 nt) that originate from the DNA regions prone to structural changes (“fragile sites”); these loci are frequently amplified or deleted in tumor cells [55, 56]. miRNA play important roles in cell proliferation and apoptosis by controlling the expression of tumor suppressor genes and oncogenes. This regulation is achieved primarily through the binding of miRNA to the 3’ untranslated region of mRNA inducing their degradation or blocking translation [57, 58]. Biogenesis of miRNA is a complex process [59]; mutations in proteins involved in miRNA synthesis

may result in alterations of miRNA processing, stability, and targeting, causing subsequent biological insults, including carcinogenesis [56].

Long non-coding RNA are transcripts longer than 200 nt and represent one of the largest and most diverse RNA families [60]. LncRNA regulate gene expression through a variety of mechanisms, including somatic mutation and altered transcriptional regulation [61, 62], and aberrant expression has been significantly linked to cancer carcinogenesis and progression [63, 64]. LncRNA may act as scaffolds for various epigenetic endpoints, inducing a cascade of events. For example, lncRNA may induce histone modifications by linking proteins together to form ribonucleoprotein complexes [62] and interact with DNA methyltransferase enzymes to affect DNA methylation [65].

Figure 3-1. Graphic of epigenetic endpoints.



A general graphic summary of the epigenetic alterations that were searched for in this review. Abbreviations: Ac – acetylation; Ph – phosphorylation; Me – methylation; Ub – ubiquitination; Su – sumoylation; mRNA – messenger RNA; miRNA – micro RNA; RISC – RNA induced silencing complex; lncRNA – long non-coding RNA.

4. EPIGENETIC EFFECTS ASSOCIATED WITH IARC GROUP 1 HUMAN CARCINOGENS INCLUDED IN IARC MONOGRAPH 100F

A total of 11 human genotoxic carcinogens that met inclusion criteria for this systematic literature review are discussed herein. A description of the common routes of exposure, associated cancers, and genotoxicity of each of these, except for coal tar pitch (see section 4.10), was also evaluated previously [10].

4.1. Benzo[a]pyrene

4.1.1. DNA Methylation.

As detailed in Chappell et al [10], changes in DNA methylation induced by benzo[a]pyrene exposure were primarily studied using human and rodent in vitro models. Modifications of the methylome were found to be gene-specific (e.g. CDH13 and p16). Since 2016, most studies concerning benzo[a]pyrene exposure and DNA methylation focused on human in vitro and in vivo models. Recent studies provide additional evidence for gene-specific methylation (e.g. tumor suppressor genes) and identified biomarkers to aid in diagnosis and prognosis of benzo[a]pyrene-associated cancer. For example, exposure to benzo[a]pyrene via cigarette smoke was significantly associated with increased promoter hypermethylation of p16 and DAPK (genes involved in carcinogenesis pathways), as well as with DNA damage [66]. Furthermore, benzo[a]pyrene increased DNA methyltransferase 1 (an enzyme essential for methylated metabolite accumulation) in smokers diagnosed with bladder cancer; in vitro validation

of these results showed dysregulated metabolite levels may be used as potential predictive biomarkers for benzo[a]pyrene associated carcinogenesis [67]. Several studies have proposed additional methylated genes as biomarkers of benzo[a]pyrene-associated cancer [68, 69] with changes to the global methylome being associated with the development and progression of cancer [68, 70]. Benzo[a]pyrene exposure in human lymphocytes induced aberrant expression of DNMT1 and hypomethylation of nuclear and mitochondrial DNA, initiating histone methylation and modifying chromatin structure; this cascade of epigenetic events may provide insight into the mechanisms of benzo[a]pyrene associated toxicity [71].

4.1.2. Histone Modifications.

In publications reviewed by Chappell et al [10], most studies of histone modifications by benzo[a]pyrene were conducted in human cell lines. The most frequent type of the reported effects was changes in global post-translational modifications of various histones (e.g., H3K4me3 and H3K9ac) as a result of treatment. Since 2016, most of the studies of benzo[a]pyrene and histone modifications also used human cells as a model of choice; however, the emphasis is more on the mechanisms of the benzo[a]pyrene effects on the histones and chromatin, and the utility of histone modifications as biomarkers of exposure to benzo[a]pyrene. For example, hyperacetylation of histones H3 and H4 and upregulation of histone deacetylases (HDAC2 and HDAC3) were found to be associated with global loss of DNA methylation following benzo[a]pyrene-exposure of human bronchial epithelial (16HBE) cells; these effects were concentration-dependent [70]. The effect of benzo[a]pyrene on the

chromatin states indicative of reactivation of long interspersed nuclear element-1 (LINE-1) retrotransposon was shown to involve functional modulation of nucleosomal and remodeling deacetylase NuRD, leading to oncogenic transformation of human bronchial epithelial cells (HBECs) [72]. Several studies showed that poly(ADP-ribose) glycohydrolase (PARG) may also play an important role in benzo[a]pyrene-associated changes in histone H2 expression; these studies were conducted in both human lung cancer cell lines and a mouse model of benzo[a]pyrene-induced carcinogenesis [73, 74]. Downregulation of *PARG* and histone modifications similar to those in human lung cancers were found in benzo[a]pyrene-treated human lung cancer cell lines suggesting that *PARG* silencing may be a biomarker for benzo[a]pyrene exposure-related lung carcinogenesis [74].

4.1.3. Non-Coding RNAs

Formerly reviewed [10] publications that investigated effects of benzo[a]pyrene on the expression of non-coding RNAs utilized human *in vitro* models; these studies demonstrated the role of aberrant miRNA expression in the dysregulation of DNA repair mechanisms and tumor suppressor genes. Since 2016, the majority of studies also used human cells, with focus on the role of miRNAs in non-genotoxic mechanisms of benzo[a]pyrene-associated carcinogenesis, such as inflammation. For example, benzo[a]pyrene-treated human bronchial epithelial cells (HBECs) demonstrated differential expression of specific miRNAs was significantly associated with the upregulation of activated leukocyte cell adhesion molecule (ALCAM) [75]. *In vitro* and *in vivo* (tumor model within nude mice) studies using *ALCAM*-knockout HBECs showed

inhibition of colony formation and increased cell migration and decreased tumor growth and enhanced metastasis, respectively. Benzo[a]pyrene exposure within human mammary cells resulted in the upregulation of inflammation pathways and dysregulation of tumorigenic miRNAs, progressing benzo[a]pyrene-induced tumorigenesis through non-mutagenic pathways [76]. The influence of benzo[a]pyrene-induced inflammation was further explored within human adenocarcinoma mammary cells cotreated with benzo[a]pyrene and a pro-inflammatory cytokine IL-6 [77]; genotoxicity was enhanced due to the downregulation of miRNA-27b, allowing increased CYP1B1 metabolism of benzo[a]pyrene to reactive intermediates. Several studies have shown *in vitro* benzo[a]pyrene exposure caused prominent changes in the expression of miRNAs and their respective target genes, providing mechanistic insight into benzo[a]pyrene-associated toxicity [71, 78].

In addition to the focus on miRNA, recent studies investigated the role of lncRNA in mediating benzo[a]pyrene-associated effects. For example, the significant upregulation of linc00673 following benzo[a]pyrene exposure within human non-small cell lung cancer cells (A549) increased cell migration, invasion, and epithelial-mesenchymal transition [79]. Human lung-derived cell lines treated with benzo[a]pyrene showed increased lncRNA H19 expression, initiating the downregulation of the S-adenosylhomocysteine hydrolase (SAHH) pathway, and overall mediation of LINE-1 methylation [80].

4.2. Benzene

4.2.1. DNA Methylation.

As detailed in Chappell et al. [10], earlier studies of DNA methylation effects of benzene exposure used human *in vivo* and *in vitro* models. The research focused on gene-specific aberrant methylation, explicitly genes associated with tumor suppression and apoptosis inhibition, following benzene treatment. Since 2016, evidence concerning benzene-induced DNA methylation has been gathered from human *in vivo* and *in vitro* studies. The majority of new human *in vivo* studies focus on gene-specific methylation, the identification of potential biomarkers for early detection of benzene toxicity, and the influence of benzene on DNA methyltransferase expression. For example, peripheral blood cells derived from benzene-exposed workers demonstrated hypomethylation of *STAT3*, which was significantly correlated with oxidative stress variables [81], and *MT-COI* a gene associated with oxidative phosphorylation [82]; these epigenetic modifications represent potential predictive marks for benzene exposures. Also, it was found that low-level occupational benzene exposure was associated with hypermethylation of tumor suppressor genes *p15^{INK4b}* [83], *p14ARF*, and *p16^{INK4A}* [84]; aberrant methylation of *p16^{INK4A}* was significantly correlated with genomic instability caused by benzene-induced chromosomal abnormalities. Furthermore, occupational benzene exposure led to aberrant promoter methylation of *CSF3R* that is essential for neutrophil production [85] and *ERCC3* that is involved in nucleotide excision repair [86]; these effects were significantly associated with a reduction in neutrophils and CpG methylation-induced hematotoxicity, respectively. Prolonged benzene exposure in

workers was associated with chromosomal damage and increased frequency of micronuclei; *in vitro* validation of these results showed even low-level benzene concentrations cause hypermethylation of DNA repair genes and hypomethylation of LINE-1 within acute myeloid leukemia (AML-5) cells [87].

Specific single nucleotide polymorphisms (SNPs) for DNA methyltransferase 3A and 3B (DNMT3A/3B; *de novo* DNMTs) were positively associated with hypomethylation following benzene exposure within shoe factory workers; this effect was hypothesized by the authors to have influenced the induction of DNA damage (e.g. micronuclei frequency) [88]. Increased expression of DNMT3b expression following benzene exposure within human *in vivo* and *in vitro* studies was correlated with increased promoter hypermethylation, silencing HOXA transcript antisense RNA myeloid-specific 1 (HOTAIRM1) expression, a putative tumor suppressor gene [89]. The DNA methylation status within erythroleukemic cells (K562) exposed to various benzene metabolites determined erythroid gene expression and cell differentiation [90-92]; for example, phenol exposure modulated the methylome to suppress erythroid-specific genes [92] while catechol upregulated gene expression [91].

4.2.2. Histone Modifications.

Previously summarized reports [10] investigating the effect of benzene exposure on histones and/or chromatin structure were conducted within human and mouse *in vivo* and rat *in vitro* models. These studies focused on species specificity of benzene-induced histone modifications and the role histone alterations play in topoisomerase II α (Topo II α) activity, expression, and mRNA levels. Since 2016, reports of histone modifications

have utilized human *in vivo* and *in vitro* models to investigate gene-specific modifications due to aberrant histone expression and the effects of histone alterations on Topo II α . For example, human bone marrow mononuclear cells treated with benzene increased histone deacetylase (HDAC) activity and decreased Topo II α activity, subsequently inducing apoptosis [93]. A follow-up full-body inhalation exposure within mice confirmed decreased Topo II α expression by impaired acetylation of histone H4 and H3. Human bone marrow samples from chronic benzene poisoning cases also showed significant decreases in histone H4 and H3 acetylation levels lowered mRNA expression of TOPOII α promoter regulation factors [93]. Recent studies of benzene exposure in workers showed amplified trimethylation of histone 3 lysine 4 (H3K4me3) was positively correlated with increased DNA damage [94]. A follow-up study within primary human lymphocytes revealed enrichment of H3K4me3 occurred within DNA damage responsive genes, potentially highlighting the mechanism in which histone modifications mediate benzene-induced hematotoxicity.

4.2.3. Non-Coding RNAs.

Studies originally summarized in Chappell et al. [10] concerning the aberrant expression of non-coding RNAs following benzene exposure used human and mouse *in vivo* models. The most common endpoints covered within these reports included differentially expressed miRNA and lncRNA-associated immune response pathways, the effects of *in utero* exposure on immune regulation, and aberrant miRNA expression-associated hematotoxicity. Since 2016, perturbed non-coding RNA expression induced by benzene exposure has been studied within human biological samples and cells. For

example, levels of circulating miRNA-221 following benzene exposure can be utilized as a reliable, minimally invasive biomarker of benzene-induced disease [95]; this finding was corroborated by several recent human *in vivo* and *in vitro* studies [96-98]. Downregulation of miRNA-451a and miRNA-486-5p was identified as being responsible for the regulation of erythroid differentiation in human cells [99, 100]. Alteration of miRNA-486-5p expression effected expression of *MAGII* (protein scaffolding gene) and *RASSF5* (tumor suppressor gene), suggesting miRNA-486-5p may be involved in Ras-associated protein-1 signaling pathway-associated genes to inhibit benzene-induced erythroid differentiation. Aberrant expression of miRNAs induced by poly(ADP-ribose)polymerase-1 (PARP-1) expression has been investigated using TK6 lymphoblastoid [101] and TK6 with miR-7-5p mimic (TK6- miR-7-5p) lines [102]. PARP-1 was found to decrease miRNA-155 expression through acetylation by modulating MBD2 protein levels [101]. An increase in miR-7-5p expression, following hydroquinone exposure, further aggravated PARP-1 and BRCA1 downregulation in TK6-miR-7-5p cells, subsequently enhancing cell apoptosis and inhibiting proliferation [102]. Investigation into the benzene metabolite 1,4-benzoquinone showed exposure-induced genotoxicity and significant upregulation of miR-222 within human lymphoblast cells [103]; this epigenetic modification inhibited MDMR-p53 expression and subsequently caused inactivation of p53 and abnormal DNA repair capacity.

Expression of lncRNAVNN3 was upregulated within exposed workers and treated human lymphocytes; a positive correlation was noted between this lncRNA and serum autophagy- and apoptosis-associated proteins [104]. Furthermore, lncRNAVNN3

was found to mediate benzene-induced autophagy and apoptosis through the mediation of beclin1 and Bcl-2 phosphorylation. Overexpression of lncRNA-OBFC2A decreased cell proliferation by interacting with Smad3 (a key protein in TGF- β signaling cascade) to control the cell cycle through altered Cyclin D1 expression [105, 106]. A human *in vivo* and *in vitro* study demonstrated crosstalk between lncRNA FAS-AS1 and DNMT3b with both being dynamically expressed via a mutual inhibition loop; the expression of FAS was downregulated through histone acetylation [107].

4.3. Aflatoxins

4.3.1. DNA Methylation.

Studies previously reviewed elsewhere [10] predominately focused on modification of the methylome following aflatoxin exposure within human *in vivo* models. The endpoints most frequently investigated included gene-specific methylation and resulting expression alterations (e.g., RASSF1A), the effect of DNA methylation on telomere length, and global methylation alterations. Since 2016, extensive research has been conducted concerning gene-specific methylation following aflatoxin exposure using human cell lines. For example, chronic aflatoxin B1 exposure produced persistent DNA hypomethylation within six identified cancer-related genes: *TXNRD1*, *PCNA*, *CCNK*, *DIAPH3*, *RAB27A*, and *HIST1H2BF* [108]. This study is the first to report persistent epigenetic effects induced following AFB1 exposure within the transcriptome and may provide key insight into potential biomarkers of hepatocellular carcinoma (HCC) development (a disease strongly associated with AFB1 exposure). Minimally

cytotoxic *in vitro* exposures of aflatoxin B1 (AFB1) and aflatoxin B2 (AFB2) (a structurally-related non-carcinogenic analog) produced distinct gene-expression profiles with significant DNA methylation alterations [109]; however, no correlation between gene-specific DNA methylation and gene expression changes was noted. In contrast, non-toxic, low-dose aflatoxin B2 exposure identified a large number of differentially expressed genes but few effects on the global DNA methylome, implying a transcriptomic response rather than significant DNA methylation modifications [110].

Additional human *in vitro* studies have shown the interplay of multiple epigenetic mechanisms pivotal to the aflatoxin-induced carcinogenesis pathway. S phase-arrested L02 cells exhibited aflatoxin-induced apoptosis, decreased mitochondrial membrane potential, increased reactive oxygen species generation, and upregulation of global DNA methylation at all exposure concentrations [111]. Results concluded DNA methylation induced by AFB1 exposure played a key role in regulating multiple hormonal and cell fate pathways. Aflatoxin B1 treatment within immortalized human cells caused *p21* promoter (a tumor suppressor gene) hypermethylation via increased DNMT3a expression, a pathway initiated by the upregulation of H3K26me3 and H2AK119Ub [112]; this cascade of epigenetic events resulted in the deregulation of cell cycle regulatory molecules. A secondary study of aflatoxin B1 treatment within human epithelial cells showed the enhancement of DNA methyltransferases (DNMT1, DNMT3A, and 3B), histone modifiers (HDAC 1, 2, 4, and 6), and histone marks (H2K27me3 and H2AK119Ub) [113]. Together these studies provide strong mechanistic evidence for epigenetic regulation of aflatoxin B1 effects.

4.3.2. Histone Modifications.

Previously reviewed publications [10] investigating aflatoxin exposure-associated histone modifications use mouse and alternative animal models. The studies investigated the correlation between histone repression and DNA hypermethylation within oocytes. Since 2016, the effects of benzene on histone marks have only been investigated within human cell lines. As summarized above, crosstalk was identified between histone modifications and genome methylation following AFB1 exposure, specifically the association between repressive histone marks (H3K27me3 and H2AK119Ub) and DNMTs [112, 113]. Chromatin immunoprecipitation analysis showed increased enrichment of H3K27me3 following AFB1 exposure upregulated DNMT3a, causing promoter demethylation (*Cyclin D1*) and hypermethylation (*p21*). These epigenetic modifications ultimately caused the deregulation of cell cycle regulatory molecules.

4.3.3. Non-Coding RNAs.

As summarized in Chappell et al. [10], previously reviewed studies investigated the effects of aflatoxin on non-coding RNA expression in human and rodent models. These studies focused on upregulated miRNA within liver tumors, identification of sensitive biomarkers of aflatoxin exposure, and miRNA-mRNA interactions associated with repression of DNA damage repair. Since 2016, human, rodent, and alternative animal models continue to be used and focused on the identification of potential biomarkers of aflatoxin exposure, differentially expressed miRNA associated with carcinogenesis pathways, and non-coding pathways enhancing immune response pathways. For example, two miRNAs have been identified as biomarkers for the early

detection of aflatoxin-associated hepatocellular carcinoma. MiRNA-4651 was identified as a potential biomarker due to its increased accuracy and sensitivity when compared to α -fetoprotein (a serum marker to identify patients at high risk of HCC) [114].

Upregulation of miRNA-182 was detectable months before clinical HCC symptoms were present and represent a non-invasive, serum biomarker for disease detection [115]. Utilization of miRNA-4651 and -182 in identifying small-size and early-stage HCC may improve AFB1-induced HCC disease diagnosis and prognosis.

MiRNA-155 expression in isolated spleen T cells from AFB1-exposed mice was found to be significantly reduced [116]. In the same tissue, significant upregulation of phosphatidylinositol-3, 4, 5-trisphosphate 5-phosphatase 1 (Ship1), and suppressor of cytokine signaling 1 (Socs1) was observed within exposed T cells. These outcomes suggest miR-155 and targeted proteins (Ship1 and Socs1) are involved within AFB1-induced immunotoxicity. Following AFB1 exposure, differentially expressed miRNAs were identified, all being significantly associated with cancer development [117]. Subsequent bioinformatic analyses established miRNA-mRNA regulatory networks related to the onset of AFB1-induced hepatocellular carcinoma.

A long-term exposure study in Roman hens fed AFB1-contaminated food indicated the expression levels of protein-coding genes, miRNAs, and lncRNAs within the liver were altered [118]; there was a strong association between these epigenetic effects and upregulation of *PPARG* (fatty acid storage regulator) and downregulation of *Bcl-6* (autoimmune response regulator).

4.4. Coke production

4.4.1. DNA Methylation.

As summarized in Chappell et al. [10], DNA methylation following coke production exposure was only investigated within human *in vivo* models. The endpoints studied within these publications were global and gene-specific promoter methylation, chromosomal abnormalities, and identification of urinary biomarkers for coke production exposure. Since 2016, studies have continued to primarily use human *in vivo* models, reporting retrotransposon (e.g. LINE-1) and gene-specific methylation (e.g. *CYP1A1* and *FLT1*). For example, coke oven workers with reported heavy smoking habits showed significant associations between *CYP1A1* hypomethylation and increased levels of 8-OHdG (a biomarker of oxidative DNA damage) [119]; these results indicate the co-exposure of smoking and coke oven work increases the risk for oxidative DNA damage, a key player in disease pathogenesis, through epigenetic alterations of *CYP1A1*. Urinary samples collected from coke oven workers demonstrated high levels of urinary 1-hydroxypyrene (1-OHP; a PAH metabolite) that were significantly associated with LINE-1 hypomethylation [120]; the additional exposure of tobacco use was found to increase aryl-hydrocarbon receptor repressor (*AhRR*) hypomethylation. Peripheral blood lymphocytes collected from coke oven workers possessed hypermethylation of fms related tyrosine kinase 1 (*FLT1*), a gene previously found to be hypermethylated within several human cancers [121]. The aberrant *FLT1* methylation status was positively correlated with 1-OHP urinary levels and DNA damage, indicating *FLT1* may act as a tumor suppressor and its anomalous modulation may contribute to coke-production-

associated carcinogenesis. Coke oven emission-transformed HBE cells demonstrated *TRIM36* hypermethylation and overall gene suppression [69]; this gene-specific methylation was significantly associated with 1-OHP levels and DNA damage, indicating aberrant methylation of *TRIM36* may contribute to coke oven-associated malignancy. Human *in vivo* (peripheral blood lymphocytes) and *in vitro* [human bronchial epithelial (HBE) cells] models highlighted potential crosstalk of several epigenetic endpoints following coke oven exposure [122]. Hypomethylation of *LINE-1* and *O*⁶-methylguanine-DNA methyltransferase (*MGMT*) were significantly correlated with enhanced H3K36me3 modifications in a dose-dependent manner, indicating co-interaction of DNA methylation and histone modifications in the induction of coke production-associated DNA damage.

4.4.2. Histone Modifications.

Trimethylation levels of H3K27me3 and H3K36me3 were elevated in peripheral blood lymphocytes collected from coke oven workers [123]. Notably, a positive correlation was identified between H3K36me3 expression and the level of internal PAH-exposure in all subjects (exposed and control), indicating PAH-induced DNA damage may be mediated by H3K36me3 modifications and could be an indicator of PAH exposure. Additionally, PAH exposure was found to lower global H3K79me2 presence, a histone mark that plays a critical role in DNA damage regulation [124].

4.4.3. Non-Coding RNAs.

Studies previously reviewed elsewhere [10] that investigated the effects of coke oven emission exposure on non-coding RNA expression used human *in vivo* models.

These reports focused on miRNA expression-mediation of coke production-associated oxidative DNA damage and micronuclei frequency. Since 2016, studies have continued to use human *in vivo* models for both miRNA and lncRNA investigations. For example, several plasma miRNAs with significant dysregulation were inversely associated with heart rate variability, representing potential biomarkers for coke production-associated cardiovascular insults [125]. Peripheral blood lymphocytes of coke oven workers demonstrated upregulation of HOTAIR and MALAT1 lncRNA expression; these epigenetic modifications were correlated to DNA damage in an exposure level-dependent manner [126]. Furthermore, elevated HOTAIR expression was significantly associated with H3K27me3 modifications (a biomarker of DNA damage), indicating lncRNA expression may regulate DNA damage through interaction with histone modifications.

4.5. Sulfur mustard

4.5.1. DNA Methylation.

Studies previously reviewed elsewhere [10] that investigated the effects of coke oven emission exposure on non-coding RNA expression used human *in vivo* models. These reports focused on miRNA expression-mediation of coke production-associated oxidative DNA damage and micronuclei frequency. Since 2016, studies have continued to use human *in vivo* models for both miRNA and lncRNA investigations. For example, several plasma miRNAs with significant dysregulation were inversely associated with heart rate variability, representing potential biomarkers for coke production-associated cardiovascular insults [125]. Peripheral blood lymphocytes of coke oven workers

demonstrated upregulation of HOTAIR and MALAT1 lncRNA expression; these epigenetic modifications were correlated to DNA damage in an exposure level-dependent manner [126]. Furthermore, elevated HOTAIR expression was significantly associated with H3K27me3 modifications (a biomarker of DNA damage), indicating lncRNA expression may regulate DNA damage through interaction with histone modifications.

4.5.2. Histone Modifications.

Human early endothelial cells exposed to low sulfur mustard concentrations [127] were tested for histone acetylation (H3-K9, H3-K27, H4-K8) or di-methylation (H3-K9, H3-K27, H3-K36) at three passage timepoints. No clear generational variation or dose-dependent effects could be discerned within the cells, indicating post-translational histone alterations do not play a key role in sulfur-mustard-induced carcinogenesis.

4.5.3. Non-Coding RNAs.

Previously summarized reports [10] predominately investigated the modulation of miRNA expression following sulfur mustard exposure within human *in vivo* models; additional studies occurred in human and murine cells. The most common endpoints reported within these studies include diagnostic methods and therapeutic intervention discoveries for sulfur mustard exposure, mechanisms of sulfur mustard-associated cellular function impairment, and identification of miRNA involved in exposure-induced pathogenesis. Since 2016, modulation of miRNA expression following sulfur mustard exposure has continued to be conducted within human *in vivo* and *in vitro* and rodent *in*

vivo models. These studies report proposed miRNA-mediated pathways of carcinogenesis and potential biomarkers for sulfur mustard exposure. For example, comparative evaluation of miRNA-143 and -9 (miRNA associated with key regulatory pathways such as NF-kB signaling) within urine samples collected from occupationally exposed patients showed significantly decreased expression levels [128]. An additional study reported four significantly upregulated miRNA within serum samples collected from sulfur mustard-exposed veterans [129]. These studies identify specific miRNAs that may represent non-invasive biomarkers of sulfur mustard exposure. MiRNA and SMAD7 expression were upregulated within sulfur mustard exposed lung tissue samples, identifying a potential mechanism by which sulfur mustard exposure regulates inflammatory/fibrotic modifications within the lung [130]. Human keratinocyte cells that were made resistant to sulfur mustard (HaCat/SM) displayed significant differences in miRNA expression patterns when compared to controls [131]. The authors postulated that miRNA-125 and miRNA-181 upregulation was the mechanism for transcriptional effects and sulfur-mustard resistance in HaCat/SM cells.

Male Sprague-Dawley rats exposed to sulfur mustard by inhalation and immortalized rat lung-derived airway epithelial cells (CRL-10354) identified miRNA-140 expression as a mediator of sulfur mustard effects [132]. Additionally, it was proposed levels of miRNA-140 can be used as a predictor of the severity of sulfur mustard-induced health insults.

4.6. Formaldehyde

4.6.1. DNA Methylation.

A study previously reviewed within Chappell et al. [10] showed long-term, low-dose formaldehyde treatment within human bronchial epidermal cells (16HBE) induced global DNA hypomethylation. Since 2016, one publication concerning global methylome modifications following occupational formaldehyde exposure has been published. Beauty salon workers exposed to low levels (up to 0.24 ppm) of formaldehyde demonstrated DNA hypermethylation in whole blood samples [133].

4.6.2. Histone Modifications.

Studies published before 2016 [10] investigating histone modifications following formaldehyde exposure were conducted within human cell lines and histone 4 isolated from calf thymus. The publications focused on the phosphorylation of H3 through JNK and PI3K/Akt pathways (responsible for cellular process regulation) and the inhibition of post-translational modifications by formaldehyde-induced Schiff bases. Since 2016, one publication investigating the epigenetic effects of formaldehyde exposure within human cells has been published. Human bronchial epithelial cells (BEAS-2B) demonstrated the inhibition of covalent modifications of histones H3 and H4 following formaldehyde treatment [134]. This inhibition resulted in aberrant chromatin assembly, altering the expression of multiple cancer-related genes (e.g. JUN, JUNB, CDKN1A, and SERPINB5).

4.6.3. Non-Coding RNAs.

Previously summarized studies [10] concerning the dysregulation of non-coding RNA expression due to formaldehyde exposure utilized human *in vitro* and animal *in vivo* models (rodent and alternative species). These publications focused on the identification of potential miRNAs biomarkers for formaldehyde exposure and the role of inflammatory response pathways in formaldehyde-associated carcinogenesis. No studies concerning the effect of formaldehyde on non-coding RNAs were published between 2016 and 2020.

4.7. 1,3-Butadiene

4.7.1. DNA Methylation.

Previously summarized studies in Chappell et al. [10] reported modification of the methylome following 1,3-butadiene exposure utilized mouse *in vivo* models; these reports focused on strain- and tissue- specificity of DNA methylation levels. Since 2016, mouse *in vivo* studies have continued to be the model of choice; scientific interest has focused on variation within damage responses (e.g. sex-specificity) as well as the influence of genetic variation on epigenomic modifications. For example, sex-specific variation in cytosine DNA hypomethylation patterns within the target (liver and lung) and non-target (kidney) tissues was identified in a mouse population-wide model [135]. Furthermore, previous findings of strain-specific effects on DNA methylation were corroborated with respect to the levels of methylation of LINE-1 and short interspersed nuclear element-B1 (SINE B1) retrotransposons detected following an *in vivo* exposure within a murine model [136]. The liver showed a significant correlation between 1,3-

butadiene-induced DNA damage and DNA methylation at the population level; a negative correlation between DNA adduct levels and the extent of SINE B1 methylation was reported. Inter-strain variability in 1,3-butadiene-associated global DNA methylation, as measured by methylation of LINE-1, was far greater than that for DNA adducts or histone methylation.

4.7.2. Histone Modifications.

As stated above, all prior publications concerning histone modifications following 1,3-butadiene exposure were reported within mouse *in vivo* models. Since 2016, all new research has been collected via mouse *in vivo* inhalation studies to highlight how population variability may influence susceptibility to epigenetic modifications following 1,3-butadiene exposure. For example, inter-strain variability of global histone modifications following 1,3-butadiene inhalation exposure was demonstrated within 20 Collaborative Cross strains [136]. In the lung, trimethylation of H3K9 and acetylation of H3K27 were significantly decreased after exposure in all 20 strains. Conversely, trimethylation of H4K20 in the liver and H3K6 in the kidney showed significant increases. Interestingly, butadiene-associated effects on variability in chromatin accessibility across tissues in exposed mouse strains only partially explained the observed variability in gene transcription effects, indicating that effects on histones and chromatin are not exclusive regulators of the gene expression effects of butadiene [137].

4.7.3. Non-Coding RNAs.

In a study of two inbred mouse strains (C57BL/6J and CAST/EiJ), exposed to 1,3-butadiene via inhalation, strain-specific differences in miRNA expression in response to

treatment were observed [138]. No effects were observed in CAST/EiJ mice; however, in C57BL/6J mice, exposure resulted in changes in the expression of 109 miRNAs. Notably, miR-326-2p, a key mitigator of cell proliferation and migration within lung carcinogenesis, and miR-150-5p, a regulatory miRNA for glucuronosyltransferases *Ugt1a1*, *Ugt1a2*, and *Ugt1a5*, were affected. These results suggest that butadiene-associated altered miRNA expression change are strain-specific.

4.8. Vinyl Chloride

4.8.1. DNA Methylation.

As shown within Chappell et al [10], publications concerning epigenetic modifications induced by vinyl chloride exposure exclusively reported DNA methylation within human *in vivo* models. Specifically, these studies focused on the role genetic mutations (e.g. p53 abnormalities) play within vinyl chloride-associated carcinogenesis and the specific methylation of DNA repair genes (e.g. MGMT). Since 2016, one additional report concerning vinyl chloride exposure causing alterations to the methylome within rodents has been published. Hepatocytes collected from rats intraperitoneally injected with vinyl chloride showed a positive correlation between DNA damage and promoter methylation of RASSF1A (gene associated with cell cycle regulation) and MGMT [139].

4.8.2. Histone Modifications.

No studies of histone modifications were identified.

4.8.3. Non-Coding RNAs.

A recent study within human peripheral blood lymphocytes collected from vinyl chloride-exposed workers possessed four significantly dysregulated miRNAs (MiRNA - 222-3p, MiRNA -146a-5p, MiRNA -151a-5p, MiRNA -22-3p). MiRNA-22-3p upregulation was positively correlated with high micronuclei frequency, a sign of genotoxicity [140]. These sites of modified expression could act as potential biomarkers of vinyl chloride monomer exposure and resulting carcinogenesis.

4.9. Occupational exposure as a painter

4.9.1. DNA Methylation.

No studies of DNA methylation were identified.

4.9.2. Histone Modifications.

No studies of histone modifications were identified.

4.9.3. Non-Coding RNAs.

A recent study of four professional painters within the naval industry aimed to better understand the role of blood miRNAs as biomarkers of exposure to common volatile organic solvents (VOCs) (ethylbenzene, toluene, xylene) used within shipyard painting [141]. Significant upregulation of miRNA-6819-5p and miRNA-6778-5p was noted within exposed workers when compared to controls. Evaluated urinary metabolites of exposed workers showed significant associations between miRNAs and organic solvents' absorbed dose. The authors concluded a more robust study could provide key insight into specific miRNAs that may be used as biomarkers of exposure to match with specific VOCs at defined concentrations.

4.10. Coal tar pitch

4.10.1. Routes of exposure, associated cancers, and genotoxicity.

Coal tar pitch is predominately an occupational exposure as it is used in electrode manufacturing and construction (paving and roofing). Limited data is available for coal tar pitch but an association between exposure and cancer of the bladder has been noted within human studies. Genotoxicity investigations identified sister chromatid exchanges and DNA strain breaks induced by coal tar pitch exposure. Further exposure information and mechanistic data are available at: <https://monographs.iarc.who.int/wp-content/uploads/2018/06/mono100F-17.pdf>.

4.10.2. DNA Methylation.

Coal-tar pitch extract (CTPE) induced down-regulation of genomic DNA methylation in BEAS-2B cells (lung epithelial cells) [142]. CTPE-induced BEAS-2B cells in culture possessed upregulated DNMT1, DNMT3a, DNMT3b, HDAC1, let-7a, and miR-21 levels within early passages, representing potential biomarkers for coal tar pitch-induced carcinogenesis.

4.10.3. Histone Modifications.

No studies of histone modifications were identified.

4.10.4. Non-Coding RNAs.

Coal tar pitch extract (CTPE)-induced BEAS-2B cells, when compared to controls, showed 707 differentially expressed lncRNA [143]. Specifically, lncRNA ENST00000501520 expression increased in malignant-transformed cells exposed to coal

tar pitch [143, 144]. When ENST00000501520 was knocked down, cell arrested in G1 phase and increased apoptosis was observed.

4.11. 4-Aminobiphenyl

4.11.1. DNA Methylation.

No studies of DNA methylation were identified.

4.11.2. Histone Modifications.

No studies of histone modifications were identified.

4.11.3. Non-Coding RNAs.

As summarized elsewhere [10], previous studies reporting epigenetic alterations due to 4-aminobiphenyl exposure utilized human cells. The reports focused on histone modifications and the role of miRNAs in DNA damage response following exposure. Since 2016, one additional study has been published further investigating the role of miRNA in 4-aminobiphenyl genotoxicity. Human liver carcinoma cells (HepG2) exposed to 4-aminobiphenyl demonstrated increased reactive oxidative species (ROS) generation, leading to the increased expression of cAMP-response element-binding protein (CREB; a transcription factor) [145]. The upregulation of CREB increased the expression of miRNA-630 which modulates RAD18 and MCM8 (genes associated with homologous recombination repair pathway) expression. These findings may increase understanding of molecular mechanisms concerning the role of miRNA-630 in reducing DNA damage repair within 4-aminobiphenyl-treated hepatic cells

5. EPIGENETIC EFFECTS ASSOCIATED WITH IARC GROUP 1 HUMAN CARCINOGENS AND ASSOCIATED OCCUPATIONS INCLUDED IN MONOGRAPHS 105-118

Eight additional human genotoxic carcinogens that were evaluated by IARC after Monograph Volume 100 also met inclusion criteria for this systematic literature review. A description of the scientific evidence concerning DNA methylation (global, gene-specific, and DNMT associated), histone modifications (global, gene-specific, histone enzyme associated, or chromatin structure), and non-coding RNAs (miRNA and lncRNA targeted and untargeted testing) for each carcinogen is summarized herein. Additionally, a brief description of the common routes of exposure, associated cancers, and mechanism of genotoxicity is provided. Further background information for each carcinogen is available in the corresponding IARC monograph (**Table 1**).

5.1. Outdoor Air Pollution

5.1.1. Routes of exposure, associated cancers, and genotoxicity.

Outdoor air pollution is a multi-source, complex exposure of one or more pollutants that occurs continuously. As summarized within IARC Monograph 109, the main exposure source categories include vehicle emissions; stationary power generation; agricultural and manufacturing emissions; residential sources; terrestrial and aquatic re-emissions; chemical production, distribution, and use; and natural processes. Outdoor air pollution includes photochemical oxidants, particulate matter, sulfur dioxide, carbon monoxide,

nitrogen oxide, toxic and/or volatile metals, polycyclic aromatic hydrocarbons (PAHs), organic compounds (VOCs, SVOCs, and particulate organic matter), mineral dust/fibers, and bioaerosols. Target organs for carcinogenesis following outdoor air pollution exposure include the lung, bladder, and breast; hematological malignancies have also been associated with these exposures. Human *in vitro* and *in vivo* data significantly indicates outdoor air pollution induces pulmonary cancers through genotoxic mechanisms and promotes tumorigenesis through oxidative stress, responses to oxidative stress, and persistent inflammation. Further exposure and other information are detailed elsewhere [146].

5.1.2. Epigenetic Effects.

Scientific evidence of outdoor air pollution-associated epigenetic modifications is voluminous. A total of 163 studies were included in this review; of these, 140 publications reported effects *in vivo*. DNA methylation is the most studied epigenetic effect of outdoor air pollution (64%), followed by non-coding RNAs (34%) and histone modifications (7%). A total of 135 publications (83% of all included) reported data from studies in humans (112 *in vivo* and 24 *in vitro* studies). As most outdoor air pollution exposure publications concern human *in vivo* investigations into epigenetic modifications, the most recently published studies per endpoint (DNA Methylation, Histone Modifications, and Non-Coding RNAs) have been identified below to provide insight into current research areas. All other included studies can be reviewed within the HAWC assessment.

5.1.2.1. DNA Methylation.

Recent studies have focused on mitochondrial DNA methylation [147, 148], identification of mechanistic pathways involved in outdoor air pollution-induced toxicity [149, 150], and *in utero* or childhood exposures [151-153]. Additional studies have investigated gene-specific methylation, specifically “clock genes” associated with the circadian rhythm [154], and methylation of genes encoding inflammatory cytokines [155].

5.1.2.2. Histone Modifications.

A recent report showed air pollution exposure impairs brain chromatin silencing by decreasing histone posttranslational modifications, reducing DNA integrity overall [156]. Additionally, early-life ambient air pollution exposure was associated with cord plasma histone H3 modifications [157].

5.1.2.3. Non-Coding RNAs.

No human *in vivo* studies of epigenetic effects of outdoor air pollution investigated long non-coding RNAs; all non-coding RNA publications focus on miRNA. The latest miRNA research has centered on the identification of miRNA biomarkers for air pollution health risk assessment [158], prenatal exposure inducing inflammatory responses and carcinogenesis [159, 160], and miRNA-mediated mechanisms of air pollution biological insults [161].

5.2. Engine exhaust, diesel

5.2.1. Routes of exposure, associated cancers, and genotoxicity.

Diesel and gasoline represent major energy sources used to power motor vehicles. Most human exposures occur in occupational (e.g. mining, construction, manufacturing) and traffic settings. Significant positive correlations exist between diesel engine emission exposure and the occurrence of lung and/or bladder cancer within humans. Engine-exhaust-associated carcinogenesis is induced through genotoxic mechanisms including DNA damage, mutations and chromosomal-level events, gene expression modulation, and production of reactive oxygen species (increased inflammatory response). Lung carcinogenicity following diesel engine emission exposure is suspected to be amplified by the other known and suspected human carcinogens within diesel engine exhaust. Further exposure and other information are detailed elsewhere [162].

5.2.2. DNA Methylation.

Workers chronically exposed to diesel engine exhaust exhibited significant hypomethylation of *p16*, *RASSF1A*, and *MGMT* (DNA damage response-related genes) and increased cytokinesis-block micronucleus (CBMN) cytome index (a biomarker of carcinogenesis) [163]. A second human *in vivo* study, evaluating peripheral blood mononuclear cell DNA methylation levels, demonstrated significant global hypomethylation of CpG sites involved in inflammation responses, repetitive elements, and miRNA [164]. Human umbilical vein endothelial cells (HUVECs) exhibited increased reactive oxygen species generation and DNA damage following diesel exhaust particulate matter exposure [165]. Furthermore, aberrant gene methylation was reported.

Several human *in vivo* and *in vitro* studies have analyzed diesel exhaust and allergen co-exposures. For example, human bronchial epithelial cells exposed to diesel

exhaust particles and house dust mite allergens resulted in increased TET1 and DNMT1 expression, indicating novel 5mC and 5hmC changes in inflammation and epithelial repair response genes [166]. In contrast, human *in vivo* exposure to diesel exhaust particles, allergens, or co-exposure only demonstrated minor DNA methylation changes [167]. Only when allergens and diesel exhaust were experienced in sequential insults were significant CpG site methylation levels noted, with the order of exposure determining unique epigenetic changes. An additional study identified airway hyperresponsive patients with naturally high TET enzyme expression and global hypermethylation as a susceptible population to diesel exhaust-associated exposures [168].

Several rodent studies *in vivo* investigated the epigenetic effects of diesel exhaust exposure, specifically *in utero* exposures. For example, pregnant C57BL/6J mice were exposed to diesel exhaust; their offspring were subsequently necropsied for brain tissue at either day 1 or day 21 [169]. The genome-wide DNA methylation status was disrupted at both time points investigated, suggesting potential future health insults and disease induction due to altered developmental pathways. Additional studies demonstrated *in utero* exposure within murine models caused altered methylation within cardiomyocytes, modifying cardiac transcriptional responses [170] and metabolic capability of the cells [171]. Aberrant CpG island methylation of *p16^{INK4A}* was noted within diesel exhaust-associated tumors, indicating tumorigenesis may involve p16 suppression [172]. Furthermore, the inactivation of *p16* supports a possible role for oxidative stress and inflammation in the initiation of lung cancer. Lung inflammation following diesel

exhaust exposure was further explored in Sprague-Dawley rats [172]. Diesel exhaust particle exposure suppressed Clara cell secretory protein (CC16) levels through aberrant methylation of the CCAAT/enhancer-binding protein alpha (C/EBPa) promoter.

5.2.3. Histone Modifications.

Human bronchial epithelial cells exposed to diesel exhaust particles showed enhanced expression of COX-2, a key mediator of stress-induced inflammatory responses; this effect was attributed to chromatin modifications [173]. Specifically, increased acetylation of H4 occurred through the degradation of HDAC1 (COX-2 transcription regulator).

5.2.4. Non-Coding RNAs.

Several studies utilizing human cells have demonstrated diesel exhaust exposure altered miRNA and gene expression [174-176]. For example, diesel exhaust exposure increased miRNA 21 expression and subsequently activated the PTEN/P13K/AKT pathway, a potential mechanism for diesel exhaust-associated carcinogenesis [177]. A second study demonstrated diesel exhaust exposure repressed miRNA 382 expression, increasing CXCL12/MMP9 and triggering pulmonary inflammation [178].

Human *in vivo* studies investigating diesel exposure have shown dysregulated miRNA and associated genes induce bronchial immune responses [179] and oxidative stress [180]. Additionally, peripheral blood mononuclear cells collected from diesel exhaust-exposed finishing workers showed significant DNA adduct formation and miRNA dysregulation [181]; aberrantly expressed miRNAs were associated with carcinogenesis apoptosis and antioxidant effects.

5.3. Lindane

5.3.1. Routes of exposure, associated cancers, and genotoxicity.

The International Organization for Standardization (ISO) common name of “lindane” refers to any material containing > 99% γ -hexachlorocyclohexane (γ -HCH), a stable isomer of hexachlorocyclohexane. Lindane is primarily used as an insecticide within forestry and agricultural settings but is used in some niche pharmaceutical scenarios to treat head lice and scabies (mites). Adverse effects from lindane exposure have been noted within the liver, kidney, and hematopoietic systems along with testicular toxicity. Lindane is suggested to be immunosuppressive by increasing oxidative stress and genotoxic by inducing chromosomal and micronuclei frequency alterations. Information on exposure is detailed elsewhere [182].

5.3.2. Epigenetic Effects.

One study reported epigenetic effects following lindane exposure, analyzing both DNA methylation and histone modifications. Pregnant female albino Wistar rats were exposed to lindane (0.25 mg/kg) and corn oil [183]. The resulting male offspring were further divided into three subgroups based on age (9, 18, or 27 weeks). These three subgroups were separated into subsequent treatment cohorts: lindane (5 mg/kg by weight, orally, 5 consecutive days) or corn oil (5 mg/kg by weight, orally, 5 consecutive days). Results from this study revealed upregulation in histone H3 acetylation and CpG methylation of promoter regions of cerebral cytochrome P450s (CYPs). The authors concluded the specific CpG alterations following lindane exposure may suggest the

occurrence of imprinting of cerebral CYPs, which may account for enhanced responsiveness of these indicated CYPs. More robust rat model studies were recommended to validate these hypotheses.

5.4. Pentachlorophenol

5.4.1. Routes of exposure, associated cancers, and genotoxicity.

Pentachlorophenol is produced through (i.) stepwise chlorination of phenols in the presence of ferric chloride or anhydrous aluminum chloride (catalysts) or (ii.) alkaline hydrolysis of hexachlorobenzene. While the initial production of the chemical was for wood preservation, the compound is now used as an herbicide, fungicide, germicide, and algicide; in tannery; in production of paints and adhesives; and in general construction such as insulation and brickwork. Due to its primary uses, human exposure commonly occurs in occupational settings and has been linked to non-Hodgkin lymphoma, soft tissue sarcoma, and acute lymphocytic leukemia. Human *in vitro* data has indicated pentachlorophenol induces oxidative stress and genotoxicity; non-mammalian models report increased oxidative stress markers and DNA adduct levels. Information on exposure is detailed elsewhere [146].

5.4.2. Epigenetic Effects.

One study reported epigenetic effects following pentachlorophenol, specifically modifications of non-coding RNAs. Pregnant ICR mice were exposed to pentachlorophenol (0.02, 0.2, or 2 mg/kg by weight) during gestation day (GD) 0.5 to GD 8.5 and then compared to controls [184]. Autophagy was increased within exposed

mouse placentas as indicated by upregulated mRNA expression of P62, LC3-II/LC3-I, and Beclin1. To further investigate the increased autophagy found in pentachlorophenol-exposed placenta samples, the expression of miR-30a-5p (an expression inhibitor of Beclin1) was evaluated. Increased pentachlorophenol exposure was significantly correlated with the downregulation of miR-30a-5p expression and induction of autophagy through Beclin1 upregulation. These results characterize the developmental toxicity of pentachlorophenol, completed through autophagy.

5.5. Polychlorinated bi-phenyls

References for polychlorinated biphenyls (PCB) have been separated into the nondioxin-like ortho-substituted PCB category, in which their two rings are not aligned in the same plane, and the more toxic dioxin-like category, with meta and para-substituents whose co-planar aromatic rings are aligned like those of the highly toxic polychlorinated dibenzodioxins (PCBDs). Dioxin-like PCB include PCBs 77, 81, 105, 114, 118, 123, 126, 156, 157, 167, 169, and 189, as listed within IARC Monograph 107 [185].

5.5.1. Routes of exposure, associated cancers, and genotoxicity.

PCBs are ubiquitous environmental contaminants found in all elements (earth, water, air), wildlife, and in most humans. The carcinogens are produced both naturally and during industrial processes. Exposures may occur via ingestion, inhalation, and dermal absorption. Occupational sources include fires, recycling processes, and waste incineration. Exposure to PCB in human populations has been significantly associated

with malignant melanoma, non-Hodgkin lymphoma, and cancer within the breast tissue. Multiple modes of actions have been identified for PCB and their metabolites, causing both genotoxic and nongenotoxic effects associated with tumor initiation and progression as well as cancer induction. Less chlorinated congeners induce oxidative stress and genotoxicity following oxidative metabolism. Highly chlorinated congeners interact with receptors associated with xenobiotic and androgen metabolism. PCB also affect plasma membrane-associated proteins, causing dysregulation of cell processes, and promotion of tumorigenesis. Information on exposure is detailed elsewhere [185].

5.5.2. Non-Dioxin Like PCBs.

5.5.2.1. DNA Methylation.

A large number of studies (39 publications) were identified for non-dioxin-like PCB effects on DNA methylation; therefore, only human *in vivo* data are summarized below. Studies investigating DNA methylation following PCB exposure in human cells and rodent/alternative species models can be reviewed within the HAWC assessment:

<https://hawcproject.org/assessment/1063/>.

Several studies have reported an inverse relationship between global DNA methylation and PCB blood plasma levels collected from human subjects [186, 187]. A cross-sectional analysis identified global DNA hypermethylation within elderly individuals following PCB exposure, potentially identifying a susceptible population [188]. A study of genome-wide methylation following exposure to non-dioxin-like PCB

reported a positive correlation between increased aberrant methylation and intake of fish that likely served as a source of non-dioxin-like PCB [189].

Other studies have implicated potential sex-specific effects of epigenomic responses induced by PCB exposure. For example, global DNA hypomethylation was denoted within an Alu assay for men and DNA hypermethylation within a LINE-1 assay for women [190]. Additionally, significant enrichment was noted within specific X-chromosome sites within prenatally PCB-exposed males [191]. Global hypermethylation was noted following PCB exposure within sperm DNA [192].

Several reports have investigated maternal PCB exposure and resulting prenatal toxicity [193-195]. Aberrant methylation of several genes was detected in blood of newborns perinatally exposed to high concentrations of PCB [196]; this study is the first to indicate PCB exposure persistence in an *in utero* model and may provide further insight into the underlying epigenetic mechanisms. One study, utilizing placental samples collected from PCB-exposed women, highlighted a positive association between total PCB levels and *H19* expression [197].

Differentially methylated genes following PCB exposure in humans may provide further mechanistic evidence to better understand resulting carcinogenesis [198, 199]. Additionally, the methylation of specific genes may offer epigenetic biomarkers for early disease or adverse health effect detection [200, 201]. For example, hypermethylation of *MGMT* displayed an inverted U-shaped association following PCB exposure, indicating a potential non-monotonic dose-response [202].

5.5.2.2. Histone Modifications.

Prolonged exposure to Aroclor 1254 within human neuroblastoma cells (SH-Sy5Y) demonstrated dose-dependent cytotoxicity; cell death was proposed to occur via repressor element 1-silencing transcription factor (REST) decreasing synapsin1 and H3 and H4 acetylation [203]. Early-life PCB exposure within a rat model decreased H4K16ac and H3K4me3 levels via the induction of *SirtT1* and *Jarid1b* (chromatin-modifying enzymes) and downregulation of androgen receptors [204].

5.5.2.3. Non-Coding RNAs.

PCB exposure was shown to affect the implantation function of human endometrial cells by impairing expression of miRNAs that are involved in the epithelial-mesenchymal transition processes [205]. The authors hypothesized that miRNAs may play a role in PCB-associated endometrial toxicity. As novel prenatal biomarkers, placental miRNA profiles were suggested as predictors of *in utero* PCB exposure [206], providing insight into the role of the placental epigenome in human reproduction and development.

Additional studies identified miRNAs and target genes with modified expression following PCB exposure were associated with various types of cancer [207] and provide key insights into mechanisms of PCB toxicity [208] within humans. Rodent *in vitro* and *in vivo* models implicated dysregulated miRNA levels following PCB exposure in the induction of several biological outcomes, including neurodevelopmental disorders [209, 210], atherosclerosis [211], and cardiac development [212].

5.5.3. Dioxin Like.

5.5.3.1. DNA Methylation.

Global methylation levels were significantly decreased following dioxin-like PCB exposure within several cross-sectional human *in vivo* studies, demonstrating that exposure to carcinogenic environmental pollutants induces change in the methylome [187, 213]. Further investigation into the epigenetic modifications associated with exposures to dioxin-like PCBs in human *in vivo* models showed significant changes in CpG methylation [214]. In contrast to human *in vivo* results, human carcinoma cell lines (HepG2 and HeLa) treated with PCB 126 showed no apparent differences within methylation levels of specific CpG locations following exposure [215]. The study further investigated the expression of *CYP1A1* following PCB 126 treatment. It was shown that cell-specific variation in treatment-induced chromatin accessibility of *CYP1A1* promoter may be an important epigenetic regulatory pathway for cell-specific *CYP1A1* expression. PCB 126 exposure within a zebrafish model showed significantly altered global methylation, with most modifications occurring within genes associated with development, metabolism, and regeneration pathways [216].

PCB118 (2,3',4,4',5-pentachlorobiphenyl) is the most individually studied dioxin-like polychlorinated biphenyl due to its high toxicity rate and rapid bioaccumulation capability. Three *in vivo* studies utilizing pregnant mice orally dosed with PCB118 identified altered methylation rates of imprinted genes and modified expression of DNMT1, DNMT3a, DNMT3l, Uhrf1, and Tet3 following exposure [217]. Specifically, methylation patterns within genes H19 and Gtl2 of male offspring from the exposed

dams were altered, identifying sex-variation in response to PCB118 exposure. Within the exposed dams, PCB118 dosage caused dysregulated oocyte maturation [217], defective uterine morphology, and decreased implantation rates [218].

5.5.3.2. Histone Modifications.

Human vascular endothelial cells treated with PCBs 77 or 126 showed increased expression of jumonji domain-contain protein 2b (JMJD2b). These affects were correlated with decreased H3K9me3 repression marks and significantly increased H3K4me3 activation marks in the promoter of this gene; these histone modifications were also associated with increased expression of inflammatory genes following dioxin-like PCB treatment [219]. Human endothelial cells exposed to relevant levels of PCB 126 and/or epigallocatechin-3-gallate (EGCG-an anti-inflammatory polyphenol) were evaluated for epigenetic modifications. Co-exposure of PCB 126 and EGCC decreased PCB-induced NF-kB activation through chromatin binding and histone H3 hypo-acetylation [220]. Mouse Hepatoma-1c1c7 cells treated with various dioxin-like PCBs (77, 169, and 153) demonstrated PCB-induced histone modifications depend upon the activation of AHR by ligand before chromatin modifications critical for active transcription can occur [221].

5.5.3.3. Non-Coding RNAs.

Human *in vivo* studies have shown PCB exposure induces significantly modulated miRNA and target gene expression, with the specified miRNA locations may serve as potential biomarkers for diagnosis of subsequent biological effects [207, 222]. Human liver cells (L-02) exposed to PCB 156, a compound associated with non-

alcoholic fatty liver disease, was found to modulate the expression of mRNAs and lncRNAs associated with metabolic and inflammatory processes [223]. These identified effects may be used as potential biomarkers for early detection of the PCB-induced lipid metabolism disorder.

5.6. Trichloroethylene

5.6.1. Routes of exposure, associated cancers, and genotoxicity.

Trichloroethylene, a solvent used in medical, industrial, and consumer markets, is widely distributed in the environment (soil, air, water). The main source of exposure is during degreasing processes (an occupational exposure). Additional information on exposure is detailed elsewhere [224].

5.6.2. DNA Methylation.

The vast majority of publications concerning modifications to the methylome following trichloroethylene treatment utilized mouse *in vivo* models. Many of these studies focused on the trichloroethylene exposure-associated CD4⁺ T cell-mediated autoimmunity. For example, MRL mice were exposed to trichloroethylene via drinking water until adulthood; continuous exposure throughout development showed significant differentially methylated binding sites of PcG proteins, potentially representing a mechanism by which trichloroethylene alters the function of effector/memory CD4⁺ cells and induced immunotoxicity [225-227]. Further investigation into this mechanism demonstrated hypermethylation of CpG sites within the *IFN-γ* promoter of effector/memory CD4⁺ T cells, representing especially sensitive sites to chronic

trichloroethylene exposure [228]. DNA methylation-induced activation, silencing, and reactivation of proinflammatory genes may represent a mechanism by which autoimmune pathology occurs following trichloroethylene exposure.

Additionally, reports in mouse *in vivo* studies focused on trichloroethylene-induced DNA methylation, and resulting gene expression changes, within the liver. For example, B6C3F1 mice orally dosed with trichloroethylene for five days demonstrated dose-dependent gene expression dysregulation, specifically of *Utrf1*, *Tet2*, *DNMT1*, *DNMT3a*, and *DNMT3b*, potentially leading to tumorigenesis [229]. Trichloroethylene was found to decrease global methylation levels and hypomethylated the promoter regions of protooncogenes, *c-jun*, and *c-myc*; these mechanisms are associated with increased cell proliferation and tumor promotion [230]. Several studies utilized trichloroethylene metabolites, dichloroacetic acid, and trichloroacetic acid. These studies concluded that trichloroethylene metabolite exposure induced DNA hypomethylation [231, 232] and increased expression of *c-jun* and *c-myc* genes [233] within liver tumors. Dichloroacetic acid-associated hypomethylation was prevented by methionine co-administration [234]. Additionally, methionine, an amino acid, may represent a potential treatment following accidental trichloroethylene exposure.

Human *in vitro* studies of the epigenetic modifications induced by trichloroethylene treatment have predominately focused on the identification of biomarkers of exposure and global DNA methylation changes. For example, human lymphoblastoid (TK6) cells demonstrated global DNA methylation as an early event in the genotoxic pathway of trichloroethylene [235]. Additionally, trichloroethylene

exposure induced significant hypomethylation, subsequently inhibiting DNMT activity [236, 237] while enhancing SET levels [238] within L-02 cells. Utilizing single-molecular monitoring, it was uncovered trichloroethylene-associated DNA hypomethylation could be partially explained by disrupted DNMT3A-DNA associations [197]; in a dose-dependent manner, DNMT3A oligomers dissociated from heterochromatin, resulting in significant loss of 5-methylcytosine (5mC) and enhancement of TET enzymes. In total, these results present a mechanistic understanding of trichloroethylene toxicology and may provide the framework for comparable investigation of other environment toxicants.

5.6.3. Histone Modifications.

Multiple studies used L-02 cells to demonstrate histone modifications following trichloroethylene exposure. For example, trichloroethylene exposure increased the histone chaperone SET, enhancing H2AK9ac through HDAC1 inhibition, a mechanism associated with SET-mediated hepatic cytotoxicity [239]. Further investigation into trichloroethylene-induced hepatotoxicity suggested relative expression level changes of H3K79me2 and H3K79me3 were induced through DNA damage [240].

Naturally high levels of SET can induce significant modifications of histone deacetylases, suggesting SET is involved in trichloroethylene-induced cytotoxicity and epigenetic alterations [241]. An in-depth investigation into the mediation of trichloroethylene-induced hepatic cytotoxicity observed SET promoting acetylation of H2AK9 by suppressing HDAC1 levels [239]. This type of investigation could be utilized

to further examine underlying mechanisms in which SET modulates histone acetylation following trichloroethylene exposure.

An additional study in human hepatocytes demonstrated a negative correlation between existing DNA hypomethylation and histone expression of tumor-related genes following trichloroethylene exposure; specifically, trichloroethylene increased histone H3 Lysine 9 acetylation levels [242]. Histone three lysine four dimethylation (H3K4me2), a mark in sperm conserved across species and linked to transgenerational epigenetic inheritance, was significantly decreased in the regulatory protein kinase A signaling pathway genes following trichloroethylene exposure [243]. This epigenetic modification may account for TCE-induced spermatozoal toxicity with H3K4me2 being a potential target for trans-generational effects.

5.6.4. Non-Coding RNAs.

Trichloroethylene was found to modulate miRNA expression within multiple species; the identified locations may represent potential biomarkers for toxicity prediction [244-246]. Occupational exposure to trichloroethylene resulted in decreased serum levels of miRNAs involved in the B-cell receptor pathways and the innate immune response [247]; these results may increase understanding of the etiologic relevance of miRNAs in trichloroethylene-associated Non-Hodgkin's Lymphoma. Human hepatocytes treated with trichloroethylene showed miRNA-199b suppression [248]. Expression of miRNA-182-5p was upregulated following trichloroethylene exposure and was associated with increased cell proliferation [249, 250]. Alternative animal models identified miRNA locations that mediate trichloroethylene-induced

reactive oxygen species generation [246] and dysregulation of cell proliferation [245, 246, 251].

5.7. Welding Fumes

5.7.1. Routes of exposure, associated cancers, and genotoxicity.

Welding fumes are generated when metals reach temperatures above their melting points, vaporize, and then condense into fumes primarily consisting of fine, solid particles (diameter > 1 μm). Welders represent the highest risk population, as they experience contemporaneous exposures of welding fumes, gases, radiation (ionizing and non-ionizing), and co-exposures of solvents and asbestos. Exposure has been positively associated with cancer of the lung and kidney, as well as correlated to other biological effects such as ocular melanoma, leukemia, and non-Hodgkin lymphoma. Evidence of welding fumes-induced genotoxicity included chromosomal aberrations, dysregulated sister-chromatid exchange rates, DNA damage, and anomalous micronuclei frequency. Additional information on exposure is detailed elsewhere [252].

5.7.2. DNA Methylation.

Several studies reported findings from the same study population. Li et al. [253] reported DNA methylation in 101 welders and 127 control workers (all non-smoking males) from southern Sweden [253-255]. Blood sampling showed increased concentration of measurable respirable dust was positively associated with hypomethylation of *F2RL3*, a marker for cardiovascular disease progression and mortality [254]. A second study [255] reported decreased methylation within the

mitochondrial regulatory region D-loop and MT-TF (a tRNA encoding gene) when compared to controls, indicating welding fume exposure may be associated with oxidative stress. Human THP-1 and small human airway epithelial (SAEC) cells and murine (RAW254.7) cells exposed to steel welding fumes showed aberrant methylation of LINE-1, SINE, and Alu elements (transposable elements) and modification of their transcription [256]. Furthermore, the expression of DNA methyltransferases and TET genes were differentially regulated in a cell-, chemical-, and dose-dependent manner. Male Fischer-344 exposed to welding fumes by inhalation showed hypermethylation, increased telomere length, and expression of neurodegeneration markers within brain tissue [257]. By contrast, similarly exposed Sprague-Dawley rats demonstrated no significant differences in DNA methylation patterns between welding fume and control groups, but still reported increased telomere length [258]; these two studies highlight inter-strain variance in the epigenetic effects of welding fume inhalation.

5.7.3. Histone Modifications.

No studies of histone modifications were identified.

5.7.4. Non-Coding RNAs.

A cross-sectional study of 30 male welders was conducted to understand the association between metal particles from welding fumes exposure and circulating miRNA concentrations [259]. The presence of certain metals, specifically chromium and nickel, within blood and urine samples were significantly associated with the downregulation of expression of miRNAs -21, -146a, and -155 (biomarkers of renal function injury).

6. SUMMARY

This systematic literature review updates the 2016 review of the published evidence on epigenetic effects related to Group 1 human carcinogens that are genotoxic and were included in IARC Monograph volume 100F [10]. Since 2015, the number of epigenetic studies on the same chemicals nearly doubled, indicating a very brisk pace of research on epigenetics as a key mechanism in environmental exposure-associated cancer (**Figure 4A**). Benzo[a]pyrene, benzene, and aflatoxins remain the most studied of the genotoxic carcinogens included in IARC Monograph volume 100F, with 49, 40, and 21 publications, respectively, of epigenetic effects published in the last five years. In 2015, no studies of epigenetic effects of coal tar pitch were available, but several recent publications reported such effects. By contrast, no additional research has been published concerning epigenetic modifications following exposure to benzidine or 4,4'-methylenebis-(2-chloroaniline) (MOCA). Of the agents with evidence of genotoxicity classified as Group 1 carcinogens by IARC in Monograph volumes 105-118 (**Figure 4B**), the evidence of epigenetic effects is similarly uneven across agents. Outdoor air pollution is by far the most studied, with respect to epigenetics as a key characteristic of human carcinogens. Over 160 publications reported epigenetic effects of exposure to outdoor air pollution. Notably, most of these newly classified carcinogens had more than 10 published reports of epigenetic alterations.

Of note is also the distribution of the types of epigenetic effects that were the subject of publications. This systematic literature review provides data to better

understand the distribution of the endpoints, models and study types investigated within the included scientific reports (**Figure 5**). Interestingly, the relative proportion of the epigenetic endpoints remains relatively consistent for the comparison of the exposures classified in the Monograph volume 100F. Studies of DNA methylation remain of great interest to the scientific community, and non-coding RNA is an increased focus in the last 5 years. Investigation of histone modifications and chromatin state alterations in toxicology are far fewer in number, as compared to the other two epigenetic modifications. This disparity is likely because of the variety and technical demands for the assays of histones or chromatin [260], as compared to more straightforward sequencing-based assays for DNA methylation and non-coding RNAs. A similar disparity is evident for the agents identified as human carcinogens in Monograph volumes 105-118; almost two-thirds of studies focused on DNA methylation.

It is also noteworthy that most studies of epigenetic effects of human genotoxic carcinogens have been performed in human models and about half or more of those were studies in exposed humans. Studies in rodents represent the second most common type of epigenetic investigations in toxicology. Most of the studies of epigenetics were conducted *in vivo*, both in humans and rodents. Indeed, across all species, one half-to-three quarters of all studies identified in this systematic literature review have been conducted *in vivo*. This observation is important because it shows that the mechanistic evidence for epigenetic effects of environmental exposures is highly relevant and useful to human cancer hazard evaluation [1]. The fraction of studies in humans, and especially studies *in vivo* in humans is increasing in recent years. Even among *in vitro* studies, human cells are

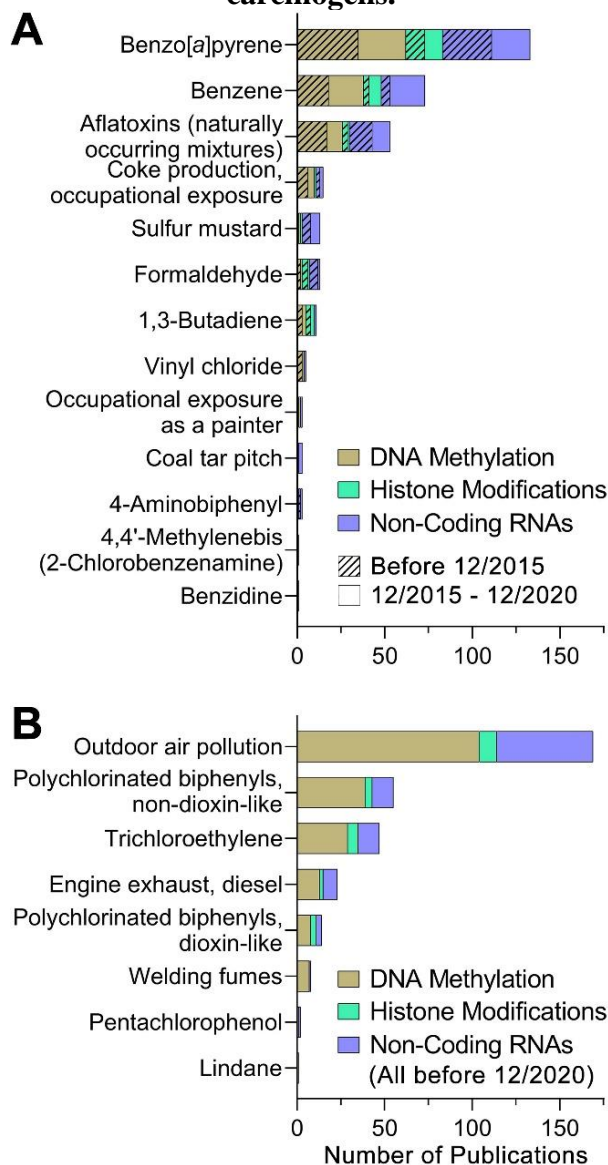
the most common model. The availability of the data in human cells provides an opportunity to fill gaps in knowledge when samples from human exposures are unavailable.

7. FUTURE RESEARCH NEEDS

Overall, this project highlights the considerable progress in epigenetic research and its relevance to elucidating carcinogen mechanisms. Even though the evidence for associations between chemicals and epigenetic effects were not deemed to be “strong” in the recent IARC monographs that utilized the key characteristics approach in the evaluation of the mechanistic evidence [261], the information provided in this review demonstrates that epigenetic effects observed in studies *in vitro* are concordant with those from *in vivo* studies.

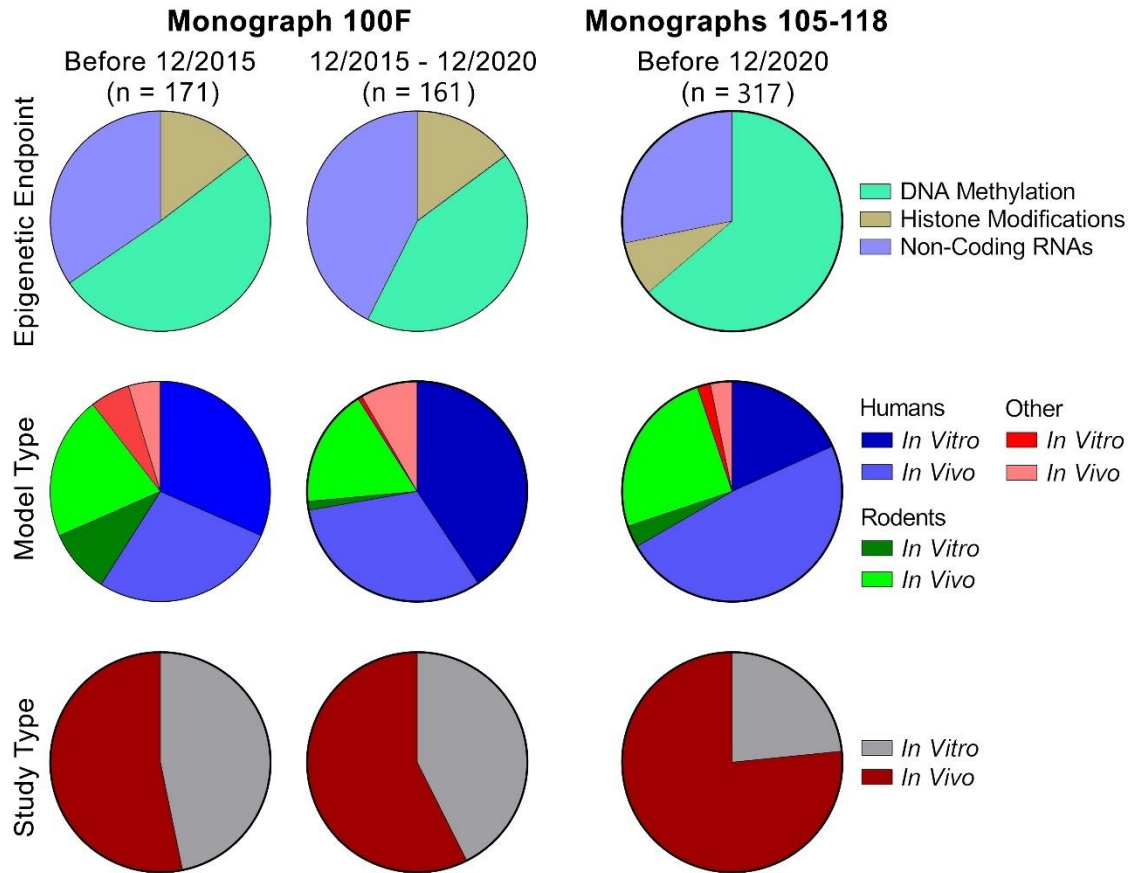
Additionally, several important future research needs were identified. In particular, increased dose- and time-dependent testing in studies of epigenetic endpoints would improve interpretation. Dose-response data, derived from both animal and human studies, may improve risk characterization, and increase understanding of an agent’s mode of action. Further study on the persistence of effects following cessation of exposure is another important data gap, especially considering the dynamic nature of most epigenetic alterations. Finally, exploration of more diverse study populations, using both *in vivo* and *in vitro* models, will facilitate greater understanding of the variability in toxic response in exposed populations. In all, these results lay important groundwork for the application of epigenetic data in hazard and risk assessment exercises.

Figure 7-1. Volume of publications per epigenetic endpoints for all review carcinogens.



The number of publications reporting epigenetic effects of exposures to the human genotoxic substances, related occupations, or types of exposure included in IARC Monographs 100F (**A**) and 105-118 (**B**). Entries are sorted in a descending order. Colors refer to the type of the epigenetic endpoints: DNA methylation, histone modifications, and non-coding RNA are colored in gold, green, and purple, respectively (see color legend inset). In (A), studies published before December 2015 (and were included in Chappell et al [10] systematic literature review) are denoted by hashed segments. Studies published from December 2015 to December 2020 are denoted by solid segments. In (B), all studies published before December 2020 are included.

Figure 7-2. Comparative analysis of study characteristics of included publications.



A relative comparison of epigenetic endpoints, model types (human, rodent, other), and study types (*in vitro* or *in vivo*) for the human genotoxic substances, related occupations, or types of exposure included in IARC Monographs 100F (left) and 105-118 (right). The pie charts for each type of comparison are color coded as shown in the legends.

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APPENDIX A

SUPPLEMENTARY TABLES

Supplementary Table 1. HAWC Project literature search terms.

Search Term Category	Search Terms
<p>Epigenetic Alterations https://hawcproject.org/assessment/1046/ https://hawcproject.org/assessment/1063/</p>	<p>("epigenomics"[MeSH Terms] OR "epigenomics"[All Fields] OR "epigenetic"[All Fields] OR "microrna"[All Fields] OR "micrnas"[All Fields] OR "miRNAs"[All Fields] OR "miRNA"[All Fields] OR "lncRNA"[All Fields] OR "rna, long noncoding"[MeSH Terms] OR "non-coding RNA"[All Fields] OR "non coding RNA"[All Fields] OR "ncRNA"[All Fields] OR "small RNA"[All Fields] OR "smallRNA"[All Fields] OR "dna methylation"[All Fields] OR "promoter methylation"[All Fields] OR "methylated DNA"[All Fields] OR "chromatin modification"[All Fields] OR "open chromatin"[All Fields] OR "histones"[MeSH Terms] OR "histones"[All Fields] OR "histone"[All Fields])</p>
<p>Monograph 100F Carcinogens https://hawcproject.org/assessment/1046/</p>	<p>("benzidine"[All Fields] OR "92-87-5"[All Fields] OR "4-aminobiphenyl"[All Fields] OR "92-67-1"[All Fields] OR "MOCA"[All Fields] OR "4,4'-methylenebis(2-chlorobenzenamine)"[All Fields] OR "4,4'-methylenebis(2-chloroaniline)"[All Fields] OR "101-14-4" OR "2-naphthylamine"[All Fields] OR "91-59-8"[All Fields] OR "ortho-toluidine"[All Fields] OR "95-53-4"[All Fields] OR "benzo[a]pyrene"[All Fields] OR "benzo-a-pyrene"[All Fields] OR "50-32-8"[All Fields] OR "coal-tar"[All Fields] OR "8007-45-2"[All Fields] OR "coal-tar pitch"[All Fields] OR "65996-93-2"[All Fields] OR "coal gasification"[All Fields] OR "coke production"[All Fields] OR "coke oven"[All Fields] OR "aflatoxin"[All Fields] OR "1402-68-2" OR "mineral oil"[All Fields] OR "benzene"[All Fields] OR "71-43-2"[All Fields] OR "1,3-butadiene"[All Fields] OR "106-99-0" OR "ethylene oxide"[All Fields] OR "75-21-8"[All Fields] OR "formaldehyde"[All Fields] OR "50-00-0"[All Fields] OR "sulfur mustard"[All Fields] OR "505-60-2"[All Fields] OR "vinyl chloride"[All Fields] OR "75-01-4"[All Fields] OR "rubber manufacture"[All Fields] OR "rubber manufacturing"[All Fields] OR "chimney sweep"[All Fields] OR "painter"[All Fields] OR "iron founding"[All Fields] OR "steel founding"[All Fields] OR "bis(chloromethyl) ether"[All Fields] OR "542-88-1"[All Fields] OR "chloromethyl methyl ether"[All Fields] OR "107-30-2"[All Fields] OR "isopropyl alcohol manufacture"[All Fields] OR "manufacture of isopropyl alcohol"[All Fields] OR "strong inorganic acid mist"[All Fields] OR "aluminum production"[All Fields] OR "production of aluminum"[All Fields])</p>

Supplementary Table 1 Continued

Search Term Category	Search Terms
<p>Monograph 105-118 Carcinogens https://hawcproject.org/assessment/1063/</p>	<p>(“welding fumes”[All Fields] OR “pentachlorophenol”[All Fields] OR “Chlorophenasic acid”[All Fields] OR “Chlorophen”[All Fields] OR “PCP”[All Fields] OR “penchlorol”[All Fields] OR “penta”[All Fields] OR “pentachlorophenate”[All Fields] OR “2,3,4,5,6-pentachlorophenol”[All Fields] OR “1-hydroxy-2,3,4,5,6-pentachlorobenzene”[All Fields] OR “87-86-5”[All Fields] OR “lindane”[All Fields] OR “γ-Benzene hexachloride”[All Fields] OR “γ-hexachlorocyclohexane”[All Fields] OR “58-89-9”[All Fields] OR “1,2-dichloropropane”[All Fields] OR “propylene dichloride”[All Fields] OR “propylene bichloride”[All Fields] OR “propylene chloride”[All Fields] OR “dichloro-1,2 propane”[All Fields] OR “chloromethylchloride”[All Fields] OR “78-87-5”[All Fields] OR “outdoor air pollution”[All Fields] OR “Polychlorinated biphenyls”[All Fields] OR “3,3',4,4'-Tetrachlorobiphenyl”[All Fields] OR “3,3',4,4'-PCB”[All Fields] OR “32598-13-3”[All Fields] OR “3,4,4',5-PCB”[All Fields] OR “1,1'-Biphenyl, 3,4,4',5-tetrachloro”[All Fields] OR “70362-50-4”[All Fields] OR “2,3,3',4,4'-Pentachlorobiphenyl”[All Fields] OR “1,1'-Biphenyl, 2,3,3',4,4'-pentachloro-“[All Fields] OR “32598-14-4”[All Fields] OR “2,3,4,4',5-PCB”[All Fields] OR “1,1'-Biphenyl, 2,3,4,4',5-pentachloro-“[All Fields] OR “74472-37-0”[All Fields] OR “2,3',4,4',5-PCB”[All Fields] OR “1,1'-Biphenyl, 2,3',4,4',5-pentachloro-“[All Fields] OR “31508-00-6”[All Fields] OR “1,1'-Biphenyl, 2,3',4,4',5'-pentachloro-“[All Fields] OR “2,3',4,4',5'-Pentachlorobiphenyl”[All Fields] OR “65510-44-3”[All Fields] OR “1,1'-Biphenyl, 3,3',4,4',5-pentachloro-“[All Fields] OR “3,3',4,4',5-Pentachlorobiphenyl”[All Fields] OR “57465-28-8”[All Fields] OR “1,1'-Biphenyl, 2,3,3',4,4',5-hexachloro-“[All Fields] OR “2,3,3',4,4',5-Hexachlorobiphenyl”[All Fields] OR “38380-08-4”[All Fields] OR “1,1'-Biphenyl, 2,3,3',4,4',5'-hexachloro-“[All Fields] OR “2,3,3',4,4',5'-Hexachlorobiphenyl”[All Fields] OR “69782-90-7”[All Fields] OR “1,1'-Biphenyl, 2,3',4,4',5,5'-hexachloro-“[All Fields] OR “2,3',4,4',5,5'-Hexachlorobiphenyl”[All Fields] OR “52663-72-6”[All Fields] OR “1,1'-Biphenyl, 3,3',4,4',5,5'-hexachloro-“[All Fields] OR “3,3',4,4',5,5'-Hexachlorobiphenyl”[All Fields] OR “32774-16-6”[All Fields] OR “1,1'-Biphenyl, 2,3,3',4,4',5,5'-heptachloro-“[All Fields] OR “2,3,3',4,4',5,5'-Heptachlorobiphenyl”[All Fields] OR “39635-31-9”[All Fields] OR “chlorinated biphenyls”[All Fields] OR “chlorinated diphenyls”[All Fields] OR “chlorobiphenyls”[All Fields] OR “polychlorobiphenyls”[All Fields] OR “1336-36-3”[All Fields] OR “Engine exhaust”[All Fields] OR “diesel”[All Fields] OR “trichloroethylene”[All Fields] OR “ethinyl trichloride”[All Fields] OR “ethylene trichloride”[All Fields] OR “TCE”[All Fields] OR “1,1,2-trichloroethylene”[All Fields] OR “79-01-6”[All Fields])</p>

Supplementary Table 2. Inclusion and Exclusion criteria.

Inclusion Criteria	Exclusion Criteria
<p>The study investigated a carcinogen that:</p> <ul style="list-style-type: none"> • Was categorized as Group 1 (carcinogenic to humans) • Has one or more demonstrated genotoxic mechanism(s) of carcinogenesis <p>Publication evaluated epigenetic alterations that occurred following independent exposure of a chemical of interest</p> <p>Publication included original data</p> <p>Study was published in English</p>	<p>The study investigated a chemical of interest but did not report any epigenetic alteration testing</p> <p>The study did not investigate a chemical of interest</p> <p>Access to the full scientific report was impeded (e.g. private journal, alternative language)</p> <p>The study was a review or commentary that did not provide primary data</p>